Prenatal Diagnosis of Congenital Adrenal Hyperplasia Caused by P450 Oxidoreductase Deficiency


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Context: Mutations in the electron donor enzyme P450 oxidoreductase (POR) result in congenital adrenal hyperplasia with apparent combined 17α-hydroxylase/17,20 lyase and 21-hydroxylase deficiencies, also termed P450 oxidoreductase deficiency (PORD). Major clinical features present in PORD are disordered sex development in affected individuals of both sexes, glucocorticoid deficiency, and multiple skeletal malformations.

Objective: The objective of the study was to establish a noninvasive approach to prenatal diagnosis of PORD including assessment of malformation severity to facilitate optimized prenatal diagnosis and timely treatment.

Design: We analyzed 20 pregnancies with children homozygous or compound heterozygous for disease-causing POR mutations and 1 pregnancy with a child carrying a heterozygous POR mutation by recording clinical and biochemical presentations and fetal ultrasound findings. In 4 of the pregnancies (3 homozygous and 1 heterozygous for disease-causing POR mutations), prenatal analysis of steroid metabolite excretion in maternal urine was carried out by gas chromatography/mass spectrometry during gestational weeks 11–23.

Results: Pregnancy complications in our cohort included maternal virilization (6 of 20) with onset in the second trimester. Seven pregnant women presented with low unconjugated estriol at prenatal screening (triple or quadruple antenatal screening test). Overt dysmorphic features were noted in 19 of the 20 babies at birth but observed in only 5 by prenatal ultrasound. These 5 had the most severe malformation phenotypes and poor outcome, whereas the other babies showed normal development. Steroid profiling of maternal urine revealed significantly increased steroids of fetal origin, namely the pregnenolone metabolite epiallopregnanediol and the androgen metabolite androsterone, with concomitant low values for estriol. Diagnostic steroid ratios conclusively indicated PORD as early as gestational week 12. In the heterozygous pregnancy, steroid ratios were only slightly elevated and estriol excretion was normal.

Conclusion: Prenatal diagnosis in PORD is readily established via urinary steroid metabolite analysis of maternal urine. Visible malformations at prenatal ultrasound predict a severe malformation phenotype. (J Clin Endocrinol Metab 98: E528–E536, 2013)
Congenital adrenal hyperplasia due to P450 oxidoreductase deficiency (PORD) is a condition with autosomal recessive inheritance, caused by mutations in the P450 oxidoreductase (POR) gene (1–3). PORD patients present with biochemical evidence of partial deficiencies in 21-hydroxylase (CYP21A2) and 17α-hydroxylase/17,20-lyase (CYP17A1) activities and manifest clinically in many but not all cases with disorder of sex development (DSD) in both sexes; glucocorticoid deficiency; and skeletal malformations including craniosynostosis, midface hypoplasia, radiohumeral synostosis, and phalangeal abnormalities (4–8). POR is the crucial electron donor to all microsomal cytochrome P450 (CYP) enzymes including the steroidogenic enzymes CYP21A2, CYP17A1, and P450 aromatase (CYP19A1), thus explaining the deficiencies in steroid synthesis observed in patients with PORD (2–3). POR also serves as the mandatory electron donor to squalene epoxidase and 14α-lanosterol demethylase (CYP51A1) catalyzing important steps in cholesterol synthesis. Sterols are involved in hedgehog-mediated regulation of fetal bone development. Hence, decreased 14α-lanosterol demethylase activity as documented ex vivo in a patient with PORD (9) may explain the observed skeletal malformation phenotype, which resembles that observed after the disruption of downstream processing of lanosterol by fluconazole (10–13). Distinct POR mutations can differentially affect the activities of electron-accepting CYP enzymes, a likely explanation for the observed phenotypic variability (5, 14–16).

The prognosis and outcome of PORD patients relies heavily on early diagnosis and timely treatment with optimal perinatal care including assessment of glucocorticoid replacement requirements as well as corrective treatment of skeletal malformations and DSD manifestations. Therefore, the aim of this study was to establish tools for prenatal diagnosis and estimation of disease severity.

**Subjects and Methods**

**Patients**

We analyzed pregnancy data from 20 patients (8 males, 12 females) with genetically confirmed PORD and 1 pregnancy with a fetus carrying a heterozygous mutation in the POR gene. Four of these pregnancies have been prospectively monitored (PORD A, B, C, and the heterozygous baby D) and are described in detail below.

