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Impaired neutrophil extracellular trap formation: a novel defect in the innate immune system of aged individuals

Jon Hazeldine, 1 Phillipa Harris, 2 Iain L. Chapple, 2 Melissa Grant, 2 Hannah Greenwood, 3 Amy Livesey, 1 Elizabeth Sapey 2 and Janet M. Lord

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Summary

Neutrophil extracellular traps (NETs) are a recently discovered addition to the defensive armamentarium of neutrophils, assisting in the immune response against rapidly dividing bacteria. Although older adults are more susceptible to such infections, no study has examined whether aging in humans influences NET formation. We report that TNF-α-primed neutrophils generate significantly more NETs than unprimed neutrophils and that lipopolysaccharide (LPS)- and interleukin-8 (IL-8)-induced NET formation exhibits a significant age-related decline. NET formation requires generation of reactive oxygen species (ROS), and this was also reduced in neutrophils from older donors identifying a mechanism for reduced NET formation. Expression of IL-8 receptors (CXCR1 and CXCR2) and the LPS receptor TLR4 was similar on neutrophils from young and old subjects, and neutrophils challenged with phorbol-12-myristate-13-acetate (PMA), as well as microbes (e.g. bacteria, fungi and protozoa) and their products (Cooper et al., 2013), NETs, as their name suggests, are highly efficient at pathogen entrapment. Indeed, in vitro, NETs have been shown to trap bacteria, parasites and fungi (Brinkmann et al., 2004; Urban et al., 2006; Guimaraes-Costa et al., 2009), whilst in vivo, increased bacterial dissemination has been observed in pathogen-infected mice administered exogenous DNases (Meng et al., 2012; Yipp et al., 2012). Interestingly, despite the fact that the granule-derived proteins resident on NETs retain their activity when bound to DNA (Parker et al., 2012), NETs are themselves not directly microbicidal (Menegazzi et al., 2012; Parker et al., 2012). Instead, these structures are thought to confer protection by neutralizing the virulence factors of invading microbes (Brinkmann et al., 2004) and via their immunomodulatory effects, which include activating the human complement system (Oehmcke et al., 2009) and enhancing the activity of bystander T cells (Tillack et al., 2012).

Three key molecular processes underlie the formation of NETs, namely reactive oxygen species (ROS) generation, autophagy and the hypercitrullination of histones H3 and H4, a process catalysed by the calcium-dependent nuclear enzyme peptidylarginine deiminase 4 (PAD4) (Remijien et al., 2011). To date, attention has focused predominantly on the generation of ROS, which play an essential role in NET formation. Pretreating neutrophils from healthy subjects with inhibitors of the ROS-generating enzyme nicotinamide adenine dinucleotide phosphate-oxidase (NADPH oxidase) or the antioxidant protein taurine prevents NET formation to a wide array of stimuli (Fuchs et al., 2007). Similarly, when challenged with pathogens or PMA, neutrophils isolated from patients with chronic granulomatous disease (CGD), an inherited disorder in which subunits of NADPH oxidase are either absent or dysfunctional (Seger, 2008), fail to generate NETs (Fuchs et al., 2007; Bianchi et al., 2009; Nishinaka et al., 2011). However, NETs are produced when these cells are challenged with exogenous hydrogen peroxide (H2O2), further highlighting the importance of ROS in NET generation. H2O2, however, does not appear to be the active oxygen species that triggers NET generation, with this function attributed instead to its downstream products hypochlorous acid (HOCl) and singlet oxygen (Nishinaka et al., 2011; Palmer et al., 2012).

