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Internalization of GPCRs: implication in receptor function, physiology and diseases

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1 Abstract

2 G protein-coupled receptors (GPCRs) are the largest family of membrane receptors and mediate the 3 effects of numerous hormones and neurotransmitters. The nearly 1,000 GPCRs encoded by the human 4 genome regulate virtually all physiological functions and are implicated in the pathogenesis of prevalent 5 human diseases such as thyroid disorders, hypertension or Parkinson's disease. As a result, 30 to 50% of 6 all currently prescribed drugs are targeting these receptors. Once activated, GPCRs induce signals at the 7 cell surface. This is often followed by internalization, a process that results in the transfer of receptors 8 from the plasma membrane to membranes of the endosomal compartment. Internalization was initially 9 thought to be mainly implicated in signal desensitization, a mechanism of adaptation to prolonged 10 receptor stimulation. However, several unexpected functions have subsequently emerged. Most 11 notably, accumulating evidence indicates that internalization can induce prolonged receptor signaling on 12 intracellular membranes, which is apparently required for at least some biological effects of hormones 13 like TSH, LH and adrenaline. These findings reveal an even stronger connection between receptor 14 internalization and signaling than previously thought. Whereas new studies are just beginning to reveal 15 an important physiological role for GPCR signaling after internalization and ways to exploit it for 16 therapeutic purposes, future investigations will be required to explore its involvement in human disease.

17

18 Keywords

19 GPCR, cAMP, receptor internalization, TSH, PTH, LH, endosomal signaling.

20

21 Abbreviations

22 G protein-coupled receptor (GPCR), protein kinase A (PKA), cyclic adenosine monophosphate (cAMP),

23 mitogen-activated protein kinase (MAPK), thyroid stimulating hormone (TSH), parathyroid hormone

24 (PTH), protein kinase A (PKA), neurokinin (NK), clathrin-mediated endocytosis (CME), clathrin-coated pits

25 (CCPs).

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1 Introduction

G protein-coupled receptors (GPCRs), with a share of almost 4% of the human genome [1], constitute the largest family of receptors that allow cells to sense extracellular stimuli [2, 3]. These external stimuli range from sensory cues like light, odorants and tastants to small-molecule neurotransmitters, peptides and hormones [2, 3]. This high diversity underscores the fundamental role that GPCRs play in the function of the endocrine, nervous, cardiovascular, sensory and immune systems.

The main initial steps of GPCR activation and signaling have been elucidated in detail [2, 4]. These events are initiated by binding of an agonist to a receptor, which triggers a series of conformational changes in the receptor that culminate in its activation. The activated receptor, in turn, binds to and activates heterotrimeric G proteins, which are composed of an α , β and γ subunit and exist in different isoforms. The α and $\beta\gamma$ subunit finally modulate the activity of membrane-localized effectors, including ion channels and enzymes like phospholipase C β (PLC β) and adenylyl cyclase.

13 A classic example of the role of these receptors in physiology is their involvement in the regulation of heart contractility. β -adrenergic receptors located on the surface of cardiomyocytes mediate the 14 15 positive ionotropic and chronotropic effects of adrenalin and noradrenalin, released upon sympathetic activation. Binding of adrenalin or noradrenalin to these receptors, which are coupled to the G_s protein, 16 activate adenylyl cyclases to produce cAMP, which stimulates protein kinase A (PKA). PKA, in turn, 17 phosphorylates different molecules involved in cardiac contractility, including L-type Ca^{2+} channels, 18 phospholamban and troponin I, ultimately leading to enhanced cardiomyocyte contractility [5]. In 19 20 addition, cAMP directly promotes the opening of pacemaker (HCN) channels in the conductive tissue, 21 thus increasing heart rate [6, 7]. Parasympathetic activation counteracts these effects via release of 22 acetylcholine, which binds to muscarinic (M2) receptors coupled to $G_{i/o}$, thus inhibiting adenylyl cyclase 23 activation. In addition, the $\beta\gamma$ subunits released upon G_{i/o} activation stabilize the membrane potential via activating potassium (GIRK) channels in the conductive tissue [8-12]. In the endocrine system, GPCRs 24 25 play an essential role as receptors for several hormones, hypothalamic releasing factors and local 26 modulators. All major known hypothalamic releasing (TRH, GnRH, CRH, GHRH) and inhibiting 27 (somatostatin, dopamine) hormones act via specific GPCRs [13-17]. With the exception of GH and PRL, 28 anterior (TSH, LH, FSH, ACTH, MSH) and posterior (vasopressin, oxytocin) pituitary hormones also signal 29 through activation of GPCRs [18]. For an extensive discussion of the specific roles of GPCRs and G 30 proteins in human physiology we refer the reader to the comprehensive review by Wettschureck and 31 Offermanns [19].

