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RESEARCH ARTICLE

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DKK1 expression by synovial fibroblasts in very early rheumatoid arthritis associates with lymphocyte adhesion in an in vitro flow co-culture system

Maria Juarez^{1†}, Helen M. McGettrick^{1†}, Dagmar Scheel-Toellner¹, Lorraine Yeo¹, Julia Spengler¹, Banesa de Paz¹, Rowan Hardy², Mark Cooper³, Karim Raza^{1,4}, Christopher D. Buckley^{1,4} and Andrew Filer^{1,5*}

Abstract

Background: Synovial fibroblasts play a key role in joint destruction and regulation of the inflammatory infiltrate in established rheumatoid arthritis (RA). The mechanisms by which this occurs in the earliest stages of RA are largely unknown. We investigated the role of Dickkopf-related protein 1 (DKK1) produced by synovial fibroblasts of patients with very early rheumatoid arthritis (VeRA).

Methods: Fibroblasts were isolated from the disease-modifying anti-rheumatic drug-naïve Birmingham early arthritis cohort of patients with new onset of clinically apparent arthritis and inflammatory symptoms of ≤ 12 weeks' duration, who at follow-up had either resolving arthritis or RA. Endothelial fibroblast co-cultures were formed using porous filters, and lymphocyte adhesion to co-cultures was assessed using phase-contrast microscopy. DKK1 gene expression and secretion were quantified by quantitative polymerase chain reaction and enzyme-linked immunosorbent assay, respectively.

Results: Synovial fibroblasts from patients with VeRA expressed significantly higher levels of DKK1 messenger RNA than those from patients with resolving arthritis. A similar trend was observed for DKK1 protein secretion. In co-culture constructs, more DKK1 tended to be secreted in co-cultures incorporating fibroblasts from VeRA than in co-cultures from non-inflamed joints and resolving arthritis. DKK1 secretion during co-culture positively correlated with lymphocyte adhesion.

Conclusions: Alterations in DKK1 could be involved in the pathogenesis and perpetuation of the inflammatory response in the earliest clinically apparent stages of RA.

Keywords: DKK1, Synovial fibroblasts, Early inflammatory arthritis, Lymphocyte adhesion

Background

Clinical studies have shown that early aggressive treatment of rheumatoid arthritis (RA) leads to better clinical outcomes without unacceptably high adverse event profiles [1]. The concept of an early window of opportunity is further supported by evidence of a distinct and transient

cytokine profile in the synovial fluid of patients with RA of less than 3 months' duration [2]. However, disease mechanisms during this very early stage of RA development remain largely undefined.

In established RA, multiple epigenetic changes drive the acquisition of a pathogenic phenotype in synovial fibroblasts that underpins their aberrant behaviour [3, 4]. This pathogenic phenotype is evidenced by the ability to invade human cartilage in a severe combined immunodeficiency model of arthritis [5, 6]. Additionally, rheumatoid synovial fibroblasts secrete proteases (e.g., matrix metalloproteinases and cathepsins) that degrade cartilage and bone tissue [7, 8]. Furthermore, they alter the dynamics of bone repair

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by releasing receptor activator of nuclear factor κ B ligand (RANKL), causing osteoclast differentiation leading to bone erosion [9] whilst simultaneously producing Dickkopf-related protein 1 (DKK1) to inhibit osteoblast-driven repair of these erosions [10].

DKK1 is an inhibitor of the Wnt signalling pathway that has been proposed as a master regulator of joint remodelling [10]. Serum levels of DKK1 positively correlate with joint erosions and inflammation in RA [11]. Indeed, patients with a genetic variant of DKK1 which results in higher DKK1 serum levels have more progressive joint destruction [12], suggesting a fundamental role for DKK1 in the pathogenesis of RA. Treatment with antibodies against DKK1 has restored bone loss in murine models of arthritis [10], suggesting it has promise as a novel therapeutic target. In this study, to determine the role of DKK1 in the pathogenesis of early RA, we analysed, for the first time to our knowledge, the expression of DKK1 in synovial fibroblasts from patients with early arthritis that eventually resolved compared with patients whose arthritis developed into RA.