Compared with younger subjects, older adults report an increased incidence of bacterial and fungal infection (Kauffman, 2001; Laupland et al., 2003). As neutrophils represent the frontline defence against these pathogens, this increased susceptibility to infection implies that the protective nature of neutrophils wanes with age. Indeed, as a result of impairments in several defensive strategies such as phagocytosis (Wenisch et al., 2000; Butler et al., 2001), degranulation (McLaughlin et al., 1986) and ROS production (Fulop et al., 2004), the microbicidal activity of neutrophils from older adults is markedly reduced (Wenisch et al., 2000; Simell et al., 2011). Recently, Tseng and colleagues (Tseng et al., 2012) made the novel observation that NET formation is also impaired with age, with the group demonstrating neutrophils from aged...
mice generated significantly fewer NETs when challenged with *Staphylococcus aureus* (Tseng et al., 2012). *In vivo*, this defect in NET generation was associated with marked bacteraemia, leading the group to hypothesize that aberrant NET formation may explain why older adults are more susceptible to invasive bacterial disease following skin and soft tissue infection (Tseng et al., 2012). However, the study did not investigate the impact of physiological aging on NET generation by human neutrophils or the mechanisms involved.

To address this important issue, we isolated neutrophils from healthy young and old donors and compared both NET production and ROS generation by these cells in response to IL-8, LPS or PMA stimulation. In addition, in order to provide clinical relevance to this study, we compared NET generation by neutrophils isolated from young and older patients with chronic periodontitis, a chronic infectious bacterial disease of humans that increases in prevalence with aging (White et al., 2012). Periodontitis is initiated by the accumulation of a pathogenic plaque biofilm at and below the gingival margin, but the resulting connective tissue damage is largely mediated by an abnormal host response, dominated by neutrophil inflammation. Thus, neutrophils are able to migrate to the site of infection, but the inflammatory immune response fails to eliminate the pathogenic bacteria (Dias et al., 2013), leading to a chronic nonresolving inflammation (Van Dyke, 2008).

**Results**

**Neutrophil priming significantly enhances NET production and ROS generation**

In the majority of NET studies published to date, resting neutrophils have been used in NET generation assays. Although capable of generating NETs, the use of resting neutrophils does not reflect the *in vivo* state of neutrophils at times of infection when exposure to pro-inflammatory cytokines and bacterial products leads to ‘priming’, which heightens neutrophil responses and microbicidal activity. Thus, to mimic more closely the conditions under which neutrophils *in vivo* would generate NETs, we exposed neutrophils to tumour necrosis factor-alpha (TNF-α), a pro-inflammatory cytokine whose levels are increased during infection and in inflammatory states, prior to stimulation with IL-8 or LPS.

Figure 1(A) shows that NET generation, measured as the DNA content of cell-free supernatants, by TNF-α-primed neutrophils was significantly higher than by resting, unprimed neutrophils treated with IL-8 (P = 0.001) or LPS (P = 0.007), showing that priming enhances NET production. Indeed, when NET formation was studied by fluorescence microscopy, it was evident that in response to both stimuli, primed neutrophils had extruded a greater amount of DNA (Fig. 1B). In addition to enhancing NET production, TNF-α priming significantly increased ROS generation by neutrophils, following IL-8 (P < 0.0001) or LPS (P < 0.0004) treatment (Fig. 1C).

**Age-associated reduction in IL-8 and LPS-induced NET formation**

To investigate the effect of aging on NET formation, neutrophils isolated from healthy young and healthy older adults were primed with TNF-α and stimulated with either IL-8 or LPS, after which the DNA content of cell-free supernatants was measured. Fluorometric quantification revealed that significantly lower amounts of extracellular DNA were extruded by neutrophils of older adults treated with IL-8 (P < 0.02) or LPS (P < 0.04) (Fig. 2A), suggesting that aging in healthy adults is associated with reduced NET production. Fluorescence microscopy...
images confirmed that following IL-8 or LPS stimulation, TNF-α-primed neutrophils from healthy older adults exhibited lower levels of NET formation (Fig. 2B).

Interestingly, no age-associated difference in NET production was observed when neutrophils were treated with PMA (Fig. 2C), a stimulus that bypasses proximal cell membrane receptor signalling events to directly activate protein kinase C, which subsequently phosphorylates and activates NADPH oxidase to generate ROS.

