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DOI:

[10.1371/journal.pone.0164253](https://doi.org/10.1371/journal.pone.0164253)

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Document Version

Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Singh, A, Varela Ramirez, CH, Bhatt, K, Veerapen, N, Lee, OYC, Wu, HHT, Besra, GS, Minnikin, DE, Fujiwara, N, Teramoto, K & Bhatt, A 2016, 'Identification of a Desaturase Involved in Mycolic Acid Biosynthesis in *Mycobacterium smegmatis*', *PLoS ONE*, vol. 11, no. 10, e0164253.
<https://doi.org/10.1371/journal.pone.0164253>

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Checked 04/11/2016

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RESEARCH ARTICLE

Identification of a Desaturase Involved in Mycolic Acid Biosynthesis in *Mycobacterium smegmatis*

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Citation: Singh A, Varela C, Bhatt K, Veerapen N, Lee OYC, Wu HHT, et al. (2016) Identification of a Desaturase Involved in Mycolic Acid Biosynthesis in *Mycobacterium smegmatis*. PLoS ONE 11(10): e0164253. doi:10.1371/journal.pone.0164253

Editor: Laurent Kremer, Centre National de la Recherche Scientifique, FRANCE

Received: May 18, 2016

Accepted: September 7, 2016

Published: October 14, 2016

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Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: KT is an employee of Jeol Ltd. The funder provided support in the form of salaries for author KT, but did not have any additional role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript. The specific roles of this author is articulated in the 'author contributions' section. AB received grant funding from the Medical Research Council (UK). The funder provided support did not have any additional role in the study design, data collection

Abstract

Mycolic acids are unique long chain fatty acids found in the cell walls of mycobacteria including the tubercle bacillus, *Mycobacterium tuberculosis*. The introduction of double bonds in mycolic acids remains poorly understood, however, genes encoding two potential aerobic desaturases have been proposed to be involved in this process. Here we show that one of these genes, *desA1*, is essential for growth of the saprophytic *Mycobacterium smegmatis*. Depletion of *desA1* in a *M. smegmatis* conditional mutant led to reduction of mycolic acid biosynthesis and loss of viability. The DesA1-depleted cells exhibited two other phenotypes: using ¹⁴C-labelling, we detected the accumulation of minor mycolic acid-related species that migrated faster in a silver TLC plate. Spiral Time of Flight Mass Spectroscopic analysis suggested the presence of species with sizes corresponding to what were likely monoenoic derivatives of α-mycolic acids. Additionally, conditional depletion led to the presence of free fatty acyl species of lengths ~C₂₆-C₄₈ in the lysing cells. Cell viability could be rescued in the conditional mutant by *Mycobacterium tuberculosis desA1*, highlighting the potential of *desA1* as a new drug target in pathogenic mycobacteria.

Introduction

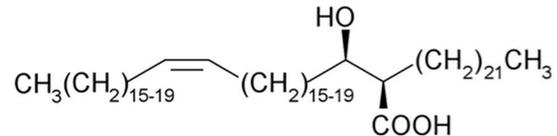
Mycobacterium tuberculosis, the bacterium that causes tuberculosis (TB), synthesises a distinct class of long-chain fatty acids termed mycolic acids. These α-alkyl, β-hydroxy fatty acids are abundant in the waxy cell wall of the tubercle bacillus and the cell walls of other mycobacteria including the leprosy bacillus *Mycobacterium leprae*, bovine TB-causing *Mycobacterium bovis* and the saprophytic *Mycobacterium smegmatis*. Mycolic acids are found either esterified to the terminal carbohydrate moieties of the peptidoglycan-arabinogalactan complex, or as part of the outer glycolipids, glucose monomycolate, trehalose monomycolate and trehalose dimycolate

and analysis, decision to publish, or preparation of the manuscript. The specific roles of this author is articulated in the 'author contributions' section.

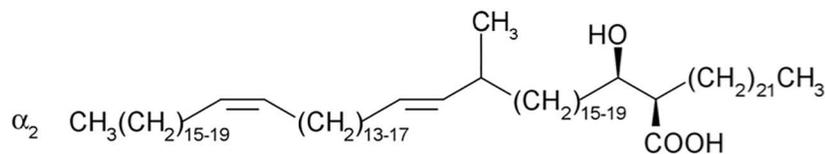
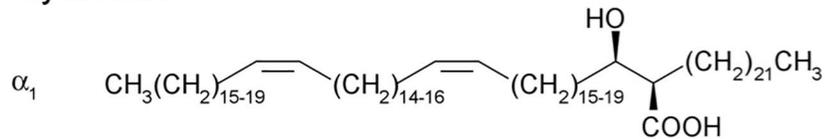
Competing Interests: This affiliation does not alter our adherence to PLOS ONE policies on sharing data and materials.

(TDM; also known as cord factor). A combination of early biochemical studies and subsequent development of mycobacterial genetics has helped unravel the pathways leading to the biosynthesis of mycolic acids [1–3]. Mycolic acids in mycobacteria consist of a longer mero-chain, and a shorter α -chain. Biosynthesis of mycolic acids is initiated by the 'housekeeping' fatty acid synthase, the multidomain fatty acyl synthase—I (FAS-I) that produces C_{16} – C_{18} and C_{24} – C_{26} fatty acids in a bimodal fashion. The latter forms the α -chain of mycolic acids. The mero-chain is synthesised by the multienzyme fatty acid synthase II (FAS-II) that extends FAS-I derived palmitoyl/stearoyl-S-CoA to yield fatty acids of chain length C_{60} . The mero-chain then undergoes Claisen condensation with the α -chain; a reaction catalysed by a polyketide synthase, Pks13 to yield a 3-oxo, α -alkyl fatty acid [4,5], which is subsequently reduced by a mycolyl reductase to form a mycolic acid [6,7]. Core to mycolic acid structure is the generation of double bonds found in the mero-chain. *M. smegmatis* produces three subclasses of mycolic acids: α , α' and epoxy mycolic acids (Fig 1); α mycolic acids contain two double bonds in the mero-chain, while α' mycolates contain one double bond. In *M. tuberculosis*, formation of *cis*-double bonds can be a precursor for subsequent modification steps such as cyclopropanation [8–10]. Asselineau *et al.* [11] hypothesised that a $C_{24:0}$ fatty acyl chain is first desaturated to a *cis*- Δ^5 - $C_{24:1}$ intermediate which is then elongated by FAS-II, and subsequently desaturated to yield *cis*- Δ^3 , *cis*- Δ^{15} - $C_{34:2}$ merochains which are eventually elongated further and incorporated into mycolic acids. In *Escherichia coli*, fatty acid desaturation is brought about by the action of FabA, a dual function enzyme that catalyses both the formation of a *trans*-double bond in a fatty acyl-chain, and its subsequent isomerisation to the *cis* configuration [12]. In the absence of FabA homologues in the genomes of mycobacteria, two alternative pathways to double bond