PORD pregnancy A was the second pregnancy of healthy, nonconsanguineous parents. Their first child (46,XX) was affected by PORD, born with overt craniofacial malformations and severely virilized external genitalia (Prader IV); a cosynotropin test confirmed adrenal insufficiency. The pregnancy had been complicated by signs of maternal androgen excess (acne from gestational weeks 14 to 16 and maternal voice deepening noted from week 24). Genetic analysis detected homozygous POR p.A287P [g.29,536 G>C] mutations, with both parents being heterozygous carriers.

During the second pregnancy, the mother did not experience signs of androgen excess. Prenatal ultrasound examinations for detection of skeletal malformations were carried out every 4–6 weeks, showing normal fetal development without visible malformations throughout the pregnancy. Amniocentesis was carried out during the 16th week of gestation and direct sequencing of amniocyte DNA revealed the same homozygous p.A287P mutation as in the first child; the karyotype was confirmed as 46,XY. After an otherwise uneventful pregnancy, labor initiated spontaneously at gestational week 35. Birth weight was 2830 g [0.9 SD score (SDS)], length was 53 cm (3.2 SDS), and head circumference was 34 cm (1.4 SDS) (SDS according to British 1990 growth reference data). Initially the patient suffered from respiratory distress and had to be referred to the neonatal intensive care unit for continuous positive airway pressure ventilation therapy. The baby had midface hypoplasia, a pear-shaped nose, a soft palate cleft, and mild acrocephaly without overriding cranial sutures and a mild left elbow contracture. Other malformations included arachnodactyly, rocker-bottom feet, and a narrow chest with mild pectus excavatum. The penis was of normal size and the testes were bilaterally descended. He was started on hydrocortisone replacement therapy in the first week of life. At 6 weeks of age, surgery for bilateral inguinal and umbilical hernias was performed. He underwent surgery for coronal and metopic synostosis at age 6 months; the soft palate cleft was repaired at 1 year of age. Postnatal growth and development including cognitive function were normal.

PORD pregnancy B was the first pregnancy of healthy, nonconsanguineous parents. Routine prenatal ultrasound at 16 weeks revealed an abnormally shaped skull, bowed femora, bilateral radioulnar synostosis, and ambiguous genitalia. Based on the severe malformation phenotype, the parents decided for termination of pregnancy (TOP), which was carried out in the 25th week of gestation. The postmortem on the fetus was consistent with a severe malformation phenotype including coronal synostosis, severe midface hypoplasia, bilateral radioulnar synostosis, and bowed femora (all confirmed on posttermination x-ray, Figure 1A) as well as microgenia, absent scrotum, and undescended testes. The karyotype was 46,XY.

Sequencing analysis of fetal DNA commenced after termination revealed compound heterozygosity for p.A287P and a frameshift mutation I444fsX6 (g.31,146dupC).

PORD pregnancy C was the second pregnancy of healthy, nonconsanguineous parents (same father as in the first pregnancy). The first pregnancy had resulted in a healthy baby girl. The course of the second pregnancy was unremarkable, without signs of maternal virilization. At 19 weeks’ gestation the triple test (human chorionic gonadotropin, α-fetoprotein, and unconjugated estriol) showed undetectable levels of estriol (<0.02 ng/mL). At 20 weeks’ gestation, the following anomalies were detected by prenatal ultrasound: bowed femurs bilaterally, craniosynostosis, intrauterine growth retardation, and abnormal scissoring of the lower extremities (right foot positioned on left flank). A fetal magnetic resonance imaging did confirm the above. The amniocentesis performed at gestational week 20 revealed a female karyotype (46, XX). In gestational week 39, the baby was delivered by cesarean section without any complications. After birth the baby experienced respiratory distress due to...
choanal stenosis [Apgar indices 8 at 1 minute, 9 at 5 minutes; birth weight 2610 g (−1.16 SDS), length 48 cm (−1.67 SDS), head circumference 31 cm (−2.89 SDS)] (SDS according to British 1990 growth reference data). Postnatally the following malformations were evident: craniosynostosis without hydrocephalus (Figure 1B), bilateral choanal stenosis, craniovertebral junction deformity, bilateral vesicoureteral reflux, bilateral hydronephrosis, contractures of the fingers and toes, arachnodactyly, humeroradial-ulnar synostosis of the upper extremities, shortened femurs, and humeri bilaterally. The external genitalia had normal female appearance. The baby was admitted to the intensive care unit and initially ventilated; concurrently hydrocortisone replacement was initiated. On day 69 she died due to respiratory insufficiency, most likely as a consequence of central apnea. Genetic analysis revealed compound heterozygous POR mutations (p.A287P/p.Q455Rfs166X).