**Effect of age on ROS generation**

Reactive oxygen species generation is a fundamental event required for NET formation as shown by the inability of neutrophils from patients with chronic granulomatous disease to generate NETs upon stimulation (Fuchs et al., 2007; Bianchi et al., 2009; Nishinaka et al., 2011). Having observed an age-related decline in NET generation, we investigated whether reduced ROS production might underlie this observation. Using a luminol-based chemiluminescence assay that detects the generation of MPO-derived oxygen metabolites, which are responsible for triggering NET formation (Dahlgren & Karlsson, 1999; Nishinaka et al., 2011; Palmer et al., 2012), we found that when compared to those from younger adults, TNF-α-primed neutrophils from older adults generated significantly less ROS following either IL-8 or LPS challenge (Fig. 3A–B).

As observed for NET formation, no age-associated differences were found in ROS production by PMA-treated neutrophils (Fig. 3C).

**Effect of age on IL-8 and LPS receptor expression on the neutrophil surface**

The data so far show no age-associated difference in PMA-induced NET or ROS production (Figs 2C and 3C), suggesting that the decline in NET and ROS generation observed following IL-8 and LPS treatment results from an altered proximal signalling response to these stimuli. Neutrophils respond to IL-8 and LPS challenge through their expression of the G protein-coupled receptors CXCR1 and CXCR2, and the pathogen recognition receptor Toll-like receptor 4 (TLR4), respectively. In agreement with a previous study (Fulop et al., 2004), we found no age-related differences in the percentage of neutrophils expressing TLR4 or in the surface density of this receptor (Table 1). Analysis of CXCR1 and CXCR2 expression also revealed no difference between neutrophils from young and old subjects in the percentage of receptor-positive cells or median fluorescent intensity values (Table 1).

**Effect of age on p38 MAP kinase activation in TNF-α-primed neutrophils**

As IL-8 and LPS receptor expression was not affected by aging, we investigated whether an aberrant response to TNF-α priming could have contributed to the age-related decline in LPS- and IL-8-induced NET and ROS generation. Priming neutrophils with TNF-α significantly increases the activity of the mitogen-activated protein kinase (MAPK) p38 (McLeish et al., 1998; Dang et al., 2006). Once active, this kinase has been shown to augment ROS generation in part by phosphorylating the cytosolic NADPH oxidase subunit p47PHOX, triggering its translocation to the plasma membrane (Dang et al., 2006). Thus, having observed an age-related decline in ROS production by TNF-α-primed neutrophils and given the integral role of ROS in NET generation (Fuchs et al., 2007; Bianchi et al., 2009; Palmer et al., 2012), we compared by Western blotting the activation (phosphorylation) kinetics of p38 in neutrophils isolated from healthy young and old subjects during our 15-min TNF-α-
priming protocol. As shown in Fig. 4, no age-associated differences were found in either baseline or TNF-α-induced p38 phosphorylation (Fig. 4A–B). Using p38 MAPK phosphorylation as a marker of TNF-α-priming efficiency, this result suggests neutrophils from young and old subjects were primed to an equivalent degree prior to secondary stimulation.

Table 1 Neutrophils were immunostained for expression of TLR4, CXCR1 and CXCR2. Data are shown for the percentage of cells expressing these receptors and their level of expression (MFI) and are mean ± SEM. MFI; median fluorescent intensity

<table>
<thead>
<tr>
<th></th>
<th>Percentage positive</th>
<th>Surface density (MFI)</th>
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<tbody>
<tr>
<td></td>
<td>Young ( n = 8 )</td>
<td>Old ( n = 8 )</td>
</tr>
<tr>
<td>TLR4</td>
<td>31.5 ± 6.2</td>
<td>43.3 ± 6.8</td>
</tr>
<tr>
<td>CXCR1</td>
<td>99.0 ± 0.37</td>
<td>98.40 ± 0.40</td>
</tr>
<tr>
<td>CXCR2</td>
<td>97.7 ± 1.32</td>
<td>97.44 ± 0.67</td>
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Neutrophil extracellular trap generation in adults with chronic periodontal infection

