Residual association at C9orf72 suggests an alternative amyotrophic lateral sclerosis-causing hexanucleotide repeat

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1. Introduction

Amyotrophic lateral sclerosis (ALS) is a neurodegenerative disease of motor neurons that causes relentless paralysis, with death occurring within 2–5 years. The lifetime prevalence of ALS is 1 in 100,000 (Johnston et al., 2006) with peak onset at approximately age 60 years, and an increased incidence in men (Chio et al., 2009).

Genetic studies of ALS have begun to yield insights, with 10–30% of cases resulting in loss of association for rs3849942 (p = [3 × 2] × 10^-10, rank 7/442,057; rs903603, p = [7 × 6] × 10^-6, rank 2/442,057). Removal of the mutation-carrying cases resulted in loss of association for rs3849942 (p = [2 × 6] × 10^-9, rank 1225/442,068), but had little effect on rs903603 (p = [1 × 9] × 10^-5, rank 8/442,068). Those with a risk allele of rs903603 had an excess of apparent homozygosity for wild-type repeat alleles, consistent with polymerase chain reaction failure of 1 allele because of massive repeat expansion. These results indicate residual association at the C9orf72 locus suggesting a second disease-causing repeat mutation.

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2010), but this distinction is artificial, with an increased risk of other neurodegenerative diseases including Parkinson’s disease, and frontotemporal dementia (Byrne and Hardiman, 2010; Millecamps et al., 2010), and an increased risk of ALS in relatives of those with apparently sporadic disease (Hanby et al., 2011). Furthermore, variants in every Mendelian ALS gene have also been reported in those with apparently sporadic ALS (Al-Chalabi and Lewis, 2011; van Blitterswijk et al., 2012), and the heritability of apparently sporadic ALS has been estimated at 61% (Al-Chalabi et al., 2010).

Genome-wide association studies in ALS and FTD recently identified a locus on chromosome 9 within a known linkage peak for ALS-FTD (Laaksovirta et al., 2010; Shatunov et al., 2010; van Es et al., 2009). The disease-causing mutation has now been identified as a massive expansion mutation of a hexanucleotide repeat between noncoding exons 1A and 1B of C9orf72 (Dejesus-Hernandez et al., 2011; Renton et al., 2011). Unaffected individuals have 2–23 repeats of the (GGGGCC)n hexanucleotide microsatellite, though the pathological repeat mutation might be several hundred repeats. The mutation is interesting in manifesting with at least 4 phenotypes: ALS, ALS-FTD, FTD, and normality.

Although 3 different genome-wide association studies all identified rs3849942 as the most associated SNP at the C9orf72 locus, and it is clear that this is in strong linkage disequilibrium with the repeat mutation (Smith et al., 2013), the UK subpopulation showed stronger genome-wide significant association at a different SNP, rs903603. The most parsimonious explanation is that rs903603 and rs3849942 are in strong linkage disequilibrium and both tag the pathological expansion. If that is the case, one would expect that removal of cases with the repeat mutation from the analysis would obliterate the association at both SNPs. We therefore tested this hypothesis in the UK population.

2. Methods

2.1. Sample collection

Whole blood samples from the Motor Neuron Disease (MND) DNA Bank were used. Selection was based on no family history of ALS, being of white European ancestry, and disease onset January 2002 or after. Control samples were obtained from the Depression Case Control (DeCC) study, the Bipolar Affective Case Control Study (BACCS), from Panos Deloukas of the Wellcome Trust Sanger Institute (Cambridge, UK), and the British 1958 birth cohort DNA collection. This project was ethnically approved.

2.2. Sample preparation

DNA was extracted from blood samples by standard methods within 1 week of bleeding and stored at the UK DNA Banking Network in Manchester. Samples were bar-coded in a tracking system to reduce risk of clerical error.

2.3. Genotyping

Genotyping and quality control was carried out as described previously (Shatunov et al., 2010), but in brief, genome-wide association was performed using various Illumina DNA microarrays followed by standard quality control and association testing using PLINK (Purcell et al., 2007).