32

1

2 Mechanisms of GPCR internalization

3 Like for other types of receptors, prolonged agonist stimulation often leads to GPCR internalization, 4 which can occur via different pathways [2, 20-23]. Of these pathways, clathrin-mediated endocytosis 5 (CME) is the best characterized and arguably most relevant one (Figure 1) [2, 20-23]. The first molecular 6 event involved in GPCR internalization is the binding of a family of G protein-coupled receptor kinases 7 (GRKs) to an agonist-occupied receptor, which phosphorylate multiple intracellular serine and threonine residues located in the 3rd intracellular loop or at the C-terminus of the receptor [24-27]. This is followed 8 by binding of arrestins to the phosphorylated receptor, which plays a major role in both fast signal 9 10 desensitization and receptor internalization [24, 26]. On the one hand, arrestins compete with G 11 proteins for binding to the receptor, thus leading to signal desensitization. On the other hand, they 12 promote receptor internalization via interacting with key proteins involved in the assembly of clathrin-13 coated pits (CCPs) such as the clathrin heavy chain and the clathrin adaptor protein AP2 [28, 29]. This 14 leads to the recruitment of GPCRs into CCPs, which detach form the plasma membrane in a process that 15 requires the small GTPase dynamin [30]. Receptors are then rapidly transferred to early endosomes, 16 from where they can follow either of two main trafficking pathways [21, 31]. Some GPCRs are sorted out in the endosomal compartment, where they are dephosphorylated, to be then recycled back to the 17 plasma membrane. Others are directed to lysosomes where they are degraded, leading to receptor 18 19 downregulation [24, 26].

20

21 Role of receptor internalization in MAPK signaling

22 While rapid desensitization was shown to occur before receptor internalization and be mediated by 23 receptor phosphorylation and β -arrestin recruitment, it also began to emerge that β -arrestin 24 recruitment and receptor internalization might also exert other functions. In experiments using a 25 dominant-negative dynamin mutant, Daaka et al. showed that receptor internalization is required for efficient ERK activation in response to β_2 -adrenergic receptor stimulation [32]. Subsequently, it was 26 27 shown that β -arrestins can bind several components of mitogen-activated protein kinase (MAPK) pathways [33, 34], thus promoting G protein-independent MAPK signaling. Since some GPCR are found 28 29 on early endosomes in complex with β -arrestins, it has been suggested that these events result in 30 endosomal MAPK signaling (Figure 1) [35]. Intriguingly, the activation of arrestin-bound ERK has been 31 shown to favor cytoplasmic vs. nuclear effects of MAPK activation by preventing ERK translocation to 32 the nucleus [34, 36]. However, the β -arrestin dependent activation of MAPKs can also occur while the

1 receptors are still located on the plasma membrane. Thus, it remains to be clarified what is the relative 2 contribution of cell surface vs. endosomal MAPK signaling. Moreover, some GPCRs that are poorly 3 internalized are nevertheless able to efficiently induce MAPK signaling. This can be at least partially 4 explained by the existence of other mechanisms leading to MAPK activation. Yet another possible 5 explanation for these findings comes from a recent study on the β_1 -adrenergic receptor – which 6 internalizes poorly upon agonist stimulation – indicating that receptor activation can lead to recruitment 7 of β -arrestin to CCPs and MAPK signaling from CCPs in the absence of receptors [37].

8

9 New paradigm of GPCR signaling from intracellular compartments

Although classical, G protein-dependent signaling has long been believed to be restricted to the plasma 10 11 membrane, studies performed in the last ten years have provided strong evidence that internalized 12 GPCRs can continue signaling on intracellular membranes (Figure 1). A first indication came from 13 experiments on the Ste2 receptor, which is implicated in pheromone signaling in yeast [38]. 14 Subsequently, our group and that of Jean-Pierre Vilardaga independently showed that the TSH and PTH 15 receptors induce a persistent phase of cAMP production after internalization, which could be prevented 16 by interfering with CME [39, 40]. Signaling by internalized TSH receptors was shown to differ from the 17 one occurring at the plasma membrane in that it was required for efficient phosphorylation of the 18 vasodilator-stimulated phosphoprotein (VASP) and actin depolymerization in response to TSH, which is 19 involved in thyroglobulin reuptake and, thus, in thyroid hormone release [39]. In the case of the PTH 20 receptor, signaling was shown to be turned off by retromer - which mediates retrograde trafficking 21 from endosomes to the trans-Golgi network - and endosomal acidification [41, 42]. These findings 22 challenged the classical model of GPCR signaling by indicating that G protein signaling can also occur on 23 intracellular membranes. They also pointed to early endosomes, in the case of the PTH receptor, and the Golgi/trans-Golgi network, in the case of the TSH receptor, as likely sites of intracellular GPCR signaling 24 25 (Figure 1).

Further important evidence for G protein signaling on early endosomes has been subsequently obtained for the β_2 -adrenegic receptor using fluorescently-tagged conformation-sensitive nanobodies selectively recognizing the active receptor and G_s protein [43].