Methods

Patients

Synovial tissue samples were obtained by ultrasound-guided biopsy [13, 14] from patients recruited into the Birmingham early arthritis cohort (BEACON). BEACON is a cohort of disease-modifying anti-rheumatic drug (DMARD)-naive patients with clinically apparent synovitis in at least one joint and inflammatory joint symptoms (morning stiffness and/or inflammatory joint pain and/or swelling) of ≤ 12 weeks' duration. Patients were assigned to one of two outcome categories at 18-month follow-up: resolving arthritis ($n = 11$) or RA ($n = 14$) (fulfilling 2010 American College of Rheumatology/European League Against Rheumatism [ACR/EULAR] criteria) [15]. Patients in the RA group are henceforth referred to as patients with very early rheumatoid arthritis (VeRA). Resolving arthritis was defined as no clinically apparent joint swelling with no DMARD or steroid use in the previous 3 months. In addition, healthy synovial tissue samples were collected from subjects undergoing exploratory arthroscopy for unexplained joint pain with no macro- or microscopic evidence of inflammation. Synovial fibroblasts were isolated as previously described [16] and used between passages 3 and 6 [17]. Demographic and clinical parameters, including age, sex, symptom duration, tender and swollen joint counts, erythrocyte sedimentation rate, C-reactive protein, rheumatoid factor and anti-cyclic citrullinated peptide status, were recorded.

Quantification of DKK1

Fibroblast messenger RNA (mRNA) was extracted using the RNeasy Mini Kit (QIAGEN, Manchester, UK), and

DKK1 expression was analysed by Applied Biosystems TaqMan low-density array (Life Technologies, Paisley, UK) using an Applied Biosystems 7900HT real-time polymerase chain reaction machine (Life Technologies). DKK1 levels were expressed relative to GAPDH using the $2^{-\Delta\Delta C_t}$ method. DKK1 levels in serum and culture supernatants were quantified using the VersaMAP immunoassay or the DKK1 DuoSet enzyme-linked immunosorbent assay (R&D Systems, Abingdon, UK).

Flow-based adhesion assay

Endothelial cells were isolated from umbilical cords as described elsewhere [18]. Endothelial fibroblast co-cultures were established on opposite sides of 0.4- μ m-pore Transwell inserts (BD Biosciences, Oxford, UK) [17] for 24 h before treatment with 100 U/ml tumour necrosis factor- α (R&D Systems) and 10 ng/ml interferon- γ (PeproTech, London, UK) for a further 24 h. Filters were incorporated into a parallel plate chamber, and purified peripheral blood lymphocytes (2×10^6 cells/ml) were perfused for 4 minutes as described elsewhere [17]. Digitised recordings were made of five random fields and analysed using Image-Pro Plus software (Media Cybernetics, Cambridge, UK). The number of adherent lymphocytes was averaged per field and expressed per square millimetre per 10^6 cells perfused (Fig. 2b) [17].

Ethical approval

All human samples were obtained with written informed consent and approval from the Human Biomaterial Resource Centre (Birmingham, UK), West Midlands and Black Country Research Ethics Committee or University of Birmingham Local Ethical Review Committee.

Statistical analysis

Data analyses were performed using PASW 20.0 (SPSS, Chicago, IL, USA) and Prism 5 (GraphPad Software, La Jolla, CA, USA) software. Data were expressed as percentages, mean \pm standard error of the mean and median (interquartile range) as appropriate. Two-group comparisons were performed using the χ^2 test, unpaired t test and Mann-Whitney U test for categorical, parametric continuous and non-parametric continuous data, respectively. Three-group comparisons were performed with the Kruskal-Wallis test and Dunn's post-test. Correlations were performed using Spearman's test, where the r value relates to the non-parametric Spearman's correlation coefficient. p Values less than 0.05 were considered statistically significant.