α' -Mycolates



α -Mycolates



Epoxy mycolates

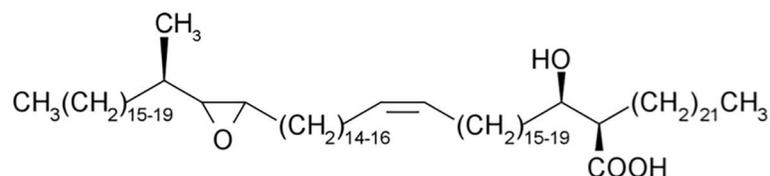


Fig 1. Structures of mycolic acid subclasses found in *Mycobacterium smegmatis*.

doi:10.1371/journal.pone.0164253.g001

formation have been suggested [3]. In *Streptococcus pneumoniae*, the novel isomerase/dehydratase pair FabM/FabZ diverts fatty acid biosynthesis to the unsaturated mode [13]. The *M. tuberculosis* homologues *echA10* and *echA11*, have been proposed as candidates, though their predicted non-essentiality makes it unlikely that they play any role in mycolic acid desaturation [3]. An alternative pathway involving a $\Delta 5$ desaturase has also been proposed to convert $C_{24:0}$ to $C_{24:1}$ prior to further elongation to yield the α -mycolate mero-chain [11]. Such a desaturase would require molecular oxygen, and interestingly the *M. tuberculosis* H37Rv genome contains genes encoding three candidate aerobic desaturases: *desA1*, *desA2* and *desA3* [14]. Of these, *desA3* has been shown to be involved in the biosynthesis of oleic acid, rather than mycolic acids [15]. Homologues of *M. tuberculosis desA1*, *desA2* and *desA3* are found in other mycobacteria including *M. bovis*, *M. leprae* and *M. smegmatis*, though *desA3* is predicted to be a pseudogene in *M. leprae* [16,17]. Dyer *et al.* [18] attempted structural studies on DesA1 and DesA2, and while the authors were unable to obtain soluble DesA1, the solved structure of DesA2 [18] revealed that DesA2 exists as a homodimer and is structurally related to plant fatty acid desaturases. However, differences in the variable metal binding pockets of DesA2, including a possible alternative access of ligand to the metal center, suggested a structurally distinct substrate for this mycobacterial fatty acid desaturase. This, and the extent of disorder in the inter-subunit region of the protein indicated a specialised role for DesA2. Here, we have investigated the role of *desA1* in the desaturation of mycolic acid mero-chains in the fast growing *M. smegmatis*.

Results

DesA1 is essential for mycobacterial viability during growth in laboratory media

The protein encoded by *M. tuberculosis desA1* (Rv0824c) shares 80% identity and 90% similarity with the *M. smegmatis* homologue encoded by MSMEG5773 (multiple sequence alignment shown in Fig 2). To test the potential essentiality of *desA1*, we first attempted to generate a null mutant of *desA1* in *M. smegmatis* by Specialized Transduction [19,20] using ph Δ MSMEG5773, a recombinant phage designed to replace *M. smegmatis desA1* (MSMEG5773) with a hygromycin resistance cassette (*hyg*). Repeated transductions of *M. smegmatis* mc²155 with ph Δ MSMEG5773 failed to generate any hygromycin resistant transductants suggesting that the gene was likely to be essential for growth in laboratory media. We then proceeded to demonstrate the essentiality of *desA1* using CESTET, a gene essentiality-testing tool for *M. smegmatis* [21]. First, we generated a merodiploid strain containing a second, integrated copy of *desA1* under the control of an acetamide-inducible promoter. Following transduction of the merodiploid strain, we were able to obtain hygromycin resistant colonies on agar plates containing acetamide. One such transductant was confirmed by Southern blot to have the native copy of *desA1* replaced with *hyg*, and was designated Δ Ms*desA1*. When broth cultures of Δ Ms*desA1* were transferred to fresh medium containing acetamide, they continued growing, however when cultures were transferred to medium without acetamide, a relative decrease in viable counts was observed (Fig 3) indicating that *desA1* was essential for growth of *M. smegmatis* in laboratory media. All together, these results validated the essentiality of *desA1* in *M. smegmatis*.

Loss of *M. smegmatis* DesA1 leads to loss of mycolic acid biosynthesis

To determine the effect of loss of DesA1 on mycolic acids we extracted ¹⁴C-labelled mycolic acids methyl esters (MAMEs) from cultures of the Δ Ms*desA1* strain grown in the presence or absence of acetamide, and analysed the extracts by 2D-argentation TLC. The second TLC dimension involves separation of saturated and unsaturated species in an AgNO₃-treated



Fig 2. Multiple sequence alignment of *M. tuberculosis* DesA1 (Rv0824c) and homologues from *M. bovis* (Mb0847c), *M. leprae* (ML2185) and *M. smegmatis* (MSMEG5773). Consensus residues found in all four amino acid sequences are indicated by uppercase letters.

doi:10.1371/journal.pone.0164253.g002

section of the TLC plate, as the presence of double bonds retards migration in this dimension. Cultures grown in the absence of acetamide showed a decrease in the biosynthesis of subclasses of α , α' and epoxy mycolic acids over time (Fig 4). Additionally, we also observed a previously described cyclopropanated α -mycolate species [22] in both cultures (Labelled X₁ in Fig 4A). Levels of X₁ did not decrease following depletion of DesA1, even at the later time point. This was surprising, as cyclopropane ring formation requires double bond formation.