29 More recently, our group used a combination of sensors based fluorescence resonance energy transfer 30 (FRET) and a nanobody recognizing the active G_s protein to localize the subcellular compartment where 31 endogenous TSH receptors are signaling in primary thyroid cells [44]. We found that the TSH receptor 32 co-internalizes with TSH and traffics retrogradely to the trans-Golgi network, where it activates an

endogenous pool of G_s protein. This leads to a delayed phase of local cAMP production and PKA 1 2 activation at a critical position near the nucleus, which appears required for efficient CREB 3 phosphorylation and gene transcription in response to TSH [44]. In contrast to previous observations 4 with the PTH receptor, however, retromer was found to promote persistent TSH receptor signaling [44]. 5 A requirement of receptor internalization for gene transcription has also been demonstrated for the β_2 6 adrenergic receptor [45]. Moreover, signaling in the Golgi complex has also been demonstrated for 7 the β_1 -adrenergic receptor [46]. However, in the case of the β_1 -adrenergic receptor, it has been 8 suggested that adrenalin, which is hydrophilic, crosses cellular membranes via the organic cation 9 transporter 3 (OCT3) and reaches a pool of β_1 -adrenergic receptors that reside in the Golgi complex [46]. In the meantime, signaling at intracellular membranes has been reported for several GPCRs, including 10 11 the dopamine D1 receptor [47], vasopressin V2 receptor [48], glucagon-like peptide 1 (GLP1) receptor [49], pituitary adenylate cyclase activating polypeptide 1 (PACAP1) receptor [50] and glucose-dependent 12 insulinotropic peptide (GIP) receptor [51]. 13

A question left open by these studies was related to the apparent contrasting role of β -arrestins, which have a well-established role in signal desensitization and, at the same time, have been suggested to promote endosomal signaling. Intriguingly, recent structural studies indicate that β -arrestins can engage with two different domains of GPCRs, i.e. with either the C-tail or the seven-transmembrane core [52]. Moreover, a complex consisting of a receptor with the G protein bound to its seven-transmembrane core and β -arrestin 1 simultaneously bound to its C-tail has been directly observed by cry-electron microscopy [53].

21 All these studies suggest the existence of multiple intracellular locations for GPCR signaling (Figure 1). 22 Some receptors, like the PTH and the β_2 -adrenergic receptor, seem to signal prevalently from early 23 endosomes. In contrast, the TSH and the β_1 -adrennergic receptor signal on membranes of the 24 Golgi/trans-Golgi network. Furthermore, there is evidence for GPCR signaling at other intracellular 25 compartments such as the nuclear envelope [54] and, more recently, mitochondria. Indeed, cannabinoid 26 CB1 receptors have been shown to be located on brain mitochondrial membranes, where they have 27 been suggested to play a role in the amnesic effects of cannabinoids [55]. Similarly, melatonin has been shown to be produced inside neuronal mitochondria, where it activates local MT1 receptors [56]. The 28 29 resulting signaling prevents stress-mediated cytochrome c release and caspase activation, thus 30 contributing to melatonin neuroprotective effects [56]. Although we are only beginning to understand 31 the implications of such a high degree of spatial control and complexity in GPCR signaling, it is likely that these mechanisms play an important role in allowing to discriminate among the multitude of
 extracellular signals that converge on a single cell.

3

4 Role of receptor internalization and trafficking in physiology and disease

5 Consistent with their crucial role of in GPCR signaling, receptor internalization and trafficking are deeply 6 implicated in human physiology and, most likely, also in disease. A first important aspect regards the 7 correct subcellular localization of receptors. Indeed, genetic mutations affecting receptor trafficking and 8 causing reduced cell surface localization of receptors are known to be implicated in various human 9 diseases, such as TSH resistance, familial idiopathic hypogonadotropic hypogonadism, Leydig cell 10 hypoplasia or familial glucocorticoid deficiency [57].

With the recent demonstration that GPCRs can continue signaling after internalization, GPCR signaling at
 intracellular sites is also emerging as an important aspect of GPCR biology with implications in
 physiology and disease.

14 For the TSH receptor, signaling at the Golgi/trans-Golgi network appears required for both rapid effects 15 of TSH – such as actin depolymerization, which is implicated in thyroglobulin reuptake and, thus, thyroid 16 hormone release – and late ones, such as those on gene transcription. Continued signaling by TSH 17 receptors after internalization might contribute to hyperthyroidism in Grave's disease, where 18 autoantibodies chronically activate the TSH receptor. Moreover, it might play a role in the pathogenesis of toxic thyroid adenomas and congenital/familial non-autoimmune hyperthyroidism, which are caused 19 20 by activating TSH receptor mutations that are often associated with intracellular receptor accumulation 21 [58, 59].