Results

The demographic and clinical characteristics of patients in both outcome groups are shown in Table 1. Patients in the VeRA group were older, and a significantly higher

Table 1 Demographic, clinical and laboratory characteristics of patients in each outcome group

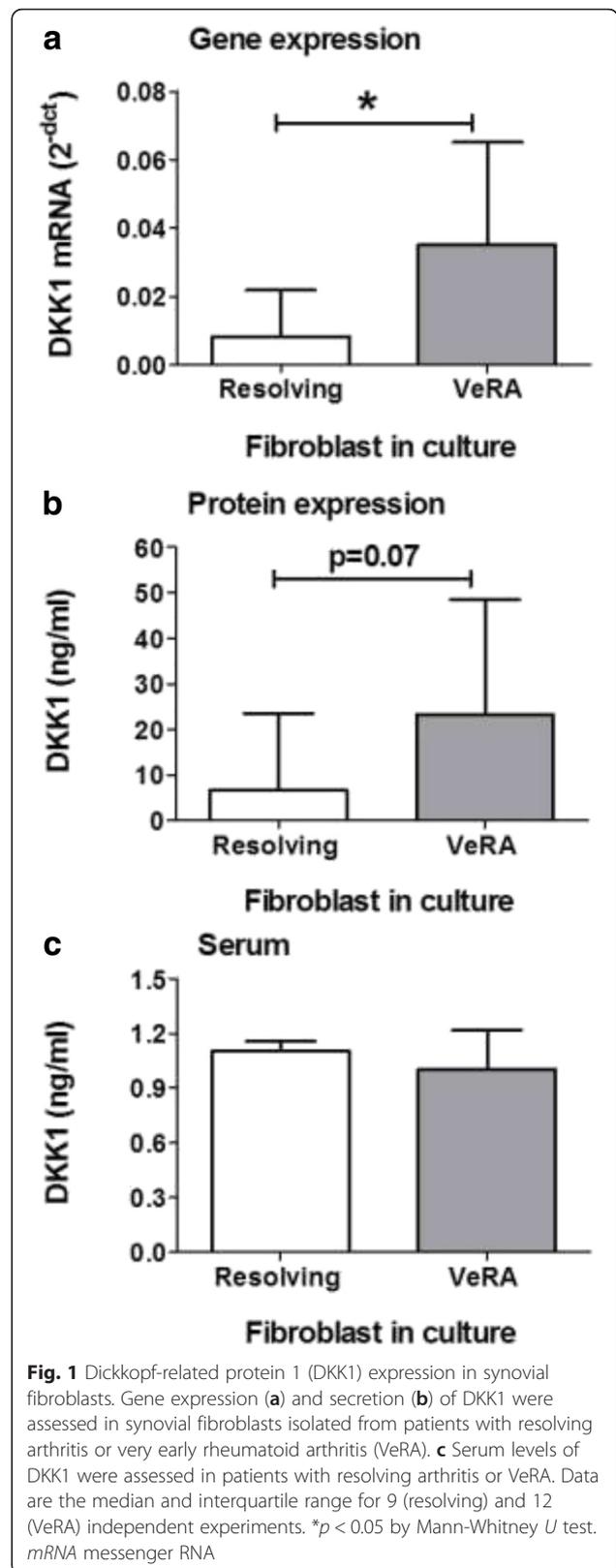
	Resolving arthritis (n = 11)	VeRA (n = 14)	p Value
Age, yr	37.4 ± 10.3	52.9 ± 9.5	0.001
Female, n (%)	3 (27.3)	9 (64.3)	<0.0001
Disease duration, wk	4.4 ± 2.9	6.1 ± 3.3	0.210
CCP-positive, n (%)	0 (0)	7 (50)	<0.0001
RF-positive, n (%)	0 (0)	6 (42.9)	<0.0001
TJC28	2.6 ± 2.0	7.9 ± 5.8	0.008
SJC28	2.7 ± 2.0	7.3 ± 5.3	0.011
CRP	8.4 ± 8.1	23.0 ± 27.0	0.097
ESR	16.4 ± 15.5	31.1 ± 22.4	0.075
VAS	42.3 ± 32.1	46.9 ± 28.1	0.703
DAS28-ESR	3.5 ± 1.0	4.9 ± 1.2	0.004
Radiographic erosions, n (%)	0 (0)	1 (7.1)	<0.0001