In addition to the above phenotypes, DesA1-depleted cultures also revealed the presence of faint but detectable levels of extra species of fatty acyl methyl esters that predominantly co-migrated with α -MAMES in the first dimension, but migrated further than the α -MAMES in the second, AgNO₃-containing dimension in the TLC plate (labelled X₂ in Fig 4A). This result suggested that the faster migrating spots (X₂) in the 2D-TLC plates were likely fatty acyl methyl esters related to α -mycolic acids.

To further characterise mycolic acid species in the conditional mutant, we first subjected purified, non-labelled α -MAMES to high-resolution matrix-assisted laser desorption/ionization time-of-flight mass spectrometry. Unlike ¹⁴C-labelled cultures, using non-labelled cells does not offer an opportunity to study mycolates in a time dependent fashion. Instead, MAMES extracted from non-labelled, no-acetamide cultures represent both 'regular' mycolates produced during the early stages of DesA1 depletion, and those produced at later stages of DesA1 depletion. The levels of these early stage 'regular' mycolates (which would also be present in the +acetamide cultures) would thus dominate in any subsequent analysis. Thus, the unknown, accumulating MAMES (species X₂) in the DesA1-depleted cultures would represent only a fraction of the total MAMES extracted from non-labelled cell pellets.

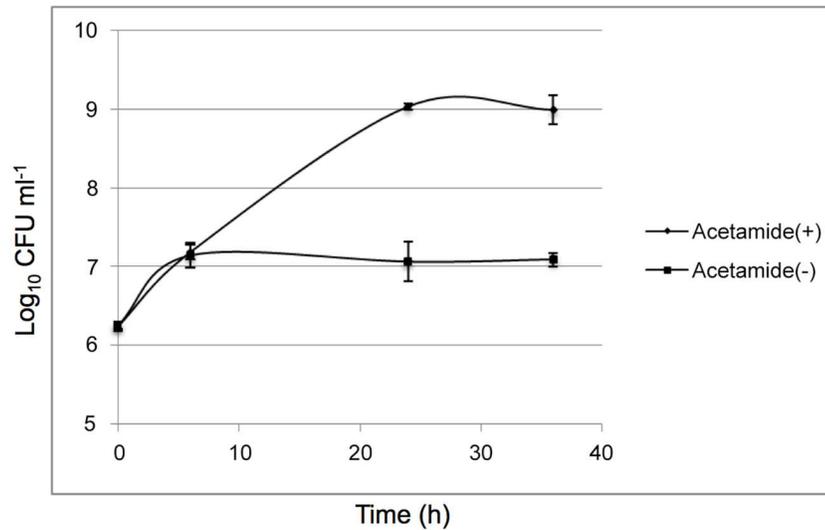


Fig 3. Conditional depletion of DesA1 leads to lysis of the $\Delta MsdesA1$ mutant. Cultures were grown for up to 36h in TSB in the presence (+Acetamide) or absence (-Acetamide) of inducer. Loss of viability was more distinct on solid media (See control strain in Fig 12).

doi:10.1371/journal.pone.0164253.g003

The molecular masses calculated for the observed MAME species in cultures grown the presence of acetamide indicated that dioenic $C_{77:2}$ and $C_{79:2}$ were the major α -MAMES in the non-depleted cultures (+acetamide; Fig 5A) though other species, including, surprisingly those with masses corresponding to monoenoic derivatives, were also present. While the DesA1 depleted cultures (-acetamide; Fig 5B) showed the presence of same species, we detected slightly higher relative peak intensities for MAMES with sizes predicted to represent monoenoic derivatives $C_{74:1}$, $C_{76:1}$, $C_{78:1}$ and $C_{80:1}$ (Fig 5C).

We further analysed total mycolates extracted from cultures of $\Delta MsdesA1$ grown in the presence or absence of acetamide, using MALDI-TOF MS with a spiral trajectory [23,24]. We chose this method over conventional MALDI-TOS MS as it gives high mass resolution and accuracy, and enables the separation of adjacent peaks with a 0.036 Da mass difference. Consistent with our findings with purified α -MAMES, we observed molecular masses corresponding to both monoenoic and dioenic α -MAMES (Fig 6; S1 and S2 Figs, S1 and S2 Tables). Given the accuracy of the method in distinguishing between species differing by 0.036 Da mass differences, we were also able to account for epoxy mycolates and distinguish these from potentially monoenoic α -mycolates of similar masses. For example the epoxy mycolate species $C_{78}H_{152}O_4Na$ (m/z 1176.158) could be differentiated from monoenoic α -mycolate $C_{79}H_{156}O_3Na$, (m/z 1176.194). However, we could not distinguish between dioenic α -mycolates and their cyclopropanated derivatives as species with the same number of carbons would have an identical mass.

While ^{14}C -acetate labelling afforded a sensitive method for visualising newly synthesised mycolates extracted from smaller culture volumes within defined time frames, further MS and NMR analysis required non-radioactive, unlabelled samples derived from scaled up cultures. Bacterial lysis limited the amount of cell material we could collect for this analysis. Furthermore, as mentioned earlier, unlabelled cell pellets would yield mycolates synthesised both in the early and late stages of depletion and thus the observed differences between the relative levels of monenoic and dioenic mycolates were perhaps under-represented here. Given the relatively low levels of species X_2 expected in non-labelled samples, we were unable to further study structural features of this species from DesA1-depleted cultures.