Heterozygous PORD pregnancy D was the second pregnancy of a consanguineous Pakistani couple. The first baby (46, XY) had presented at delivery with a dysmorphic face including frontal bossing, hypotelorism, proptosis, low nasal bridge, short columella, pronounced philtrum, and small, low-set ears. Other malformations at birth included bilateral radiohumeral synostosis and arachnodactyly. During the first trimester of pregnancy, the mother had started to develop hirsutism, resolving after delivery. POR sequencing in the index child revealed a homozygous p.R498P mutation.

Because of this history, the second pregnancy was closely monitored by ultrasound, but no malformations could be detected. Chorionic villous sampling during gestational week 11 revealed a 46,XX karyotype. The baby was well and healthy at delivery, with no sign of malformations; genetic analysis after birth revealed heterozygosity for p.R498P.

PORD malformation score

The clinical malformation phenotype was assessed after birth using a scoring algorithm we recently developed (8). In brief, this score assesses the absence or presence of 6 different types of malformations: midface hypoplasia, craniosynostosis, phalangeal malformations, large joint synostoses, femoral bowing, and additional malformations. The severity of each malformation is graded on a scale ranging from 0 (not present), 1 (mild), 2 (severe), to 3 (severe with additional complications). The minimum of the total PORD malformation score is 0 (no malformations), and the maximum score is 16 (most severe malformation phenotype).

Urinary steroid profiling

Urinary steroid metabolite excretion was analyzed by gas chromatography/mass spectrometry (GC/MS). Characterization and quantification of all excreted steroids has been previously established in a longitudinal study of a pregnancy with a PORD fetus (17). This allowed designation of key analytes for diagnosis of the condition by the development of an abbreviated selected ion monitoring quantitative method for their analysis. In the 4 prospective PORD pregnancies followed up, maternal urine for steroid metabolite assessment was collected during gestational weeks 12, 22, and 28 (PORD A), 23 (PORD B), 20 (PORD C), and 11 (heterozygous PORD pregnancy D), respectively, and results were compared with the urinary steroid profiles from 60 apparently normal pregnancies of gestational weeks 15–23.

Results

Prenatal maternal and fetal findings

Overall, 7 of 20 pregnancies with a baby affected by PORD were complicated by signs of maternal virilization,
which usually manifested during the second trimester (Table 1). Clinical signs and symptoms included acne, hirsutism, and deepening of the voice, all resolving within a few weeks after delivery. The occurrence of prenatal maternal virilization did not show any conclusive correlation to genotype or sex of the affected child. In 7 pregnancies a low unconjugated estriol were found at routine prenatal screening (triple or quadruple antenatal screening test) during gestational weeks 14–19 (cases 2, 3, 15, 17, PORD A, PORD B, and PORD C).

Prenatal ultrasound detected skeletal malformations in 5 of the 20 PORD pregnancies (Table 1), in all cases describing clearly visible bowed femora. These 5 included the above-described PORD pregnancies B and D and cases 1–3 (Table 1). In case 1, bilaterally angulated femora were detected for the first time at gestational week 21 and a prominence of the forehead in addition to short and bent femora at gestational week 25. In case 2, ultrasound showed bowed femora and flexed distal limb joints of the extremities at gestational week 16. In case 3, ultrasound in gestational week 17 revealed a flattened nasal bridge, low set ears, abnormal hand posturing, ambiguous genitalia, and bowed humeri, femora and tibiae.