To ascertain the possible clinical relevance of the age-related reduction in NET generation, we compared NET production induced by PMA or HOCL stimulation between neutrophils obtained from young (mean age 42 ± 4.9 years) and older (mean 57 ± 3.4 years) periodontitis patients, a chronic infectious bacterial disease of humans that increases in prevalence with aging (White et al., 2012). Given that periodontitis leads to tooth loss, the ‘older’ dentate patients in this periodontitis substudy had a slightly lower age range when compared to that of the group of healthy older adults already studied. As with the healthy older subjects, in response to PMA treatment, we found no difference in NET production between the two patient groups (Fig. 5A). However, when exposed to the more physiologically relevant stimulus of HOCL, neutrophils derived from older patients released significantly fewer NETs when compared to neutrophils from young periodontitis patients (P = < 0.03; Fig. 5A). Interestingly, no differences were observed in absolute NET production between patients and age-matched controls for either stimulus (Fig. 5B).

Discussion

Here, we demonstrated that priming neutrophils with TNF-α significantly increased NET formation triggered by a secondary IL-8 or LPS stimulus. This observation is in line with a previous in vitro study, where priming neutrophils with interferon (IFN)-alpha or IFN-gamma prior to their stimulation with the complement protein C5a significantly enhanced NET production (Martinelli et al., 2004), and also concurs with the findings of a recent in vivo study, where increased levels of plasma DNA (a marker of NET generation) were recorded in mice that had been treated with granulocyte colony-stimulating factor prior to low-dose LPS challenge (Demers et al., 2012). TNF-α priming significantly increased neutrophil ROS production induced by IL-8 and LPS, and given the critical role of ROS in NET formation (Fuchs et al., 2007; Bianchi et al., 2009; Nishinaka et al., 2011), this is one mechanism that could explain how pretreatment with TNF-α augments NET production. However, this may not be the only explanation. In a recent study, Wang and co-workers demonstrated that stimulating human neutrophils for 15 min with TNF-α resulted in the activation of PAD4 (Wang et al., 2009). PAD4 catalyses the hypercitrullination of histone H4, a post-translational modification that drives the chromatin decondensation that precedes NET formation (Remijsen et al., 2011). Thus, it may be that in TNF-α-primed neutrophils, the process of NET formation is already underway before these cells are subjected to secondary challenge with NET-inducing stimuli.

Physiological aging is associated with an increased incidence of bacterial infections due, at least in part, to the age-related decline in immunity. In mice, aging has recently been shown to result in a
significant impairment in NET generation both in vitro and in vivo, the latter being associated with marked bacteraemia in a model of Staphylococcus aureus infection (Tseng et al., 2012). Based on this observation, it has been proposed that aberrant NET formation may contribute to the increased incidence of invasive bacterial disease reported by older adults following skin and soft tissue infection (Tseng et al., 2012). However, no study had examined the effect of physiological aging in humans on NET generation. Here, we report that when compared to those from younger subjects, TNF-α-primed neutrophils from healthy older adults generated significantly fewer NETs when challenged with IL-8 or LPS, highlighting an additional age-related defect in neutrophil defences.

Reactive oxygen species generation by TNF-α-primed neutrophils following both IL-8 and LPS stimulation also exhibited a significant age-related decline, offering a mechanistic explanation for the reduction in NET formation with age. That aging has a detrimental effect on ROS production by primed neutrophils has previously been reported by Fulop et al., who demonstrated neutrophils from older adults were primed less efficiently by the pro-inflammatory cytokine granulocyte macrophage colony-stimulating factor, resulting in significantly lower ROS production upon subsequent stimulation (Fulop et al., 2004). Interestingly, we found no age-associated difference in ROS generation or NET formation when neutrophils were challenged with PMA. As a stimulus that bypasses proximal cell membrane receptor signalling events, these results suggest that an aberrant response to TNF-α priming and/or impaired signal transduction following IL-8 and LPS stimulation is responsible for the age-related decline in ROS and NET formation. However, the activation kinetics of p38 MAPK, a signalling element downstream of the TNF-α receptor, were comparable in neutrophils from young and old donors following TNF-α treatment, a finding that is in agreement with the work of Tortorella and colleagues who also reported no age-related difference in p38 MAPK phosphorylation in neutrophils in response to TNF-α priming (Tortorella et al., 2004). Our data also show no effect of age on expression of the IL-8 receptors CXCR1 and CXCR2 or the LPS receptor TLR4, confirming a previous observation (Fulop et al., 2004). With no difference in receptor expression, our data point to age-related changes to proximal events in receptor signalling as being responsible for the decline in NET and ROS production.