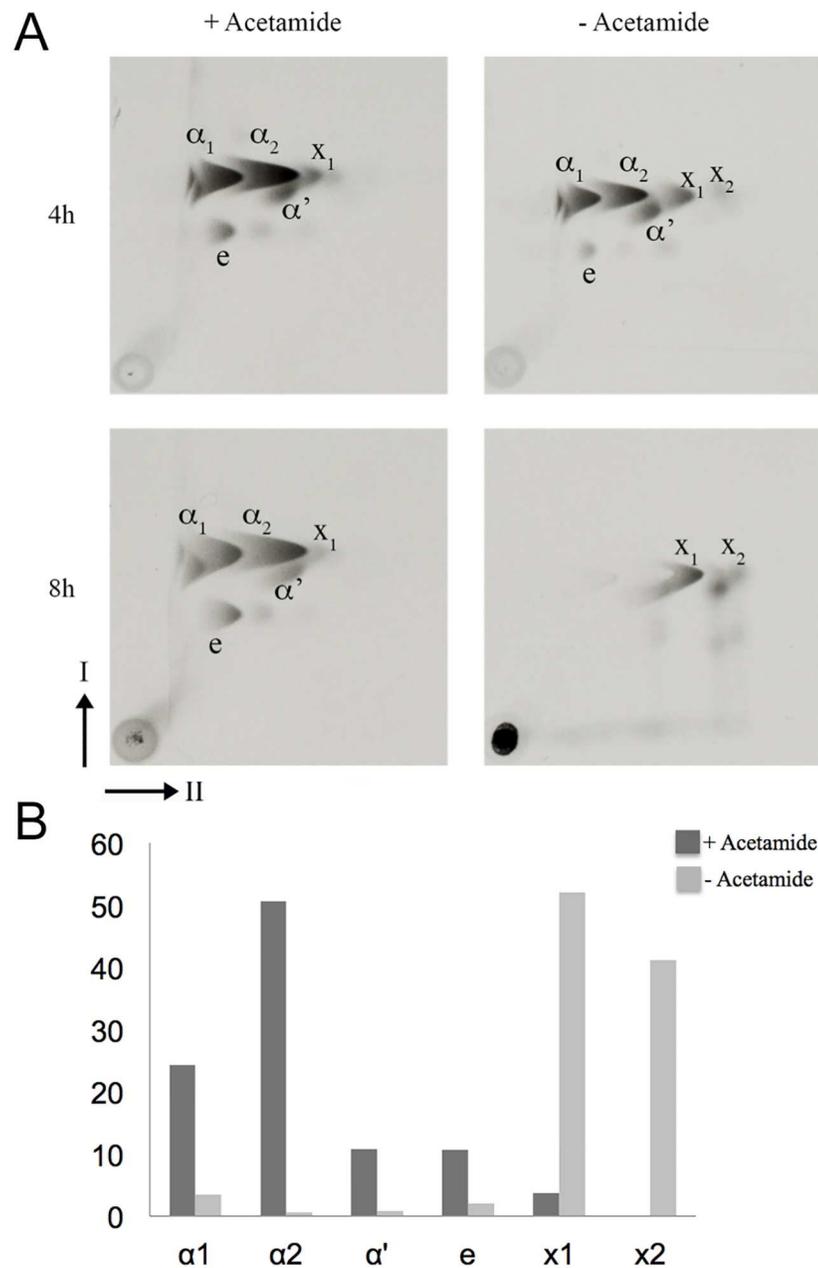


Fig 4. Argentation TLC of ¹⁴C-labelled methyl esters of cell wall mycolic acids (MAMES) extracted from the $\Delta MsdesA1$ mutant grown in the presence or absence of acetamide. (A) Direction-I separated in petroleum ether:acetone (95:5, v/v); direction-II separated thrice in AgNO₃ treated silica gel in petroleum ether:ethyl acetate (9:1, v/v). The cyclopropanated derivative is indicated as X₁ and the faster-migrating species that co-migrated with α -MAMES in direction-I are indicated as X₂. The clear 'imprint' appearing superimposed on the comet shaped X₂ spot in the 8h -acetamide TLC is likely due to unlabelled α_2 mycolates synthesised at the early stages of DesA1 depletion. (B) Bar graph showing the relative amounts of each ¹⁴C-labelled mycolate subspecies indicated as a percentages of the total amounts of ¹⁴C-labelled mycolic acids detected on the TLC plates, as determined by densitometry.

doi:10.1371/journal.pone.0164253.g004

In summary, the 2D-argentation TLC results and subsequent MS analysis indicate that loss of DesA1 leads to changes in the relative proportions of monoenoic and dienoic derivatives of α -MAMES following loss of DesA1 function. We cannot conclude that depletion of DesA1 led to an overall accumulation of monoenoic derivatives of α -mycolic acids. However, using Mass