However, despite this low detection rate by prenatal ultrasound, at birth all 20 PORD cases presented with overt skeletal malformations; the median PORD malformation score of the cohort was 7.5, ranging from 4 to 15. Of note, the median malformation score in the 15 patients without evidence of malformations on prenatal ultrasound was 6 (range 0–10), ie, spanning from absent to mild and moderate malformations. By contrast, the 5 patients with prenatally detected malformations presented with the most severe skeletal malformation phenotypes (all scores >10, median score 15) (Table 1). Of these 5 pregnancies, 1 ended after the termination of pregnancy because of the severity of the observed malformations. Three patients died, due to recurrent respiratory infections (case 1), accidental displacement of the tracheostomy tube

<table>
<thead>
<tr>
<th>Patient Number</th>
<th>Genotype</th>
<th>Karyotype</th>
<th>Malformations Detected by Prenatal Ultrasound</th>
<th>Total Malformation Score at Birth*</th>
<th>Maternal Androgen Excess</th>
<th>DSD Detected by Prenatal Ultrasound</th>
<th>DSD Evident at Birth</th>
<th>Outcome</th>
<th>Follow-Up Period</th>
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<tr>
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<td>IVS7 + 2bp/pT</td>
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<td>15</td>
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<td>15</td>
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<td>Y</td>
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<td>15</td>
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<td>N</td>
<td>N</td>
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<td>23 mo</td>
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<td>Y</td>
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<td>4 y</td>
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<td>N</td>
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<td>N</td>
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<td>Y</td>
<td>N</td>
<td>N</td>
<td>Normal, normal intelligence</td>
<td>2 y</td>
</tr>
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Abbreviations: E3, estriol; −, no mutation detected; N, no; n.d., not determined; Y, yes. Bold indicates cases with visible malformations on prenatal ultrasound and a total malformation score above 10.

* For a detailed description of the PORD malformation score, please see published reference (8).

** The malformation score for case PORD B is likely to be underestimated as scoring based on postmortem report rather than clinical examination.

^ Normal, normal postnatal development in terms of growth, speech, motor, and cognitive function.
The typical features of PORD were identified in all urine samples collected in the 3 affected pregnancies during gestational weeks 12–28. All maternal steroid metabolites in urine normalized after delivery (data not shown), consistent with a fetal source of abnormality during pregnancy.

In the heterozygous pregnancy D, the above-mentioned pathogenic urinary metabolite ratios for PORD were only slightly elevated, and the estriol concentration was completely normal.

Discussion

Analysis of the course of 20 pregnancies with fetuses affected by PORD and 1 pregnancy heterozygous for a POR mutation revealed 3 cardinal features indicative of the presence of the disorder that may be evident prenatally: maternal virilization, severe bone malformations detected by fetal ultrasound, and, invariably in the prospectively monitored cases, a distinctly altered steroid metabolite excretion in maternal urine indicative of PORD from gestational week 12 onward. Remarkably, these changes were uniformly present in all affected pregnancies and did not vary as a function of genotype or phenotype, similar to our previous experience with urine GC/MS analysis in affected patients (8).

The characteristically low maternal urinary estriol excretion during the affected pregnancies was mirrored by the finding of low serum estriol in the antenatal multiple marker screening test carried out in 7 of the patients. These findings were similar to those in the 2 previously published prenatal cases, describing low urinary and serum estriol, respectively (17, 18). The maternal blood screening test is usually performed between the 16th and 18th week of pregnancy. The triple test comprises measurement of serum α-fetoprotein, human chorionic gonadotropin, and estriol, but a quad test is gaining popularity and also includes inhibin A. The primary object of the screening test is for detection of neural tube defects and Down’s syndrome (19, 20).

In a physiological pregnancy, estriol is generated in the placenta from fetal dehydroepiandrosterone (DHEA), the major product of fetal adrenal steroid synthesis (21). In PORD, 3 enzymatic reactions that are involved in the pathway leading to estriol synthesis are dependent on electron transfer by POR and thus may be compromised. First, mutant POR results in impaired 17-hydroxylase/17,20-lyase activity of CYP17A1. This consequently diminishes the production of DHEA and downstream androgens, thereby decreasing the amount of androgenic substrate available for conversion to estriol by aromatase (Figure 3).
Second, steroid 16α-hydroxylase is required for the production of 16α-hydroxy-DHEA, but there is conflicting evidence as to whether it has diminished activity due to PORD. Finally, CYP19A1 aromatase activity itself could be impaired by mutant POR, further contributing to low estriol production, which again could show some variability in PORD with distinct mutants having been shown to have a differential impact on aromatase activity in vitro (15). Impaired aromatase activity would predictably result in an accumulation of its immediate precursor 16α-hydroxyandrostenedione and reflected in increased excretion of its metabolite 16α-hydroxyandrostenedione. These markers were modestly raised in the pregnancies described here; however, similar to our previously reported case (17), these increases were certainly not in the range that would be expected in cases of aromatase deficiency due to CYP19A1 mutations. Thus, it appears that the major determinant of low estriol in PORD is reduced synthesis of DHEA due to impairment of 17,20-lyase activity.