For the PTH receptor, which plays a critical role in regulating Ca^{2+} homeostasis and bone turnover and is 22 a major pharmacological target for the therapy of osteoporosis, it has been shown that PTH₁₋₃₄ but not 23 the PTH related peptide $PTHrP_{1:36}$ – which activates the PTH receptor in a paracrine fashion – is capable 24 25 of inducing persistent cAMP signaling [40]. Moreover, a PTH analog (M-PTH₁₋₃₄) that produces a more sustained cAMP response than PTH₁₋₃₄ has been shown to induce larger increases in trabecular bone 26 27 volume and cortical bone turnover although the responsible mechanisms have not been fully elucidated [60]. Similarly, vasopressin and oxytocin can both induce cAMP/PKA signaling upon binding to the V2 28 29 receptor but only vasopressin leads to a strong antinatriuretic and antidiuretic effect [61-63]. Feinstein et al showed that this difference in signaling strength possibly results from different spatial signaling 30 31 patterns induced by these two ligands [48]. These examples also suggest the possibility of designing 32 GPCR agonists capable of preferentially inducing cell-surface vs. intracellular signaling. This might allow developing a new generation of GPCR agonists with tailored biological effects, and thus, potentially
 improved efficacy and tolerability.

3 More recently, our group took advantage of mice expressing a FRET sensor for cAMP to investigate 4 cAMP signaling in intact ovarian follicles [64]. We found that activation of LH receptors with LH induces 5 two waves of cAMP production that propagate within the follicles. Importantly, blocking receptor 6 internalization prevented the second phase and partially inhibited the LH-induce resumption of meiosis 7 in the oocyte [64]. These data indicate that LH receptor internalization plays an important role in 8 mediating the biological effects of LH. Future studies appear required to further investigate the role of 9 LH receptor signaling at intracellular sites in both female and male reproduction and its alterations in 10 gonadal disorders.

With the growing number of studies investigating GPCR signaling at intracellular sites, the physiological implications of this phenomenon are increasing. These include a role in insulin secretion for the GLP1 receptor [49, 65], in renal water and sodium reuptake for the vasopressin V2 receptor [48] and in the excitability of cardiac neurons for the PACAP1 receptor [50].

Whereas receptor internalization has been mostly associated with prolonged cAMP signaling from intracellular sites, and thus mostly with slow biological effects, in the case of dopamine D1 receptors, it has been shown that these receptors are internalized very rapidly after agonist stimulation (within one minute) and that the resulting cAMP signaling from endosomal membranes increases neuronal excitability in striatal neurons [47].

20 So far, endosomal GPCR signaling has been mostly investigated in cellular models or using ex vivo 21 preparations. Whereas these studies indicate that receptor internalization is required to mediate the 22 biological effects of several hormones and neurotransmitters, further studies are required to investigate 23 these processes in vivo. Interestingly, two recent studies have provided first in vivo evidence for a relevant physiological role of endosomal GPCR signaling. A first study investigated the role of 24 25 internalization of the neurokinin 1 (NK1) receptor, which mediates the effects of substance P, on pain 26 sensing [66]. As a result of pain stimuli, substance P is released from the terminals of primary sensory 27 neurons in the dorsal horn of the spinal cord, where it induces activation and internalization of NK1 28 receptors expressed in second-order neurons [67, 68]. The results of the study indicate that inhibiting 29 NK1 internalization and the resulting endosomal signaling attenuate nociception in vivo. This study also 30 reports an innovative pharmacological strategy to selectively inhibit receptor endosomal signaling. For 31 this purpose, the authors developed a cholestenol-conjugated antagonist, which accumulates in endosomes and is capable of inhibiting endosomal NK1 receptor signaling – which is required for 32

nociception – without affecting NK1 receptors at the cell surface. Similar results were obtained by the
 same group for the calcitonin receptor-like receptor, which binds the calcitonin-gene related peptide

(CGRP), and is also implicated in pain transmission [69].

4 Altogether, these new findings reinforce the view that receptor internalization and signaling are 5 inextricably linked and cooperate to mediate the effects of several hormones and neurotransmitters. While genetic defects in receptor trafficking have been associated with selected human diseases and we 6 7 are beginning to explore the physiological implications of new exciting discoveries in this field, further studies are needed to investigate the involvement of receptor internalization and signaling at 8 9 intracellular sites in a large repertoire of diseases. Furthermore, there is an urgent need in drug 10 development to move away from oversimplified models of GPCR signaling to take into account the complex interplay between signaling and internalization. This might allow going far beyond the concept 11 12 of either activating or inhibiting a receptor - on which current drugs are based - and design more 13 selective drugs capable of modulating receptor signaling at the desired time and subcellular location. 14 The clinician should keep an eye on these exciting developments, which might revolutionize the way of 15 treating common diseases in the near future.