The significance of NETs in vivo, with respect to conferring protection in times of pathogenic challenge, has been illustrated in several murine models of infection (Meng et al., 2012; Yipp et al., 2012). Although there are fewer studies in humans, the literature does suggest that reduced NET generation is associated with increased susceptibility to bacterial infections and sepsis (Yost et al., 2009). Recently, evidence has emerged challenging the belief that NETs are directly microbicidal, with their related decline, offering a mechanistic explanation for the reduction in NET formation arising due to compromised NADPH oxidase activation. Therefore, the potential impact of reduced NET generation upon the course of the disease itself is unclear, as recent evidence from our group has demonstrated an autoimmune signature to periodontitis involving antibodies to citrullinated proteins and their un-citrullinated counterparts (dePablo et al., 2013). Citrullinated proteins are a feature of NET production and are associated with autoimmune disease (Kessenbrock et al., 2009), and in this respect, it has been proposed that excess NET formation or reduced NET removal from the periodontal tissues may play a role in periodontitis pathogenesis (Cooper et al., 2013), similar to other autoimmune diseases (Kessenbrock et al., 2009). Thus, the impact of reduced NET production with aging may be disease specific and may differ between diseases of a primarily infectious nature and those where NETs are associated with autoimmune tissue damage.

In summary, we have shown for the first time a propensity for impaired NET release in neutrophils from older adults and an association with reduced ROS production, although reduced NET formation was not further compromised in periodontitis patients.

**Experimental procedures**

**Participants**

Thirty-nine healthy young (mean age 25.54 ± 4.15 years (mean ± standard deviation), 23 women and 16 men) and 38 healthy old (mean age 69.89 ± 5.34 years, 19 women and 19 men) subjects, who did not report any acute infection and were not taking any medication, were recruited to the study to determine the effect of age on NET generation. In addition, 20 patients (9 women and 11 men) with chronic periodontitis (mean age 42.2 ± 7.1, range 31–61 years), defined as previously reported (Dias et al., 2013), and twenty age- and gender-matched controls were enrolled to study NET release by peripheral blood neutrophils in the context of a chronic bacterial infection. Written informed consent was obtained from all participants prior to their enrolment into the study, which was approved by the local research ethics committee (West Midlands Research Ethics Committee number 10/H1208/48).

**Neutrophil isolation and priming**

Peripheral blood neutrophils were isolated by Percoll density gradient centrifugation as described previously (Afford et al., 1992). Neutrophil purity and viability was determined by Giemsa staining (Diff-Quick; Gentaur Europe, Brussels, Belgium) and trypan blue staining, respectively. Routinely, the purity of neutrophil preparations and cell viability were ≥ 98%. Following isolation, neutrophils were resuspended at a concentration of 1–5 × 10⁶ mL⁻¹ in RPMI-1640 media (Sigma-Aldrich, Dorset, UK) supplemented with 2 mM L-glutamine, 100 U mL⁻¹ penicillin and 100 μg mL⁻¹ streptomycin (hereafter referred to as assay media) or Hank’s balanced salt solution (HBSS) supplemented with calcium and
Quantification of extracellular DNA in cell-free supernatants