Low estriol on its own, however, is a nonspecific marker that can be associated with a number of conditions, including several single gene disorders underlying sterol and steroid synthesis defects such as Smith-Lemli-Opitz syndrome, steroid sulfatase deficiency, aromatase deficiency, and severe neonatal adrenal insufficiency due to mutations in NR0B1 (dosage-sensitive sex reversal-adrenal hypoplasia congenita critical region on the X chromosome, gene 1), NR5A1 (steroidogenic factor 1), CYP11A1 (P450 side chain cleavage enzyme), and TBX19 (TPIT, corticotroph specific T-box transcription factor) (22–35). However, here a distinct combination of low estriol and other fetal steroid markers enabled us to make the specific diagnosis of PORD based on the analysis of maternal urine at midpregnancy. Interestingly, normal estriol in the pregnancy with the heterozygous baby clearly distinguished this pregnancy from the PORD pregnancies. In this case the PORD specific ratios showed only very mild aberrations from the control cohort due to a normal estriol, suggesting that the baby was not affected by PORD. In addition to low estriol, a characteristic finding in the PORD pregnancies was the highly increased excretion of EpialloPD, a metabolite of fetal pregnenolone, which accumulates in PORD due to the concurrent impairment of CYP17A1 and CYP21A2 activities. Thus, in PORD EpialloPD essentially replaces estriol as the major ter-

![Figure 2. Proposed urine steroid metabolite ratios for a prenatal diagnosis of PORD. Illustrated are normative values from gestational week 15 to week 23 (n = 60) and 4 pregnancies with affected PORD babies and 1 pregnancy with a heterozygous carrier. Panel A shows reduced estriol relative to the excretion of the progesterone metabolite PD. Panel B shows increased value of the EpialloPD to estriol ratio resulting from pregnenolone accumulation caused by attenuated 17-hydroxylase/17,20-lyase and decreased estriol production. Panel C shows increased 16α-hydroxyandrostenedione excretion in relation to decreased estriol, indicative of the effect of mutant POR on CYP19A1 aromatase activity. Panel D shows increased androsterone excretion compared with etiocholanolone, indicative of alternative pathway androgen synthesis activity (2). Panel E gives the abbreviation and systematic name of the urinary steroid metabolites and from which they derive. The ratios denoted by an asterisk were obtained from a previously published report on a patient (17).](image-url)
minal product of fetal adrenal steroid synthesis and metabolism (17). Consequently, the altered fetal synthesis and metabolism in PORD pregnancy is most effectively documented by measuring the urinary ratio of the abnormal end-product (EpialloPD) to the normal end-product (estriol). Importantly, we could identify all 4 prospectively monitored PORD cases by the above-described urinary steroid ratios despite an appreciable degree of biochemical variability observed in PORD. The identified ratios in our view reflect the key biochemical changes detectable in prenatal PORD cases. However, further studies in larger cohorts will be needed to further validate our findings. We should add that 2 other rare disorders, 17-hydroxylase deficiency and cytochrome b5 deficiency, are also associated with attenuated 17-hydroxylase/17,20 lyase and pregnenolone build-up and may therefore give similar results regarding the increased excretion of EpialloPD and low excretion of estriol.

However, in both conditions the maternal urine androstosterone to etiocholanolone ratio would be normal and not elevated as was observed in PORD.