16

3

17 Author contributions

18 A.G. and D.C. wrote the manuscript.

19

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25

26 Conflicts of interests

- 27 The authors declare no competing financial interests.
- 28

29 Practice points

GPCRs are the largest family of receptors and mediate the effects of several hormones and
 neurotransmitters

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- 1 GPCRs are major pharmacological targets (at least 30% of all drugs on the market target these 2 receptors) 3 Prolonged stimulation with hormones or drugs leads to GPCR internalization 4 Receptor internalization serves different functions and has been unexpectedly shown to be 5 required for the biological effects of hormones and neurotransmitters Defects in receptor trafficking are involved in some genetic disorders and their involvement in 6 7 common diseases needs to be further explored. 8 The new finding that GPCRs signal not only at the plasma membrane but also on membranes of 9 endosomes and the Golgi/trans-Golgi network might allow to develop a new generation of drugs 10 with improved efficacy and less side effects. 11 12 **Research agenda** Further explore the role of GPCR internalization in human physiology. 13 14 Investigate the involvement of receptor internalization and GPCR signaling on intracellular 15 membranes in the pathogenesis of human diseases where GPCRs play an important role. 16 Develop new drugs capable of selectively activating or inhibiting GPCR at the cell surface vs. at • 17 intracellular sites or to modify GPCR internalization and/or intracellular trafficking. 18 19 **Figure legend** 20 Figure 1: The complex interply between GPCR signaling and innternalization. Binding of a ligand to a receptor (1) induces a first phase of G protein-dependent signaling at the plasma membrane (2). This is 21 22 followed by GRK-mediated phoshorylation of the receptor and β -arrestin binding, which results in rapid desensitization. At the same time, β -arrestin promotes MAPK signaling (3). β -arrestin also induces 23 24 receptor internalization via clathrin-mediated endoctosis (CME). The internalized receptor can induce a 25 second phase of G protein-dependent signaling from either early endosomes or the Golgi/trans-Golgi 26 network (4). This second signaling phase has been shown to be biologically relevant for a growing
- number of GPCRs. Afterwards, the receptor is either degraded in lysosomes or recycled back to the
 plasma membrane (5) to undergo another round of signaling.

1 References

- 2 [1] Bjarnadottir TK, Gloriam DE, Hellstrand SH, Kristiansson H, Fredriksson R, Schioth HB. Comprehensive
- 3 repertoire and phylogenetic analysis of the G protein-coupled receptors in human and mouse.
- 4 Genomics. 2006;88:263-73.
- 5 *[2] Pierce KL, Premont RT, Lefkowitz RJ. Seven-transmembrane receptors. Nat Rev Mol Cell Biol.
- 6 2002;3:639-50.
- 7 [3] Lefkowitz RJ. Historical review: a brief history and personal retrospective of seven-transmembrane
- 8 receptors. Trends Pharmacol Sci. 2004;25:413-22.
- 9 [4] Lefkowitz RJ. A brief history of G-protein coupled receptors (Nobel Lecture). Angew Chem Int Ed Engl.
- 10 2013;52:6366-78.
- 11 [5] Bers DM. Cardiac excitation-contraction coupling. Nature. 2002;415:198-205.
- 12 [6] Baruscotti M, Bucchi A, Difrancesco D. Physiology and pharmacology of the cardiac pacemaker
- 13 ("funny") current. Pharmacol Ther. 2005;107:59-79.
- [7] DiFrancesco D, Tortora P. Direct activation of cardiac pacemaker channels by intracellular cyclic AMP.
 Nature. 1991;351:145-7.
- 16 [8] Huang CL, Slesinger PA, Casey PJ, Jan YN, Jan LY. Evidence that direct binding of $G\beta\gamma$ to the GIRK1 G
- protein-gated inwardly rectifying K⁺ channel is important for channel activation. Neuron. 1995;15:113343.
- 19 [9] Inanobe A, Morishige KI, Takahashi N, Ito H, Yamada M, Takumi T, et al. G beta gamma directly binds
- to the carboxyl terminus of the G protein-gated muscarinic K⁺ channel, GIRK1. Biochem Biophys Res
 Commun. 1995;212:1022-8.
- [10] Reuveny E, Slesinger PA, Inglese J, Morales JM, Iniguez-Lluhi JA, Lefkowitz RJ, et al. Activation of the
- cloned muscarinic potassium channel by G protein $\beta\gamma$ subunits. Nature. 1994;370:143-6.
- 24 [11] Wickman KD, Iniguez-Lluhl JA, Davenport PA, Taussig R, Krapivinsky GB, Linder ME, et al.
- 25 Recombinant G-protein $\beta\gamma$ -subunits activate the muscarinic-gated atrial potassium channel. Nature.
- 26 1994;368:255-7.
- 27 [12] Logothetis DE, Kurachi Y, Galper J, Neer EJ, Clapham DE. The $\beta\gamma$ subunits of GTP-binding proteins
- 28 activate the muscarinic K^+ channel in heart. Nature. 1987;325:321-6.
- 29 [13] Hillhouse EW, Randeva H, Ladds G, Grammatopoulos D. Corticotropin-releasing hormone receptors.
- 30 Biochem Soc Trans. 2002;30:428-32.
- 31 [14] Smith RG, Van der Ploeg LH, Howard AD, Feighner SD, Cheng K, Hickey GJ, et al. Peptidomimetic
- regulation of growth hormone secretion. Endocr Rev. 1997;18:621-45.
- 33 [15] Yu R, Hinkle PM. Signal transduction and hormone-dependent internalization of the thyrotropin-
- releasing hormone receptor in cells lacking G_q and G_{11} . J Biol Chem. 1999;274:15745-50.
- 35 [16] Millar RP, Lu ZL, Pawson AJ, Flanagan CA, Morgan K, Maudsley SR. Gonadotropin-releasing hormone
- 36 receptors. Endocr Rev. 2004;25:235-75.
- 37 [17] Gershengorn MC, Osman R. Molecular and cellular biology of thyrotropin-releasing hormone
- 38 receptors. Physiol Rev. 1996;76:175-91.
- [18] Paschke R, Ludgate M. The thyrotropin receptor in thyroid diseases. N Engl J Med. 1997;337:167581.
- [19] Wettschureck N, Offermanns S. Mammalian G proteins and their cell type specific functions. Physiol
 Rev. 2005;85:1159-204.
- 43 [20] Hanyaloglu AC, von Zastrow M. Regulation of GPCRs by endocytic membrane trafficking and its
- 44 potential implications. Annu Rev Pharmacol Toxicol. 2008;48:537-68.
- [21] Sorkin A, von Zastrow M. Endocytosis and signalling: intertwining molecular networks. Nat Rev Mol
 Cell Biol. 2009;10:609-22.