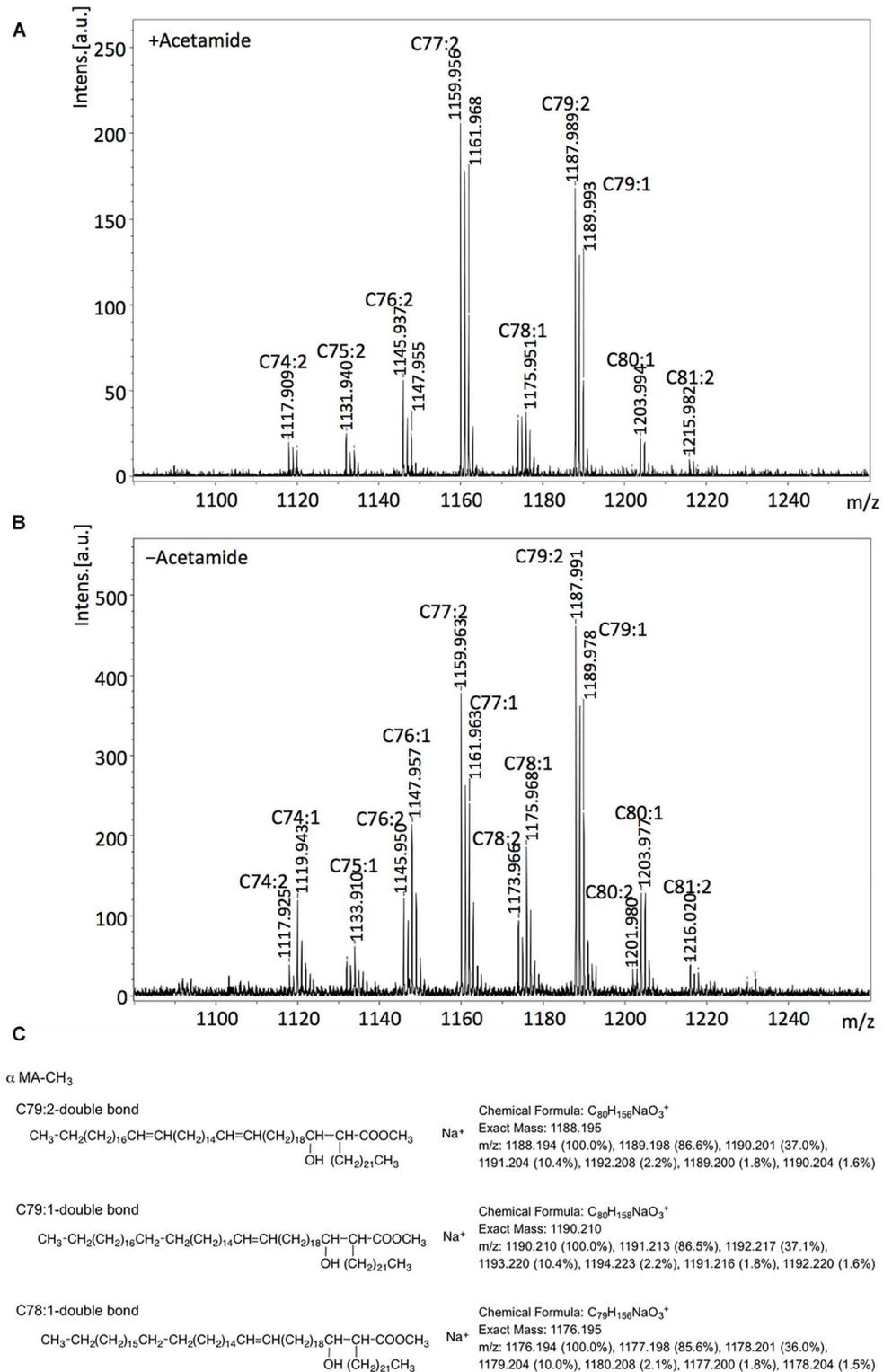


Fig 5. MALDI Mass Spectroscopy analysis of α -MAMES purified from the Δ MsdesA1 mutant grown in the presence or absence of acetamide. A, mass spectra of α -MAMES extracted from (A) + acetamide cultures and (B) -acetamide cultures. (C) Example of predicted sizes and structures based on observed masses.

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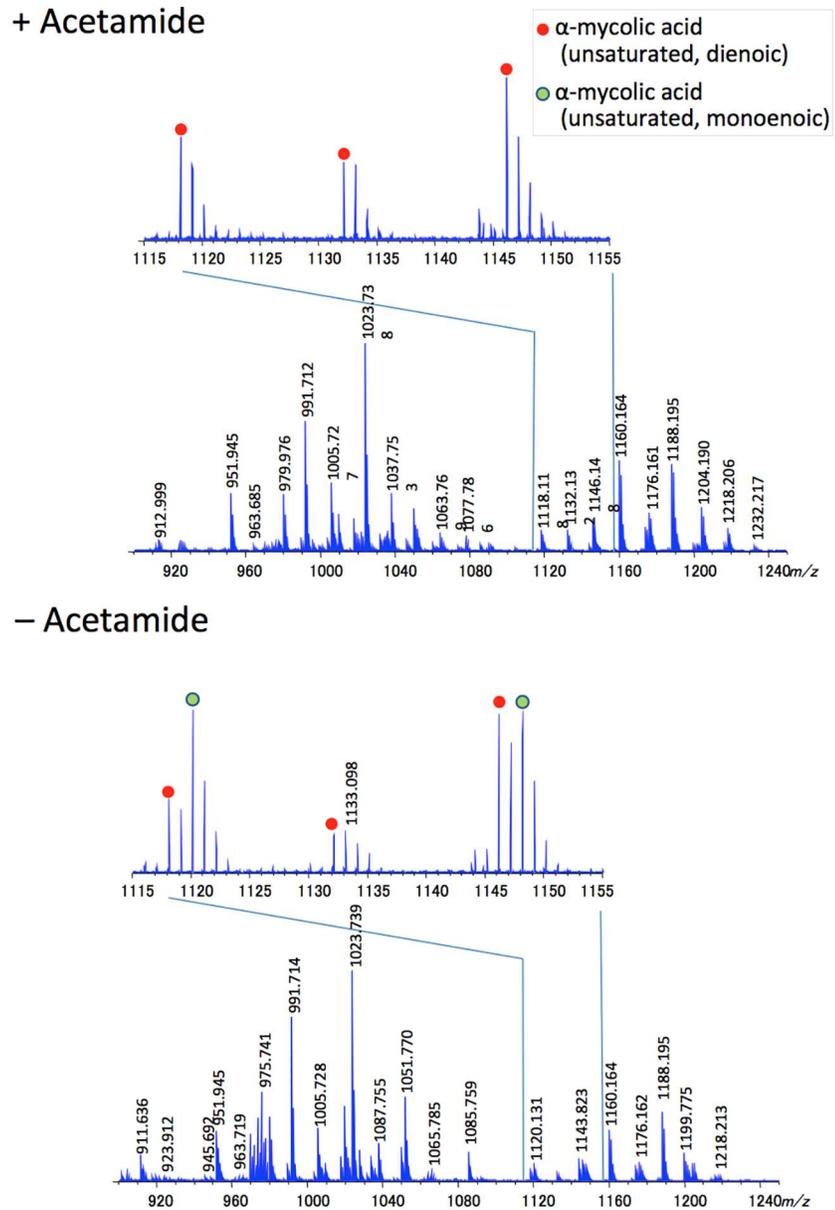


Fig 6. MALDI Spiral-TOF Mass Spectroscopy analysis of total MAMES purified from the $\Delta MsdesA1$ mutant grown in the presence or absence of acetamide.

doi:10.1371/journal.pone.0164253.g006

Spectroscopy, we were able to show changes in the relative proportions of monoenoic and dienoic derivatives following loss of DesA1 function.