The mutation was identified using repeat-primed polymerase chain reaction (PCR) as described (Dejesus-Hernandez et al., 2011). Nonmutation repeat length for alleles of the hexanucleotide repeat was quantified using amplified fragment length polymorphism (AFLP), using fluorescently-labeled primers as detailed previously (DeJesus-Hernandez et al., 2011). Both analyses were performed using an automated 3130XL capillary electrophoresis-based Genetic Analyzer and GeneMapper v4.0 software (Applied Biosystems).

2.4. Statistical quality control

Statistical quality control excluded individuals with incongruent sex, those marked for exclusion by the genotyping institution, those with low genotyping per individual <0.02, those with phenotype missingness <0.02, those with low heterozygosity (p < 0.05), those exhibiting identity by descent sharing >5% of alleles with another participant, and those not of white European ancestry.

Excluded SNP markers were those with a minor allele frequency less than 0.015, SNP missingness <0.02, significant departure from Hardy-Weinberg equilibrium (p < 0.001), those marked for exclusion by the genotyping institution, and those on chromosome X.

2.5. Phasing and imputation analyses

Phasing and imputation was completed using 1000 Genomes v2.20101123 autosomal release (http://www.1000genomes.org/), performed using MaCH 1.0.18 (http://www.sph.umich.edu/csg/abecasis/MACH/index.html) and minimac (http://genome.sph.umich.edu/wiki/Minimac). We performed association analysis on imputed data using ProbABEL (http://www.genabel.org/packages/ProbABEL), mach2dat (http://github.com/sph.umich.edu/wiki/Mach2dat:_Association_with_MACH_output), and PLINK v1.07 (http://pngu.mgh.harvard.edu/~purcell/plink/index.shtml), after creating input files using the GenGen (http://www.openbioinformatics.org/engen/) conversion tool. For GenGen, we set the r² threshold as 0.3 and, based on genotype posterior probabilities derived in MaCH, a quality control threshold of 0.9.

2.6. Association and haplotype analysis

To test for residual association and identify SNPs not related to the mutation we used logistic regression, modeling case-control comparisons with sex as a covariate, stratified by the presence or absence of the repeat mutation.

We set the Bonferroni-corrected threshold of statistical significance at 1.13 \times 10^{-5}, and the genome-wide threshold as 5 \times 10^{-8}. We used the R statistical package to construct Manhattan and Q-Q Plots, LocusZoom (http://csg.sph.umich.edu/locuszoom/) for regional Manhattan plots, and Hapioview for haplotype block maps (http://www.broadinstitute.org/scientific-community/science/programs/medical-and-population-genetics/hapioview/hapioview) for Manhattan plots, and Hapioview for haplotype block maps (http://www.broadinstitute.org/scientific-community/science/programs/medical-and-population-genetics/hapioview/hapioview).

For linear regression analysis of the nonmutated microsatellite repeat lengths as a quantitative variable we used PLINK v1.07 and IBM SPSS 20. For haplotype analyses we used PLINK v1.07 sliding windows between 2 and 100 to explore significantly associated haplotypes using the phased imputed dataset. The prevalence of the most associated haplotypes were refined in Hapioview and tested for their association with ALS using an omnibus haplotype association analysis.

We ran all genetic tests under additive, dominant, and recessive models, with little difference in results between dominant and additive models. We have reported the additive model for association of SNP alleles with the repeat mutation, and a dominant model (using the larger allele) for the analysis of repeat lengths.
2.7. Sequencing C9orf72

Four hundred eighty DNA samples of patients with sporadic ALS were pooled in sets of 8. Each pool was tagged with 1 of 12 identifying DNA sequences and then combined into a further pool containing 1 representative of each identifying sequence. Thus each final sample for sequencing contained the DNA of 96 different individuals which could be resolved down to a group of 8. Samples were sequenced using Illumina GAIIx platform generating 38-base pair paired-end, indexed reads. Preliminary analysis and quality control of reads was carried out using Illumina’s CASAVA v1.7 software and output files contained all filtered, but unaligned, sequences for each sample stored in a simple exportable FASTQ format. Alignment of short reads against reference messengerRNA sequence (uc003zqq.2, C9orf72, length = 3233) and variations calling was done using Maq 0.7.1 mapping short DNA sequencing reads and calling variants using mapping quality scores with default parameters (Li et al., 2008).