- 1 [22] Drake MT, Shenoy SK, Lefkowitz RJ. Trafficking of G protein-coupled receptors. Circ Res.
- 2 2006;99:570-82.
- 3 [23] DeWire SM, Ahn S, Lefkowitz RJ, Shenoy SK. β-arrestins and cell signaling. Annu Rev Physiol.
- 4 2007;69:483-510.
- 5 [24] Pitcher JA, Freedman NJ, Lefkowitz RJ. G protein-coupled receptor kinases. Annu Rev Biochem.
- 6 1998;67:653-92.
- 7 [25] Pitcher J, Lohse MJ, Codina J, Caron MG, Lefkowitz RJ. Desensitization of the isolated β_2 -adrenergic
- 8 receptor by beta-adrenergic receptor kinase, cAMP-dependent protein kinase, and protein kinase C
- 9 occurs via distinct molecular mechanisms. Biochemistry. 1992;31:3193-7.
- 10 [26] Krupnick JG, Benovic JL. The role of receptor kinases and arrestins in G protein-coupled receptor
- 11 regulation. Annu Rev Pharmacol Toxicol. 1998;38:289-319.
- 12 [27] Stadel JM, Nambi P, Shorr RG, Sawyer DF, Caron MG, Lefkowitz RJ. Catecholamine-induced
- 13 desensitization of turkey erythrocyte adenylate cyclase is associated with phosphorylation of the β -
- 14 adrenergic receptor. Proc Natl Acad Sci U S A. 1983;80:3173-7.
- 15 [28] Laporte SA, Oakley RH, Zhang J, Holt JA, Ferguson SS, Caron MG, et al. The β_2 -adrenergic
- 16 receptor/ β -arrestin complex recruits the clathrin adaptor AP-2 during endocytosis. Proc Natl Acad Sci U
- 17 S A. 1999;96:3712-7.
- 18 [29] Goodman OB, Jr., Krupnick JG, Santini F, Gurevich VV, Penn RB, Gagnon AW, et al. β-arrestin acts as
- a clathrin adaptor in endocytosis of the β_2 -adrenergic receptor. Nature. 1996;383:447-50.
- 20 [30] Doherty GJ, McMahon HT. Mechanisms of endocytosis. Annu Rev Biochem. 2009;78:857-902.
- 21 [31] Irannejad R, Tsvetanova NG, Lobingier BT, von Zastrow M. Effects of endocytosis on receptor-
- 22 mediated signaling. Curr Opin Cell Biol. 2015;35:137-43.
- 23 *[32] Daaka Y, Luttrell LM, Ahn S, Della Rocca GJ, Ferguson SS, Caron MG, et al. Essential role for G
- protein-coupled receptor endocytosis in the activation of mitogen-activated protein kinase. J Biol Chem.
 1998;273:685-8.
- 26 [33] McDonald PH, Chow CW, Miller WE, Laporte SA, Field ME, Lin FT, et al. β-arrestin 2: a receptor-
- 27 regulated MAPK scaffold for the activation of JNK3. Science. 2000;290:1574-7.
- 28 [34] DeFea KA, Zalevsky J, Thoma MS, Dery O, Mullins RD, Bunnett NW. β-arrestin-dependent
- endocytosis of proteinase-activated receptor 2 is required for intracellular targeting of activated ERK1/2.
 J Cell Biol. 2000;148:1267-81.
- 31 [35] Charest PG, Oligny-Longpre G, Bonin H, Azzi M, Bouvier M. The V2 vasopressin receptor stimulates
- 32 ERK1/2 activity independently of heterotrimeric G protein signalling. Cell Signal. 2007;19:32-41.
- 33 [36] Tohgo A, Pierce KL, Choy EW, Lefkowitz RJ, Luttrell LM. β-arrestin scaffolding of the ERK cascade
- 34 enhances cytosolic ERK activity but inhibits ERK-mediated transcription following angiotensin AT1a
- 35 receptor stimulation. J Biol Chem. 2002;277:9429-36.
- 36 [37] Eichel K, Jullie D, von Zastrow M. β -arrestin drives MAP kinase signalling from clathrin-coated
- 37 structures after GPCR dissociation. Nat Cell Biol. 2016;18:303-10.
- 38 [38] Slessareva JE, Routt SM, Temple B, Bankaitis VA, Dohlman HG. Activation of the
- 39 phosphatidylinositol 3-kinase Vps34 by a G protein α subunit at the endosome. Cell. 2006;126:191-203.
- 40 *[39] Calebiro D, Nikolaev VO, Gagliani MC, de Filippis T, Dees C, Tacchetti C, et al. Persistent cAMP-
- 41 signals triggered by internalized G-protein-coupled receptors. PLoS Biol. 2009;7:e1000172.
- 42 *[40] Ferrandon S, Feinstein TN, Castro M, Wang B, Bouley R, Potts JT, et al. Sustained cyclic AMP
- 43 production by parathyroid hormone receptor endocytosis. Nat Chem Biol. 2009;5:734-42.
- 44 [41] Feinstein TN, Wehbi VL, Ardura JA, Wheeler DS, Ferrandon S, Gardella TJ, et al. Retromer terminates
- 45 the generation of cAMP by internalized PTH receptors. Nat Chem Biol. 2011;7:278-84.