For the comparison of NET production by neutrophils from young and old volunteers, resting or primed neutrophils (1 × 10^6 in assay media) were treated with 25 nM PMA (Sigma-Aldrich), 10 ng mL⁻¹ IL-8 (R&D Systems, Abingdon, UK), 100 ng mL⁻¹ LPS (Sigma-Aldrich) or assay media for 3 h at 37°C in a humidified 5% CO₂ atmosphere. Postincubation, 1 U mL⁻¹ of micrococal nuclease (MNase; Sigma-Aldrich) and 1 μM of the cell-impermeable DNA binding dye SYTOX Green (Life Technologies, Paisley, UK) were added to cells in order to digest and stain extracellular DNA, respectively. After a 10 min-incubation in the dark at room temperature (RT), cells were pelleted and extracellular DNA content was determined in the cell-free supernatant by measuring fluorescence in a Fluoroskan Ascent fluorometric plate reader (Labsystems Affinity Sensors, Cambridge, UK) or a BioTek™ Synergy 2 fluorometric plate reader (NorthStar Scientific Ltd, Leeds, UK), at an excitation wavelength of 485 nm and emission at 530 nm. Experiments were performed in quadruplicate, and background fluorescence values of neutrophils treated with assay media alone were subtracted from test samples.

In the periodontitis study, 1 × 10⁷ neutrophils in RPMI 1640 media supplemented with 1% glutamine were added to a preblocked (syringe-filtered 1% BSA) 96-well plate and stimulated with either 50 nM PMA or 0.75 mM HOCl and incubated for 3 h (37°C, 5% CO₂). Post-treatment, DNA release was determined as above. No TNF-α priming was employed in this study, as peripheral blood neutrophils from periodontitis patients are already primed within the circulation by plasma cytokines (Wright et al., 2008; Dias et al., 2010).

Assessment of NET formation by immunofluorescence microscopy

Visualization of NETs by immunofluorescence was performed using an adapted version of the protocol described by Brinkmann et al. (Brinkmann et al., 2010). Briefly, resting or primed neutrophils (2 × 10⁵ in assay media) were seeded onto 13-mm circular glass coverslips (VWR International, Leicestershire, UK) and incubated for 30 min at 37°C in a 5% CO₂ atmosphere to allow for cell adherence. Postincubation, neutrophils were stimulated with PMA (25 nM), IL-8 (10 ng mL⁻¹), LPS (100 ng mL⁻¹) or assay media for 3 h (37°C, 5% CO₂), after which cells were fixed in 2% paraformaldehyde for 30 min at RT. Once fixed, coverslips were subjected to three 5-min washes in PBS at RT, after which cells were permeabilized in 0.1% Triton X-100 (Sigma-Aldrich) for 1 min at RT. After a single 5-min wash in PBS, samples were stained with 1 μM SYTOX Green for 5 min, followed by a 5-min wash in PBS. Once stained, specimens were mounted in fluoromount medium onto glass microscope slides (VWR International) and visualized using a LEICA DMI 6000 B microscope (Leica Microsystems, Milton Keynes, UK) at ×20 objective. The criterion used to define a NET was extracellular DNA in the form of a ‘line’ or ‘cloud’.

Measurement of ROS generation

Reactive oxygen species generation was assessed by luminol-amplified chemiluminescence. Resting or primed neutrophils (1 × 10⁵ in HBSS⁺/⁺) were dispensed into wells of a 96-well white-bottomed flat plate (Corning) that contained 1 μM luminol (pH 7.3; Sigma-Aldrich). Cells were then stimulated with LPS (100 ng mL⁻¹), IL-8 (10 ng mL⁻¹), PMA (25 nM) or vehicle control, after which ROS generation was assessed at 1-min intervals for 60 min using a Berthold Centro LB 960 luminometer (Berthold Technologies, Hertfordshire, UK). Experiments were performed in quadruplicate, with ROS production measured as relative light units (RLU) and calculated as area under the curve (AUC).