2.8. Sequencing the repeat

Forward (GGTTTAGACGTCGTCGTCGT) and reverse primers (CCAGCTTCGGTCAGAGAAAT) were designed to create a 424-base pair amplicon (64% GC-rich) covering the repeat sequence. A touchdown PCR protocol (see Supplementary Section S2) was used to amplify the region of interest followed by gel electrophoresis to inspect the amplicon length.

Samples were purified and analyzed using an ABI 3130xl Genetic Analyzer (Applied Biosystems) with the BigDye Terminator v1.1 Cycle Sequencing Kit. ABI’s Sequencing Analysis 5.3.1 software was used to analyze base calls and identify repeat patterns.

3. Results

3.1. C9orf72 genotyping

There were 632 case and 4519 control samples. After quality control there were 599 case and 4136 control samples. There were 585,919 SNPs; after quality control 442,057. Thirty-nine cases tested positive for the mutation; the remaining samples had repeat lengths of less than 23 (Supplementary Section 1; Supplementary Table 1).

3.2. Residual association

Genomic control factor \( \hat{\lambda}_{GC} \) was 1.00–1.03 for all analyses. Genome-wide association of all UK case versus control samples confirmed the findings of the original study (Shatunov et al., 2010), with no new associations (Supplementary Fig. 1 and Supplementary Table 2). The top 3 SNPs were rs10967976 (\( p = 6.164 \times 10^{-5} \); odds ratio [OR], 1.41), rs903603 (\( p = 7.548 \times 10^{-8} \); OR, 0.71), and rs10812611 (\( p = 9.819 \times 10^{-6} \); OR, 1.44), all at 9p21.2. Rs903603 was the most associated SNP in the previous independent analysis (Shatunov et al., 2010). In that study rs10967976 and rs10812611 were 1 of 10 most associated SNPs after removal of cases with the mutation, compared with the SNP tagging the repeat mutation, rs3849942, dropping from 7th to 1225th place, as expected (Table 1).

We confirmed that the mutation tagging SNP rs3849942 is strongly associated with ALS when only samples positive for the mutation were tested against controls (\( p = 9.77 \times 10^{-11} \); OR, 5.225; rank = 1), and rs903603 was not as strongly associated as rs3849942 (\( p = 7.73 \times 10^{-2} \); OR, 0.24; rank = 22) (Supplementary Fig. 4). A Q-Q plot (Supplementary Fig. 5) did not show evidence of inflation of the test statistic.

3.3. Residual association and mutation-specific haplotypes

To explore linkage disequilibrium in the region, we phased and imputed SNPs at C9orf72 and the surrounding region (chr9:27303051–27750375) using the 1000 Genomes Project reference panel. Haplotype association analyses and refinement of haplotype blocks in Haplovie identified a 79-SNP haplotype in 100% of the cases with the mutation \( (n = 39) \), of which 72 SNPs were from an 82-SNP haplotype published previously (Smith et al., 2013). Using the same method we also identified a distinct 11-SNP haplotype present in 76% of the samples without the mutation (Supplementary Fig. 6).

3.4. Repeat length and risk allelic distribution

We examined the relationship between the hexanucleotide repeat length of phased cases without the mutation \( (n = 448) \) and the allelic frequency and distribution of ALS-associated SNPs to explore the linkage disequilibrium pattern of the locus. The mutation-tagging SNP rs3849942 risk allele was associated with increased hexanucleotide repeat numbers for the larger allele in linear regression \( (\beta = 5.711; p = 3.02 \times 10^{-84}) \) (Supplementary Table 4). Allele frequency distributions of the repeat length stratified by rs3849942 alleles showed a marked difference between those with 6 or less against 7 or more (Fig. 1). Nonmutation samples with 7 or more repeats \( (n = 208) \) showed the strongest genome-wide association with ALS when compared with control samples \( (n = 4142) \); OR, 4.99; \( p = 1.35 \times 10^{-45} \) (Supplementary Table 5).