- 1 [42] Gidon A, Al-Bataineh MM, Jean-Alphonse FG, Stevenson HP, Watanabe T, Louet C, et al. Endosomal
- 2 GPCR signaling turned off by negative feedback actions of PKA and v-ATPase. Nat Chem Biol.
- 3 2014;10:707-9.
- *[43] Irannejad R, Tomshine JC, Tomshine JR, Chevalier M, Mahoney JP, Steyaert J, et al. Conformational
 biosensors reveal GPCR signalling from endosomes. Nature. 2013;495:534-8.
- 6 *[44] Godbole A, Lyga S, Lohse MJ, Calebiro D. Internalized TSH receptors en route to the TGN induce
- 7 local Gs-protein signaling and gene transcription. Nature communications. 2017;8:443.
- 8 *[45] Tsvetanova NG, von Zastrow M. Spatial encoding of cyclic AMP signaling specificity by GPCR
- 9 endocytosis. Nat Chem Biol. 2014;10:1061-5.
- *[46] Irannejad R, Pessino V, Mika D, Huang B, Wedegaertner PB, Conti M, et al. Functional selectivity of
 GPCR-directed drug action through location bias. Nat Chem Biol. 2017;13:799-806.
- 12 [47] Kotowski SJ, Hopf FW, Seif T, Bonci A, von Zastrow M. Endocytosis promotes rapid dopaminergic 13 signaling. Neuron. 2011;71:278-90.
- 14 [48] Feinstein TN, Yui N, Webber MJ, Wehbi VL, Stevenson HP, King JD, Jr., et al. Noncanonical control of
- 15 vasopressin receptor type 2 signaling by retromer and arrestin. J Biol Chem. 2013;288:27849-60.
- 16 [49] Kuna RS, Girada SB, Asalla S, Vallentyne J, Maddika S, Patterson JT, et al. Glucagon-like peptide-1
- 17 receptor-mediated endosomal cAMP generation promotes glucose-stimulated insulin secretion in
- 18 pancreatic β -cells. Am J Physiol Endocrinol Metab. 2013;305:E161-70.
- 19 [50] Merriam LA, Baran CN, Girard BM, Hardwick JC, May V, Parsons RL. Pituitary adenylate cyclase 1
- 20 receptor internalization and endosomal signaling mediate the pituitary adenylate cyclase activating
- 21 polypeptide-induced increase in guinea pig cardiac neuron excitability. J Neurosci. 2013;33:4614-22.
- 22 [51] Ismail S, Gherardi MJ, Froese A, Zanoun M, Gigoux V, Clerc P, et al. Internalized Receptor for
- Glucose-dependent Insulinotropic Peptide stimulates adenylyl cyclase on early endosomes. Biochem
 Pharmacol. 2016;120:33-45.
- *[52] Shukla AK, Westfield GH, Xiao K, Reis RI, Huang L-Y, Tripathi-Shukla P, et al. Visualization of arrestin
 recruitment by a G-protein-coupled receptor. Nature. 2014;512:218-22.
- 27 [53] Thomsen AR, Plouffe B, Cahill TJ, 3rd, Shukla AK, Tarrasch JT, Dosey AM, et al. GPCR-G Protein-β-
- Arrestin Super-Complex Mediates Sustained G Protein Signaling. Cell. 2016;166:907-19.
- 29 [54] Tadevosyan A, Vaniotis G, Allen BG, Hebert TE, Nattel S. G protein-coupled receptor signalling in the
- 30 cardiac nuclear membrane: evidence and possible roles in physiological and pathophysiological function.
- 31 J Physiol. 2012;590:1313-30.
- 32 [55] Hebert-Chatelain E, Desprez T, Serrat R, Bellocchio L, Soria-Gomez E, Busquets-Garcia A, et al. A
- cannabinoid link between mitochondria and memory. Nature. 2016;539:555-9.
- 34 [56] Suofu Y, Li W, Jean-Alphonse FG, Jia J, Khattar NK, Li J, et al. Dual role of mitochondria in producing
- 35 melatonin and driving GPCR signaling to block cytochrome c release. Proc Natl Acad Sci U S A.
- 36 2017;114:E7997-E8006.
- 37 [57] Lania AG, Mantovani G, Spada A. Mechanisms of disease: Mutations of G proteins and G-protein-
- 38 coupled receptors in endocrine diseases. Nat Clin Pract Endocrinol Metab. 2006;2:681-93.
- 39 [58] Van Sande J, Parma J, Tonacchera M, Swillens S, Dumont J, Vassart G. Somatic and germline
- 40 mutations of the TSH receptor gene in thyroid diseases. J Clin Endocrinol Metab. 1995;80:2577-85.
- 41 [59] Alberti L, Proverbio MC, Costagliola S, Weber G, Beck-Peccoz P, Chiumello G, et al. A novel germline
- 42 mutation in the TSH receptor gene causes non-autoimmune autosomal dominant hyperthyroidism. Eur J
- 43 Endocrinol. 2001;145:249-54.
- 44 [60] Okazaki M, Ferrandon S, Vilardaga JP, Bouxsein ML, Potts JT, Jr., Gardella TJ. Prolonged signaling at
- 45 the parathyroid hormone receptor by peptide ligands targeted to a specific receptor conformation. Proc
- 46 Natl Acad Sci U S A. 2008;105:16525-30.
- 47 [61] Chou CL, DiGiovanni SR, Mejia R, Nielsen S, Knepper MA. Oxytocin as an antidiuretic hormone. I.
- 48 Concentration dependence of action. Am J Physiol. 1995;269:F70-7.