VeRA very early rheumatoid arthritis, CCP cyclic citrullinated peptide, RF rheumatoid factor, TJC28 28-joint tender joint count, SJC28 28-joint swollen joint count, CRP C-reactive protein, ESR erythrocyte sedimentation rate, DAS28-ESR Disease Activity Score in 28 joints based on erythrocyte sedimentation rate, VAS visual analogue scale
Data are presented as mean ± standard deviation unless otherwise indicated

number of them were female, compared with patients in the resolving group (9 vs. 3, respectively; $p < 0.001$). Patients with VeRA had more severe disease activity, as evidenced by higher tender joint counts (7.9 vs. 2.6, $p = 0.02$) and swollen joint counts (7.3 vs. 3.0, $p = 0.04$), higher disease activity scores (Disease Activity Score in 28 joints based on erythrocyte sedimentation rate 4.9 vs. 3.5, $p = 0.004$) and higher number of radiographic erosions at baseline (1 vs. 0, $p < 0.001$) than the patients in the resolving group. Seven of the fourteen patients in the VeRA group were undifferentiated at the time of biopsy (ACR/EULAR 2010 score <6). Fibroblasts from two male and two female individuals undergoing exploratory arthroscopy for unexplained joint pain with no macro- or microscopic evidence of inflammation were included as controls for co-culture recruitment assays (mean age 45.8 years, standard deviation 10.3).

Synovial fibroblasts from patients with VeRA expressed significantly higher levels of DKK1 mRNA compared with those with resolving arthritis (Fig. 1a). A similar trend was observed for DKK1 secretion (Fig. 1b). Expression of DKK1 mRNA did not correlate with age, disease duration or any other clinical indices. There was no difference in DKK1 mRNA expression between patients with VeRA presenting with undifferentiated arthritis and those fulfilling criteria at presentation (data not shown). In contrast, serum levels of DKK1 were similar between the clinical outcome groups (Fig. 1c).

Next we assessed whether the differential expression of DKK1 in early disease had functional consequences. Endothelial fibroblast co-cultures were analysed for



DKK1 secretion and for their ability to support lymphocyte adhesion from flow (Fig. 2b). In this model, more DKK1 was secreted in co-cultures incorporating fibroblasts from patients with VeRA compared with those from non-inflamed joints (normal) or patients with resolving disease, although this was not statistically significant (Fig. 2a). VeRA fibroblast co-cultures released significantly higher amounts of DKK1 than endothelial cells cultured alone (Fig. 2a). DKK1 secretion during co-culture showed a positive correlation with the level of lymphocyte adhesion supported by co-culture (Fig. 2c).

Discussion

In this study, we examined, for the first time to our knowledge, the expression of DKK1 in synovial fibroblasts isolated from DMARD-naive patients with inflammatory arthritis of less than 3 months’ duration. Our data suggest that, even at this very early stage, fibroblasts from patients with VeRA may have gained the capacity to impair bone repair and induce bone erosion through increased expression of DKK1. Notably, this pathogenic phenotype was

not observed in fibroblasts from patients with resolving arthritis where non-RA joint inflammation was also present. We also observed an association between DKK1 expression and the ability of fibroblasts from patients with VeRA to perpetuate the inflammatory response through lymphocyte adhesion. Whilst no causal relationship can be inferred, raised DKK-1 levels may help to provide an explanation for the long-observed association between inflammation and bone destruction. Collectively, these data indicate that DKK1 expression may be involved in early RA pathogenesis, through both the perpetuation of the inflammatory response and enhanced joint destruction.