A similar relationship between risk alleles and hexanucleotide repeat length was observed for the SNPs showing residual association. Repeat numbers greater than 2 (the minimum length) for the larger allele were significantly associated with repeat alleles at all of the 3 SNPs showing residual association (Fig. 2). Modeling repeat length as a quantitative variable in a linear regression confirmed that these SNPs were significant predictors of having greater than 2 repeats (example SNP rs10812611; \( \beta = 5.835; p = 1.50 \times 10^{-52} \)) (Supplementary Table 4). Again, association testing of samples with repeat length greater than 2 \( (n = 385) \) against all controls \( (n = 4142) \) showed that SNPs showing residual association

<table>
<thead>
<tr>
<th>SNP</th>
<th>With hexanucleotide repeat mutation</th>
<th>Without hexanucleotide repeat mutation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rank</td>
<td>p</td>
<td>Odds ratio (allele)</td>
</tr>
<tr>
<td>rs10967976 First</td>
<td>6.16E-08</td>
<td>1.41 (G)</td>
</tr>
<tr>
<td>rs903603 Second</td>
<td>7.55E-08</td>
<td>0.71 (A)</td>
</tr>
<tr>
<td>rs10812611 Third</td>
<td>9.82E-08</td>
<td>1.4 (G)</td>
</tr>
<tr>
<td>rs3849942 Seventh</td>
<td>3.28E-06</td>
<td>1.39 (A)</td>
</tr>
<tr>
<td>rs2814707 Ninth</td>
<td>5.00E-06</td>
<td>1.38 (A)</td>
</tr>
</tbody>
</table>

Key: SNP, single-nucleotide polymorphism.
were most significantly associated with ALS; rs903603: OR, 0.39; \( p = 2.21 \times 10^{-28} \); rs10812611: OR, 2.78; \( p = 6.96 \times 10^{-33} \); and rs10967976: OR, 2.82; \( p = 1.19 \times 10^{-33} \) (Supplementary Table 6).

The identified 11-SNP haplotype for the residual association SNPs was found in 100% of the samples with greater than 2 repeats for larger allele (71% in control samples). Extension of the haplotype to include an additional SNP, rs2492816, was very specific for longer repeat lengths. This 12-SNP haplotype was strongly associated with ALS in the sample set excluding cases with the mutation, or with 2 or less repeats (\( \chi^2 = 114; \ p = 1.56 \times 10^{-24} \); Fig. 3). This haplotype was not significantly associated with ALS in cases with 2 hexanucleotide repeats and was not found at all in cases with the pathological repeat mutation.

An omnibus association analysis comparing mutation-specific and residual association haplotypes in those with the mutation and those with greater than 2 repeats on their larger allele supported the hypothesis that the alleles showing residual association at C9orf72 are significantly associated with ALS, but not by association with the pathological repeat mutation (Table 2).

3.5. Apparent homozygosity for larger hexanucleotide repeats associates with the risk allele of rs903603

The association of risk alleles with greater than 2 hexanucleotide repeats suggests that residual association SNPs might predispose to instability, and therefore a repeat mutation, as has been postulated previously for the rs3849942 haplotypic background. Thus, 1 possible explanation of the residual association is an interrupted or alternative repeat sequence, also pathologically expanded, but not detected by repeat-primed PCR targeted at (GGGGCC)n (Fig. 4). In that situation one would expect a similar problem to that reported by DeJesus-Hernandez et al. (2011): that cases appear homozygous because their larger repeat allele is too expanded to amplify. Most cases (93.8%; 15/16 cases) with greater than 2 repeats and the rs903603 risk allele were apparently homozygous for the hexanucleotide repeat allele, breaking Hardy–Weinberg equilibrium. In comparison, 66% (150/266) of cases with the same risk allele were heterozygous for the repeat and we found only 4% (3/83) of cases with the risk allele homozygous for 2 repeats (Table 3). We did not observe this >2 repeat homozygosity bias for any other SNPs. For example, 37% of cases with the mutation-specific risk alleles had homozygosity for repeat alleles of length greater than 2, which satisfied Hardy–Weinberg equilibrium.