- 1 [62] Chou CL, DiGiovanni SR, Luther A, Lolait SJ, Knepper MA. Oxytocin as an antidiuretic hormone. II.
- 2 Role of V2 vasopressin receptor. Am J Physiol. 1995;269:F78-85.
- 3 [63] Balment RJ, Brimble MJ, Forsling ML, Kelly LP, Musabayane CT. A synergistic effect of oxytocin and
- 4 vasopressin on sodium excretion in the neurohypophysectomized rat. J Physiol. 1986;381:453-64.
- 5 [64] Lyga S, Volpe S, Werthmann RC, Gotz K, Sungkaworn T, Lohse MJ, et al. Persistent cAMP signaling by
- 6 internalized LH receptors in ovarian follicles. Endocrinology. 2016:en20151945.
- 7 [65] Girada SB, Kuna RS, Bele S, Zhu Z, Chakravarthi NR, DiMarchi RD, et al. Galphas regulates Glucagon-
- 8 Like Peptide 1 Receptor-mediated cyclic AMP generation at Rab5 endosomal compartment. Mol Metab. 9 2017;6:1173-85.
- 10 *[66] Jensen DD, Lieu T, Halls ML, Veldhuis NA, Imlach WL, Mai QN, et al. Neurokinin 1 receptor signaling
- 11 in endosomes mediates sustained nociception and is a viable therapeutic target for prolonged pain
- 12 relief. Science translational medicine. 2017;9.
- 13 [67] Mantyh PW, DeMaster E, Malhotra A, Ghilardi JR, Rogers SD, Mantyh CR, et al. Receptor
- 14 endocytosis and dendrite reshaping in spinal neurons after somatosensory stimulation. Science.
- 15 1995;268:1629-32.
- 16 [68] Steinhoff MS, von Mentzer B, Geppetti P, Pothoulakis C, Bunnett NW. Tachykinins and their
- 17 receptors: contributions to physiological control and the mechanisms of disease. Physiol Rev.
- 18 2014;94:265-301.
- 19 [69] Yarwood RE, Imlach WL, Lieu T, Veldhuis NA, Jensen DD, Klein Herenbrink C, et al. Endosomal
- 20 signaling of the receptor for calcitonin gene-related peptide mediates pain transmission. Proc Natl Acad
- 21 Sci U S A. 2017.