DKK1 directly impairs osteoblast differentiation and indirectly enhances bone destruction by increasing RANKL-induced osteoclastogenesis [10, 19]. In established RA, expression of DKK1 within the synovium localises to synovial fibroblasts ex vivo [10] and is tightly regulated by glucocorticoid metabolism in vitro [20], supporting a role for Wnt signalling inhibition in RA bone destruction. In the work we present, differential expression of DKK1 in resolving and early persistent disease suggests that increased

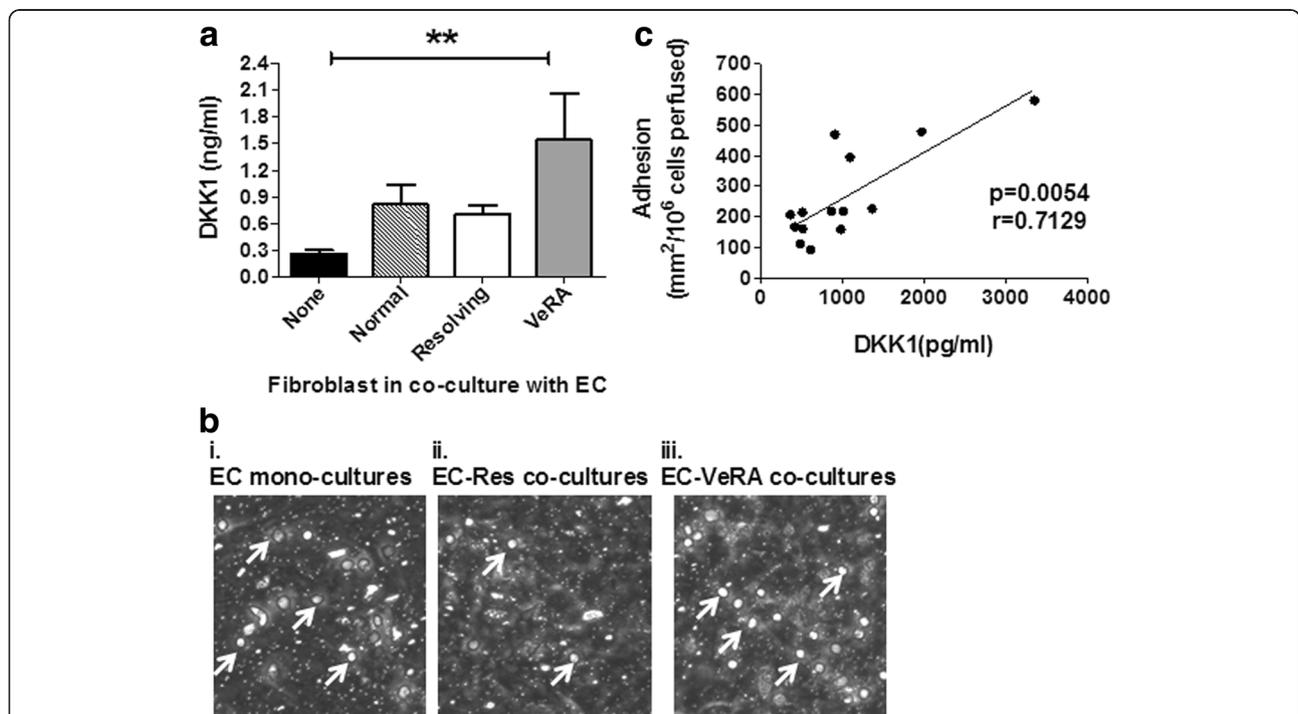


Fig. 2 Dickkopf-related protein 1 (DKK1) levels in human umbilical vein endothelial cell (HUVEC) synovial fibroblast co-cultures. **a** DKK1 release from tumour necrosis factor (TNF)-α + interferon (IFN)-γ-treated endothelial cells cultured alone (none; n = 7) or with synovial fibroblasts from non-inflamed joints (n = 4), patients with resolving rheumatoid arthritis (RA; n = 5) or patients with very early rheumatoid arthritis (VeRA; n = 5). Significantly higher DKK1 levels were observed in the supernatant of VeRA-HUVEC co-cultures than HUVEC alone (p < 0.01). No statistical differences were found between the different co-culture constructs. Data are shown as mean and standard error of the mean. Kruskal-Wallis test with Dunn’s post-test analysis were performed. **p < 0.01. **b** Photomicrographs showing lymphocyte adhesion to (i) endothelial cells cultured alone, with fibroblasts from (ii) patients with resolving RA or (iii) patients with VeRA in response to TNF-α + IFN-γ treatment. The number of adherent lymphocytes was averaged per field and is expressed per square millimetre per 10⁶ cells perfused. Arrows indicate adherent lymphocytes. **c** DKK1 release positively correlated with lymphocyte adhesion to cytokine-treated co-cultures (p = 0.0054). Correlation assessed by Spearman’s test with a non-parametric correlation coefficient of r = 0.7129. Each independent experiment incorporated a different donor for all cell types