In parallel, we also analysed fatty acid methyl esters (FAMES) obtained from polar and apolar lipid extracts by 2D-argentation TLC and GC-MS. Unlike MAMES, no additional FAME species were detected in the second dimension of the TLCs (Fig 7). We further analysed FAMES using Gas Chromatography-Mass Spectroscopy (GC-MS). The GC-MS spectra did not reveal any major differences in unsaturated species (Fig 8), indicating that the role of *desA1* was associated with mycolic acids, but not other fatty acids found in *M. smegmatis*.

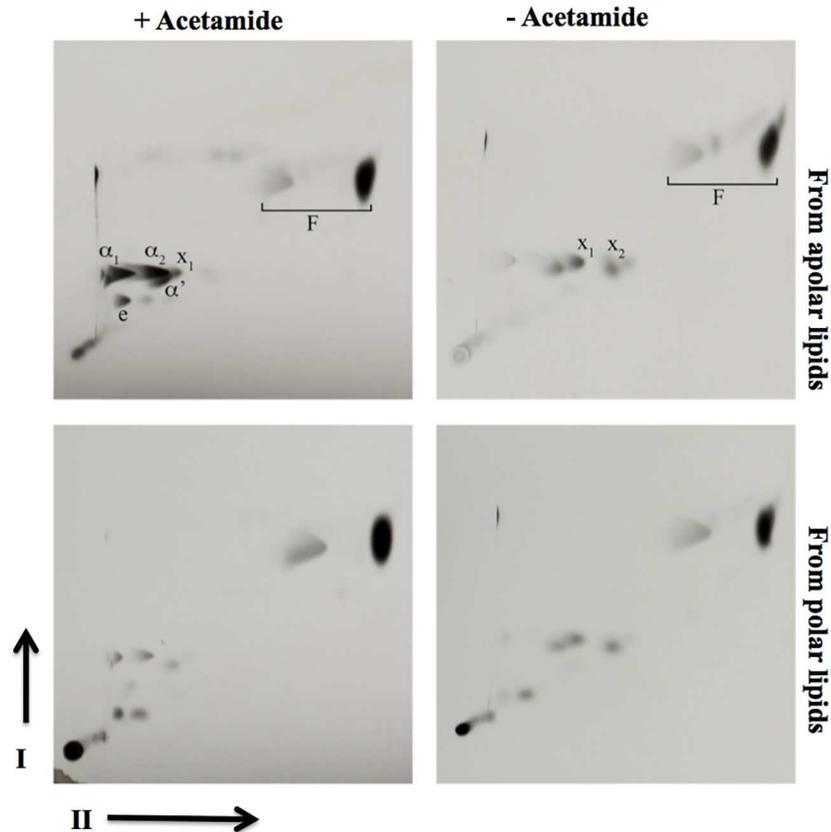


Fig 7. 2D-argentation TLC of fatty acyl methyl esters extracted from polar and apolar lipid extracts of the $\Delta MsdesA1$ mutant grown in the presence or absence of acetamide. (A) Direction-I separated in petroleum ether:acetone (95:5, v/v); direction-II separated thrice in $AgNO_3$ treated silica gel in petroleum ether:ethyl acetate (9:1, v/v). Mycolate species from apolar extracts are indicated and are also present in lesser amounts in the polar extracts. F; fatty acyl methyl esters.

doi:10.1371/journal.pone.0164253.g007

Lipid analysis of the conditional mutant

In addition to studying mycolic acid profiles of the conditional $\Delta MsdesA1$ mutant, we also compared the total lipid profile of the conditional mutant, grown in the presence and absence of acetamide, by 2D-TLC particularly using System C that allows visualisation of free fatty acid and free mycolic acid species [25]. While cultures of $\Delta MsdesA1$ grown in the presence of acetamide showed the patterns expected of the wild type strain (*M. smegmatis* mc²155), i.e. a spot each corresponding to free fatty acids (FA) and free mycolic acids (MA; Fig 9), cultures of the strain grown for 8h in the absence of acetamide showed the accumulation of a lipid species that migrated close to the free MA and FA spots (Fig 9). We then monitored the incorporation of ¹⁴C acetate by cultures of the conditional mutant at different time intervals in the presence or absence of acetamide. Analysis of ¹⁴C-labelled lipids by 2D-TLC using System C showed an accumulation of the same, unknown lipid species in cultures grown for 8h in the absence of acetamide (Fig 9). The presence of this lipid spot predominantly in the cultures growing in the absence of acetamide implied that the accumulation of the spot was directly or indirectly related to the loss of *desA1* function. Furthermore, the accumulation of this lipid species was accompanied by a decrease in the levels of free mycolic acids (Fig 9) suggesting that the accumulating lipid was related to mycolic acids. As expected, we also saw a decrease in levels of TDM after conditional depletion of *DesA1* (Fig 10), but no immediate reduction was seen in