Measurement of p38 MAP kinase activation by Western blotting

Freshly isolated resting neutrophils (5 × 10⁶ mL⁻¹ in assay media) were treated with 10 ng mL⁻¹ TNF-α for 0–15 min at 37°C in a humidified 5% CO₂ atmosphere. Poststimulation, neutrophils were pelleted by centrifugation (1500 g, 2 min, 4°C), supernatants discarded and cell lysates prepared by the addition of hot sodium dodecyl sulphate (SDS) sample buffer [4% SDS (w/v), 0.1 M dithiothreitol, 20% glycerol (v/v), 0.0625 M Tris–HCl and 0.004% bromophenol blue (w/v)]. Lysates were separated on 10% SDS polyacrylamide gels and proteins transferred to polyvinylidene difluoride membranes (Scientific Laboratory Supplies, Yorkshire, UK). To prevent nonspecific binding, membranes were incubated with 5% bovine serum albumin (BSA) in Tris-buffered saline (TBS; 200 mM Tris (pH 7.5), 1.5 mM NaCl) containing 0.1% Tween-20 (TBST) for 1 h at RT, before being probed overnight at 4°C with a rabbit anti-human phospho-p38 antibody (diluted 1:1000 in TBST containing 2.5% BSA; Cell Signalling Technology, Hertfordshire, UK). Horseradish peroxidase activity was detected using enhanced chemiluminescence (Geneflow, Staffordshire, UK). To confirm equal loading of proteins, blots were washed in TBST and incubated for 1 h at RT with a goat anti-rabbit secondary antibody conjugated to horseradish peroxidase (HRP; diluted 1:4000 in TBST; GE Healthcare, Buckinghamshire, UK). Horseradish peroxidase activity was determined using enhanced chemiluminescence (Geneflow, Staffordshire, UK). To confirm equal loading of proteins, blots were striped with stripping buffer (10% (w/v) SDS, 62.5 mM Tris–HCl (pH 6.8), 0.08% (w/v) β-mercaptoethanol) for 45 min at 50°C, washed for 2 h in distilled H₂O and blocked for 1 h at RT in 5% BSA/TBST, before being probed overnight at 4°C with a p38 antibody (diluted 1:2000). Densitometry was performed using the National Institute of Health IMAGEJ software package.

Measurement of surface receptor expression

Freshly isolated neutrophils (1 × 10⁷) were stained with 2 μg mL⁻¹ anti-CXCR1-FITC (R&D Systems; clone 42075), 3 μg mL⁻¹ anti-CXCR2-FITC (R&D Systems; clone 48311) or 5 μg mL⁻¹ anti-TLR4-APC (clone HTA125; eBioscience, Hatfield, UK) antibodies or their relevant concentration-matched isotype controls for 20 min on ice. Postincubation, cells were washed once in PBS/1% BSA, resuspended in 100 μL PBS and transferred to polypropylene FACs tubes. Flow cytometry was conducted using a CyAnADP™ bench-top cytometer (Dako, Cambridgeshire, UK) with 5000-10 000 neutrophils acquired for analysis, which was performed using Summit v4.3 software (Dako Colorado Incorporation, Fort Collins, CO, USA). The percentage of antigen-positive neutrophils was recorded along with the corresponding median fluorescence intensity (MFI) values.

Statistical analysis

Statistical analyses were performed using GraphPad Prism® software (GraphPad Software Ltd, San Diego, CA, USA). Data distribution was examined using the Kolmogorov–Smirnov test. For data that followed a normal distribution, unpaired and paired student T-tests were performed to assess differences between two independent groups or matched
paired samples, respectively. A repeated measures ANOVA was used to compare more than two matched observations. For non-normally distributed data, a Mann–Whitney U test was used. Data are presented as mean ± standard error of the mean (SEM). The minimum level of confidence at which results were considered statistically significant was 
P ≤ 0.05.

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Author contributions
J.H performed the experimental work, data analysis and wrote the manuscript. M.G and P.H performed experimental work and data analysis. A.L and H.G performed experimental work. E.S assisted with subject recruitment. I.L.C and J.M.L designed the study and contributed to data interpretation and writing of the manuscript.

Conflict of interest
None declared.

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