DKK1 production could be a key event in progression to RA and occurs early in the disease process. Wnt signalling inhibition by DKK1 may therefore be an as yet undefined pathway through which synovial fibroblasts influence bone destruction in early RA. Further work in this area should be directed towards confirmation of findings in *in vivo* models of arthritis.

Conclusions

Synovial fibroblasts of patients with early inflammatory arthritis that persists as RA express higher DKK1 levels than those from patients with inflamed joints whose arthritis resolves. This phenomenon is amplified by co-culture with endothelial cells and associates with increased lymphocyte adhesion to co-cultures. We propose that DKK1 expression may be involved in early RA pathogenesis, through both perpetuation of the inflammatory response and enhanced joint destruction. Targeting of DKK1 may be a valid therapeutic approach early in the disease course.

Abbreviations

ACR/EULAR: American College of Rheumatology/European League Against Rheumatism; BEACON: Birmingham early arthritis cohort; CCP: cyclic citrullinated peptide; CRP: C-reactive protein; DAS28: Disease Activity Score in 28 joints; DKK1: Dickkopf-related protein 1; DMARD: disease-modifying anti-rheumatic drug; ESR: erythrocyte sedimentation rate; HUVEC: human umbilical vein endothelial cell; IFN: interferon; mRNA: messenger RNA; RA: rheumatoid arthritis; RANKL: receptor activator of nuclear factor κ B ligand; RF: rheumatoid factor; SJC28: 28-joint swollen joint count; TJC28: 28-joint tender joint count; TNF: tumour necrosis factor; VAS: visual analogue scale; VeRA: very early rheumatoid arthritis; Wnt: Wingless.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MJ and HMM carried out the experimental investigations and analysed and interpreted data. MJ, CDB, KR and AF recruited and diagnosed patients and acquired the clinical data. MJ, HMM and AF conceived the study, interpreted data and drafted the manuscript. DST, RH and MC contributed to study design. LY, JS and BdP contributed to experimental investigations. All authors contributed to the analysis and discussion of the data, along with editing of the manuscript. All authors read and approved the final manuscript.