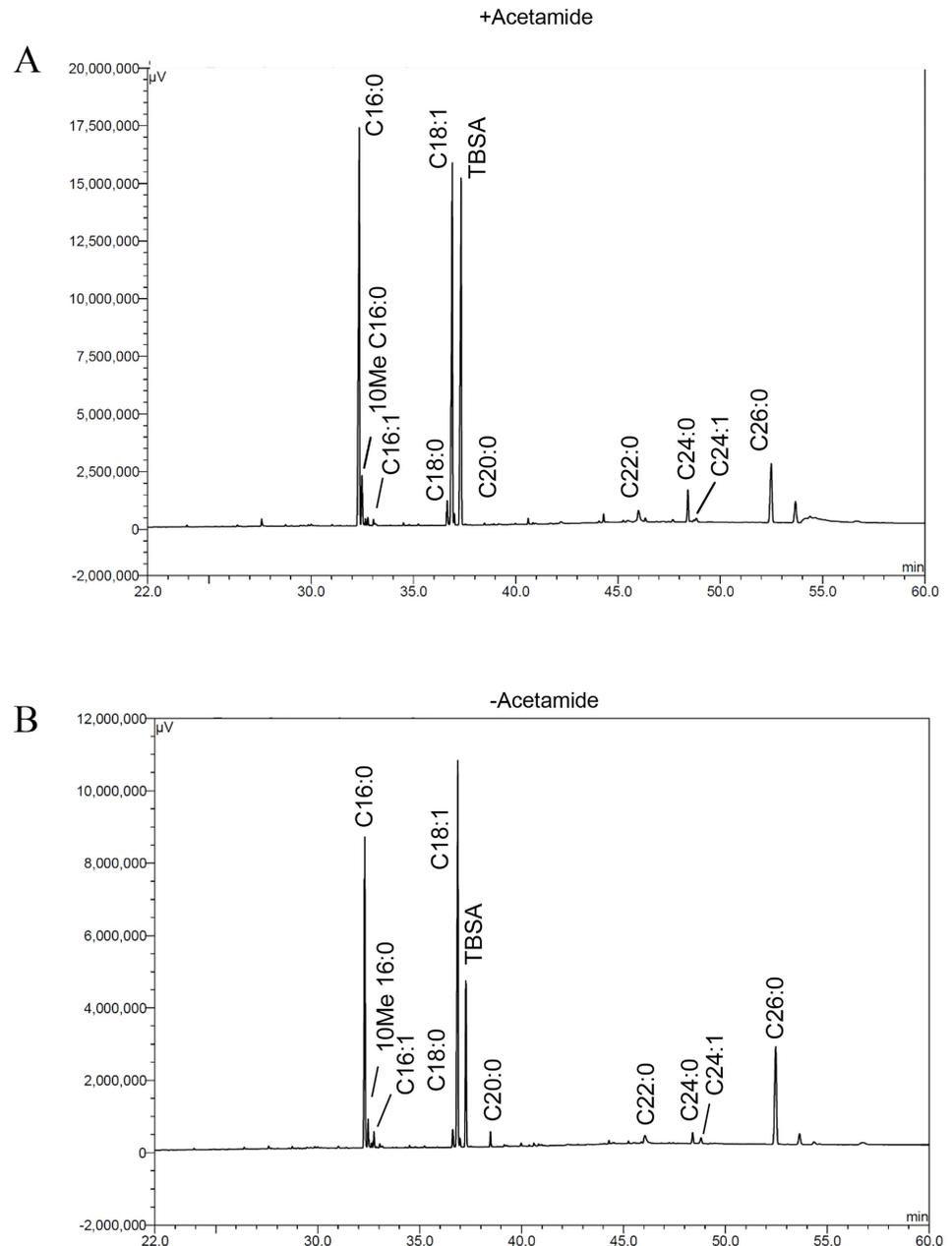


Fig 8. GC-MS analysis of fatty acyl methyl esters prepared from lipid extracts of the $\Delta MsdesA1$ mutant grown in the presence or absence of acetamide. Sigma-Aldrich fatty acyl methyl esters were used as standards. TBSA; tuberculostearic acid.

doi:10.1371/journal.pone.0164253.g008

other apolar lipids (Fig 10). Surprisingly, 2D-TLC of polar lipid fractions revealed relatively lower levels of PIMs in the DesA1-depleted cultures, though levels of other polar lipids remained similar.

Loss of DesA1 leads to the accumulation of a series of fatty acids

The 2D-TLC patterns obtained with the $\Delta MsdesA1$ mutant in Fig 9 suggested that the lipid species accumulating in DesA1-depleted cells could be a fatty acyl species. To further probe the

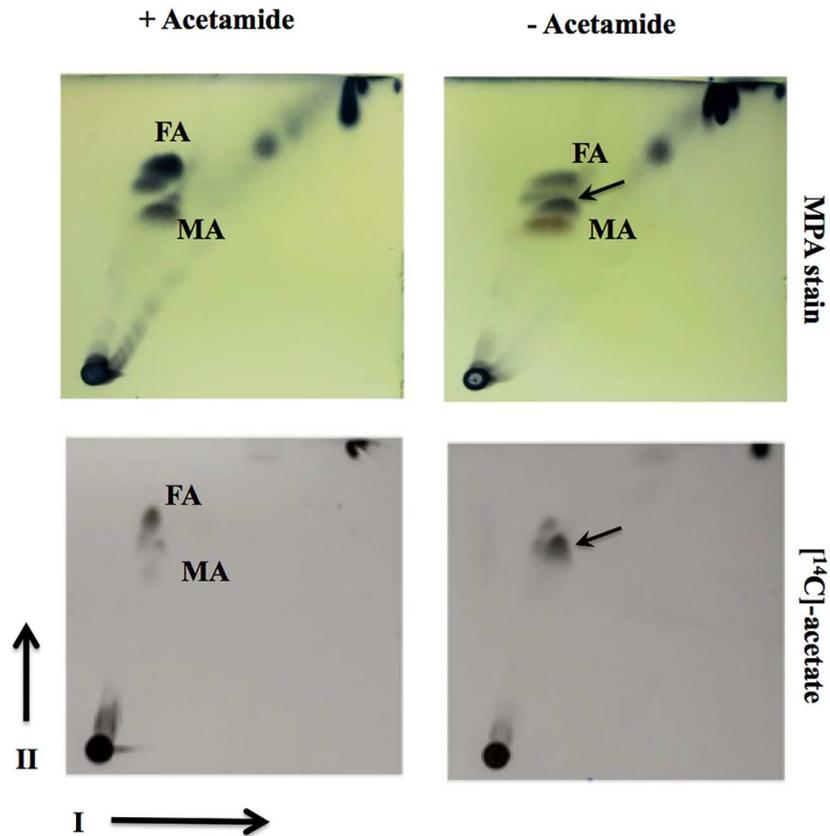


Fig 9. TLC analysis of apolar lipids extracted from the $\Delta MsdesA1$ mutant grown in the presence or absence of acetamide and separated by 2D-TLC using solvent system C. Direction-I, chloroform: methanol, 96:4 (v/v); direction-II, toluene:acetone, 80:20 (v/v). MPA, 5% ethanolic molybdophosphoric acid, followed by charring; MA, free mycolic acids; FA, free fatty acids.