In normal pregnancies the excretion of the 2 epimeric major androgen metabolites androsterone (5α-reduced) and etiocholanolone (5β-reduced) is approximately equivalent. Increased excretion of 5α-reduced androgens was demonstrated by the significantly elevated androstosterone to etiocholanolone ratios in the PORD pregnancies reported here and also in the previously reported case (17), providing a clue to the cause of maternal and female fetal virilization in PORD. We have previously proposed that this is caused by increased activity of an alternative androgen biosynthetic pathway during fetal and early neonatal life (2), resulting in the increased production of androsterone and 5α-dihydrotestosterone. PORD results in an accumulation of 17α-hydroxyprogrenenolone and 17α-hydroxyprogesterone that can enter the alternative pathway (Figure 3). This pathway is accessed through a 5α-reduced 17-hydroxy pregnanone intermediate (3α,17α-dihydroxy-5α-pregnan-20-one) that is a substrate with higher affinity for the 17,20 lyase activity of CYP17A1 than the other 17-hydroxysteroids, 17α-hydroxyprogrenenolone and 17α-hydroxyprogesterone (36). In PORD, 5α-reduced 17-hydroxy pregnanone intermediate conversion by 17,20 lyase via the alternative pathway results in increased fetal production of androsterone, which in turn can be converted to active androgen in the fetoplacental unit, even while the conventional (classic) androgen synthesis pathway is largely blocked. It is important to note that the presence of maternal virilization was independent of genotype and other phenotypic characteristics (Table 1) and, for example, was observed in 1 affected child but not in its sibling with the same genotype (PORD pregnancy A). Future research will have to be conducted to reveal the underlying mechanisms explaining this disparity.

Most of the 20 patients we reported here underwent only routine prenatal ultrasound because no malformations were suspected prior to birth. Anatomic ultrasound performed in prenatal specialist centers in the second trimester has a reported detection rate of approximately 70%–90% for fetal congenital abnormalities (37, 38). Thus, one could assume that specialized malformation ultrasound may have detected skeletal abnormalities in the milder cases. However, this assumption contrasts with the longitudinally documented findings in a PORD pregnancy (PORD pregnancy A) in which the pregnancy was very closely monitored by specialist ultrasound after the birth of an index patient with multiple skeletal malformations of intermediate severity. However, prenatal ultrasound failed to detect the malformations in the second child, which were clearly vis-
ible after birth. Of note, in our cohort, prenatal ultrasound seemed to indicate disease severity and long-term outcome in PORD. All 5 patients with the highest malformation scores based on postnatal clinical assessment were found to have skeletal malformations during routine prenatal ultrasound examinations, whereas no malformations were detected in the 15 patients with clinically intermediate or mild malformation phenotypes. This would suggest that only the most severe PORD-related malformation phenotypes can be detected reliably by routine prenatal ultrasound, and according to our data, these are invariably associated with a poor prognosis, whereas children with intermediate, mild, or absent malformation phenotypes had a good clinical outcome.

The option of prenatal diagnosis always comes with ethical challenges and when deciding on the pros and cons of termination of pregnancy, it is important to consider the normal cognitive capacity and normal intellectual development in affected children irrespective of presence and severity of malformations. The parents of the child described as PORD pregnancy A asked for a prenatal diagnosis to ascertain whether they should temporarily move from a very rural area closer to a tertiary care hospital offering peri- and postpartal fetal medicine and pediatric specialist care. Fetal sexing using fetal cells circulating in maternal blood will become available in the foreseeable future. The parents of PORD pregnancy A considered prenatal dexamethasone treatment to avoid the virilization observed in their firstborn daughter, the index child. This girl carried homozygous A287P mutations associated with 46,XX DSD but normal genital phenotype in affected 46,XY individuals (8). In PORD it is highly likely that the major part of virilization is driven by the alternative androgen synthesis pathway (2). Thus, irrespective of other concerns regarding prenatal dexamethasone treatment, in a male fetus such a treatment is highly likely to significantly reduce the substrate flow to the alternative pathway and consequently result in undervirilization.

In conclusion, our data demonstrate that diagnosis of PORD can be established prenatally via GC/MS steroid analysis of maternal urine, with further data needed on diagnostic sensitivity and on the differentiation of fetuses with homozygous POR mutations from heterozygous carriers. Prenatal ultrasound is helpful in estimating the severity of the PORD-associated skeletal malformation phenotype and appears to correlate with long-term outcome. These 2 noninvasive diagnostic tools are of significant promise for the prenatal diagnosis of PORD in pregnancies affected by maternal virilization or fetal malformations and those occurring after the birth of an index child with PORD. This diagnostic approach will hopefully facilitate improved perinatal care and the timely involvement of a multidisciplinary specialist team including endocrine expertise. Although on an individual patient basis, urinary steroid profiling of pregnant women with low estriol in the triple or quadruple screening test is readily performed and a powerful tool for the early diagnosis of PORD, more routine and widespread use of the technique for identifying steroid or sterol synthesis disorders such as PORD, steroid sulfatase deficiency, and Smith-Lemli-Opitz syndrome in all low-estriol pregnancies is unlikely to occur because of the costs involved.

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References