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References

- Cush JJ. Early rheumatoid arthritis: is there a window of opportunity? *J Rheumatol Suppl.* 2007;80:1–7.
- Raza K, Falciani F, Curnow SJ, Ross EJ, Akbar CY, Lord AN, et al. Early rheumatoid arthritis is characterized by a distinct and transient synovial fluid cytokine profile of T cell and stromal cell origin. *Arthritis Res Ther.* 2005;7:R784–95.
- Karouzakis E, Gay RE, Michel BA, Gay S, Neidhart M. DNA hypomethylation in rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum.* 2009;60:3613–22.
- Kasperkowitz PV, Timmer TC, Smeets TJ, Verbeet NL, Tak PP, van Baarsen LG, et al. Fibroblast-like synoviocytes derived from patients with rheumatoid arthritis show the imprint of synovial tissue heterogeneity: evidence of a link between an increased myofibroblast-like phenotype and high inflammation synovitis. *Arthritis Rheum.* 2005;52:430–41.
- Müller-Ladner U, Kriegsmann J, Franklin BN, Matsumoto S, Geiler T, Gay RE, et al. Synovial fibroblasts of patients with rheumatoid arthritis attach to and invade normal human cartilage grafted into SCID mice. *Am J Pathol.* 1996;149:1607–15.
- Lehmann J, Jüngel A, Lehmann I, Busse F, Biskop M, Saalbach A, et al. Grafting of fibroblasts isolated from the synovial membrane of rheumatoid arthritis (RA) patients induces chronic arthritis in SCID mice—a novel model for studying the arthritogenic role of RA fibroblasts *in vivo*. *J Autoimmun.* 2000;15:301–13.
- Korb A, Pavenstädt H, Pap T. Cell death in rheumatoid arthritis. *Apoptosis.* 2009;14:447–54.
- Pap T, Müller-Ladner U, Gay RE, Gay S. Fibroblast biology: role of synovial fibroblasts in the pathogenesis of rheumatoid arthritis. *Arthritis Res.* 2000;2:361–7.
- Shiegeyama Y, Pap T, Kunzler P, Simmen BR, Gay RE, Gay S. Expression of osteoclast differentiation factor in rheumatoid arthritis. *Arthritis Rheum.* 2000;43:2523–30.
- Diarra D, Stolina M, Polzer K, Zwerina J, Ominsky MS, Dwyer D, et al. Dickkopf-1 is a master regulator of joint remodelling. *Nat Med.* 2007;13:156–63.
- Wang SY, Liu YY, Ye H, Guo JP, Li R, Liu X, et al. Circulating Dickkopf-1 is correlated with bone erosion and inflammation in rheumatoid arthritis. *J Rheumatol.* 2011;38:821–7.
- de Rooy DPC, Yeremenko NG, Wilson AG, Knevel R, Lindqvist E, Saxne T. Genetic studies on components of the Wnt signalling pathway and the severity of joint destruction in rheumatoid arthritis. *Ann Rheum Dis.* 2013;72:769–75.
- Kelly S, Huby F, Filer A, Di Cicco M, Hands RE, Rocher V, et al. Ultrasound-guided synovial biopsy: a safe, well-tolerated and reliable technique for obtaining high-quality synovial tissue from both large and small joints in early arthritis patients. *Ann Rheum Dis.* 2015;74:611–7.
- Scire CA, Epis O, Codullo V, Humby F, Morbini P, Manzo A, et al. Immunohistological assessment of the synovial tissue in small joints in rheumatoid arthritis: validation of a minimally invasive ultrasound-guided synovial biopsy procedure. *Arthritis Res Ther.* 2007;9:R101.
- Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham 3rd CO, et al. 2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum.* 2010;62:2569–81.
- Filer A, Parsonage G, Smith E, Osborne C, Thomas AM, Curnow SJ, et al. Differential survival of leukocyte subsets mediated by synovial, bone marrow and skin fibroblasts: site-specific versus activation-dependent survival of T cells and neutrophils. *Arthritis Rheum.* 2006;54:2096–108.

17. McGettrick HM, Smith E, Filer A, Kissane S, Salmon M, Buckley CD, et al. Fibroblasts from different sites may promote or inhibit recruitment of flowing lymphocytes by endothelial cells. *Eur J Immunol.* 2009;39:113–25.
18. Cooke BM, Usami S, Perry I, Nash GB. A simplified method for culture of endothelial cells and analysis of adhesion of blood cells under conditions of flow. *Microvasc Res.* 1993;45:33–45.
19. Goldring SR, Goldring MB. Eating bone and adding it: the Wnt pathway decides. *Nat Med.* 2007;13:133–4.
20. Hardy R, Juarez M, Naylor A, Tu J, Rabbitt EH, Filer A, et al. Synovial DDI1 expression is regulated by the local glucocorticoid metabolism in inflammatory arthritis. *Arthritis Res Ther.* 2012;14:R226.

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