3.6. Trimodal pattern of repeats

We found a trimodal pattern of repeat allele sizes, where the occurrence of 2, 5, or 8 repeats was significantly more frequent than

![Fig. 1. Relationship between rs3849942 alleles and repeat length in nonmutation cases. The risk allele is strongly associated with longer repeat lengths, suggesting it might lie on a haplotype promoting repeat length instability.](image1)

![Fig. 2. The relationship between hexanucleotide allele repeat length and single-nucleotide polymorphisms (SNPs) showing residual association at the C9orf72 locus SNP rs903603 (A), rs10967976 (B), and rs10812611 (C). Just as for the relationship for rs3849942 shown in Fig. 1, the risk allele for SNPs on the alternative risk haplotype is overwhelmingly likely to be associated with repeat sizes greater than 2.](image2)

![Fig. 3. Twelve single-nucleotide polymorphism haplotypes for case samples with repeat length greater than size 2.](image3)
Table 2
Association analyses of haplotypes with ALS in mutation and case samples with hexanucleotide repeat length >2

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>Mutation p-value for association with ALS</th>
<th>Residual</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mutation cases (n = 39)</td>
<td>7.53 × 10^{-14}</td>
<td>1.30 × 10^{-8}</td>
</tr>
<tr>
<td>Cases with hexanucleotide repeat length &gt;2 (n = 385)</td>
<td>9.87 × 10^{-9}</td>
<td>5.91 × 10^{-23}</td>
</tr>
</tbody>
</table>

Key: ALS, amyotrophic lateral sclerosis.
Values in bold highlight association between mutation groups and haplotypes.

Table 3
Frequency of cases by residual rs10967976 alleles stratified by repeat length and zygosity

<table>
<thead>
<tr>
<th>Repeat zygosity</th>
<th>Nonrisk allele A (%)</th>
<th>Risk allele G (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heterozygous</td>
<td>76 (34)</td>
<td>150 (66)</td>
</tr>
<tr>
<td>Homozygous 2 repeats</td>
<td>80 (96)</td>
<td>3 (4)</td>
</tr>
<tr>
<td>Homozygous (repeats &gt;2)</td>
<td>1 (5)</td>
<td>18 (85)</td>
</tr>
</tbody>
</table>

Because the SNPs showing residual association are not associated with the mutation but are associated with ALS, repeat instability, and apparent homoyzosity for the repeat, 1 possible explanation is an alternative pathological expansion that is not detected by repeat-primed PCR targeting (GGGGCC)n. Such an alternative repeat sequence might be the result of an interrupted repeat. Interrupted repeat sequences are found in ALS resulting from intermediate expansion of a CAG trinucleotide repeat in the ATXN2 gene, with CAA interrupting the sequence and resulting in an alternative (Corrado et al., 2011). Interrupted sequences are also found in Fragile X syndrome, a condition with a massive pathological repeat expansion (Rebczuk et al., 2009).

Others have reported evidence of pathological C9orf72 hexanucleotide expansions using Southern blot analysis without corresponding detection of repeat-primed PCR in FTD cases, consistent with an alternative repeat sequence (Pickering-Brown et al., 2012).

Current evidence supports the idea that the sheer size of the disease-associated C9orf72 hexanucleotide repeat expansion causes pathology, with no evidence for repeat lengths below 23 being associated with ALS. One would expect that an alternative repeat that is also pathological should also be very large and not readily amplified by PCR. In that case, samples carrying the alternative repeat should appear homoyzosity for the nonexpanded allele because the pathological allele would appear null using PCR. Consistent with this, we find cases with more than 2 repeats are overwhelmingly likely to appear homoyzosity.

One factor increasing the likelihood of pathological expansion in transmission to the next generation is the current size of the repeat, and it is particularly interesting that the SNPs showing residual association also strongly associated with alleles showing larger numbers of nonpathological repeats. The C9orf72 region is very GC-rich, known to increase vulnerability toward repeat mutations. Additionally, a familial study examining inheritance of the repeat mutation suggests that larger repeat lengths are indeed more unstable and prone to expansion (Beck et al., 2013).

We found a trimodal pattern of repeat allele sizes with significant departure from a uniform distribution for repeat lengths 2, 5, and 8. This pattern is also apparent in previous publications (Djeunes-Hernandez et al., 2011; Smith et al., 2013), although not explicitly described. The tendency to increase the allele size by

![Fig. 4](image)

**Fig. 4.** Repeat-primed polymerase chain reaction results. Repeat-primed polymerase chain reaction of example case samples showing a small expansion in the nonpathological size range (A), and a sample known to have a pathological (GGGGCC)n expansion mutation (B).
18 nucleotides might be related to the underlying pathological mechanism of expansion, either because the mechanism results in preferential expansion in units of 18 base pairs, or because expansion in other unit sizes is unstable and leads to massive pathological expansion.

A weakness of this study is that we relied on repeat-primed PCR to identify cases with the mutation. It is possible that some cases with the mutation were missed and this explains the residual association. Several factors make this unlikely. First, the mutation is tagged by rs3849942, and as expected, this SNP lost association with ALS after the removal of mutation-positive cases, and rs903603 remained in the top 10 associated SNPs. Second, the frequency of identified mutation cases in our sample is among the highest in the world (Smith et al., 2013), suggesting we are not missing a large proportion of these. Third, we performed a standard PCR of the hexanucleotide repeat and assayed the length polymorphism of the alleles (AFLP); in all cases, there were no heterozygous cases with the expansion, suggesting that the detection method worked. Fourth, in all cases in which sequence data were available, the repeat size on sequencing matched our estimate of size based on AFLP and repeat-primed PCR.

Another explanation for our findings is a substitution or similar mutation, rather than an alternative repeat mutation. This seems unlikely because for several years many groups, including ours, performed sequencing through the region finding no segregating point mutations or indels. Furthermore, the point mutations we identified on sequencing do not occur in individuals carrying the alternative risk haplotype. It does however remain possible that for a small subset of individuals with sporadic ALS, point mutations of C9orf72 are pathogenic, but it is then difficult to explain the association between rs903603 risk alleles and hexanucleotide repeats greater than a length of 2. Nevertheless, we cannot exclude this possibility.

A third possibility is that SNPs showing residual association tag repeat alleles greater than size 2 because such repeats are themselves pathogenic. Case and control samples share the same frequency and distribution of repeat lengths (Rutherford et al., 2012) making this unlikely, but recent evidence suggests that moderate expansion in the number of repeats can actually be pathogenic (Gomez-Tortosa et al., 2013).

We present evidence of residual association at the C9orf72 locus in ALS, not accounted for by the known pathological GGGGCC hexanucleotide repeat expansion but consistent with the hypothesis of an interrupted GGGGCC or alternative sequence repeat. We present a new pathological haplotype associated with ALS at this locus that is associated with having greater than 2 repeats for the larger allele, suggesting this haplotype predisposes to repeat instability. Further investigation including Southern blot analysis of individuals carrying the risk haplotype and sequence analysis of the region will help to identify the cause of the residual association signal we have identified.

Disclosure statement

Ammar Al-Chalabi discloses the following. Personal payments: royalties from the books, The Brain (Oneworld Publications) and Complex Disease Genetics, a Laboratory Manual (Cold Spring Harbor Laboratory Press); institutional payments: consultancy to Biogen Idec and Cytokinetics Inc; grant income: from various charities and governmental organizations. The remaining authors have no conflicts of interest to disclose.

Patients provided ethnically approved consent for withdrawal of blood used for genotyping.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.neurobiolaging.2013.03.003.

References