doi:10.1371/journal.pone.0164253.g009

chemical composition of this lipid species, apolar free fatty acyl species (free mycolic acids, free fatty acids and unknown accumulating species) from cultures grown in the presence or absence of acetamide were extracted from preparative TLC plates (separated in System C), and converted into fluorescent dimethoxycoumarin (DMC) esters of fatty acids. The DMC esters were then analysed by reverse phase HPLC analysis (Fig 11). Differences were observed in the HPLC profiles from the extracts of cultures grown in the presence or absence of acetamide. The *DesA1*-depleted cultures showed the presence of a series of fatty acyl species which eluted with a retention times ranging from 15 to 50 min. Comparison of elution times to fatty acid standards and those of mycolic acids suggested that these potential fatty acyl species had chain lengths ranging from approximately C_{26} to C_{48} . Thus, the apolar lipid species migrating close to the free MAs in System C consisted of a mix of fatty acid species ranging from $\sim C_{26}$ - C_{48} .

The *M. tuberculosis desA1* ortholog is functional in *M. smegmatis*

To test whether *M. tuberculosis desA1* (*Rv0824c*), could function compensate for the loss of *M. smegmatis desA1* in the conditional mutant, we first transformed $\Delta MsdesA1$ with a replicating plasmid, pMV206*MtdesA1*, containing *Rv0824c* and its native promoter. We then tested the ability of this strain to survive following growth in the absence of acetamide. Three

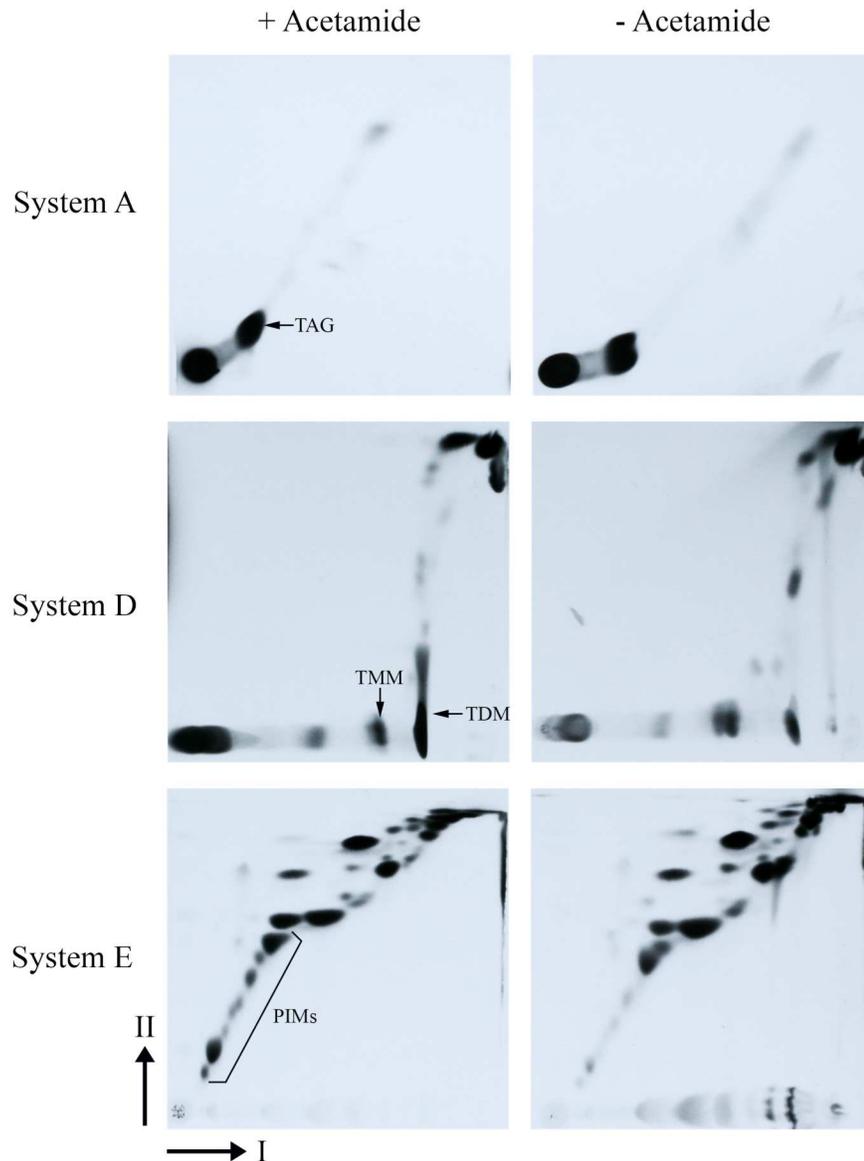


Fig 10. TLC analysis of apolar (Systems A and D) and polar lipids (System E) extracted from the $\Delta MsdesA1$ mutant grown in the presence or absence of acetamide and separated by 2D-TLC using solvent system C. System A direction I; petroleum ether:ethyl acetate 98:2 (X3), direction II; petroleum ether:acetone 98:2. System D direction I; chloroform:methanol:water 100:14:0.8, direction II; chloroform:acetone:methanol:water 50:60:2.5:3. System E direction I; chloroform:methanol:water 60:30:6, direction II; chloroform:acetic acid:methanol:water 40:25:3:6. TAG, triacylated glycerol; TMM, trehalose monoacycolcate; TDM, trehalose dimycolate; PIMs, phosphatidyl inositol mannosides.

doi:10.1371/journal.pone.0164253.g010

independently obtained transformants were able to survive in the absence of acetamide in the media (Fig 12). In contrast, control strains of $\Delta MsdesA1$ containing only the plasmid vector were unable to grow on plates without acetamide (Fig 12). These results demonstrated that *M. tuberculosis desA1* was able to compensate for the loss of *M. smegmatis desA1* and rescue the viability of cultures of $\Delta MsdesA1$ grown in the absence of inducer.

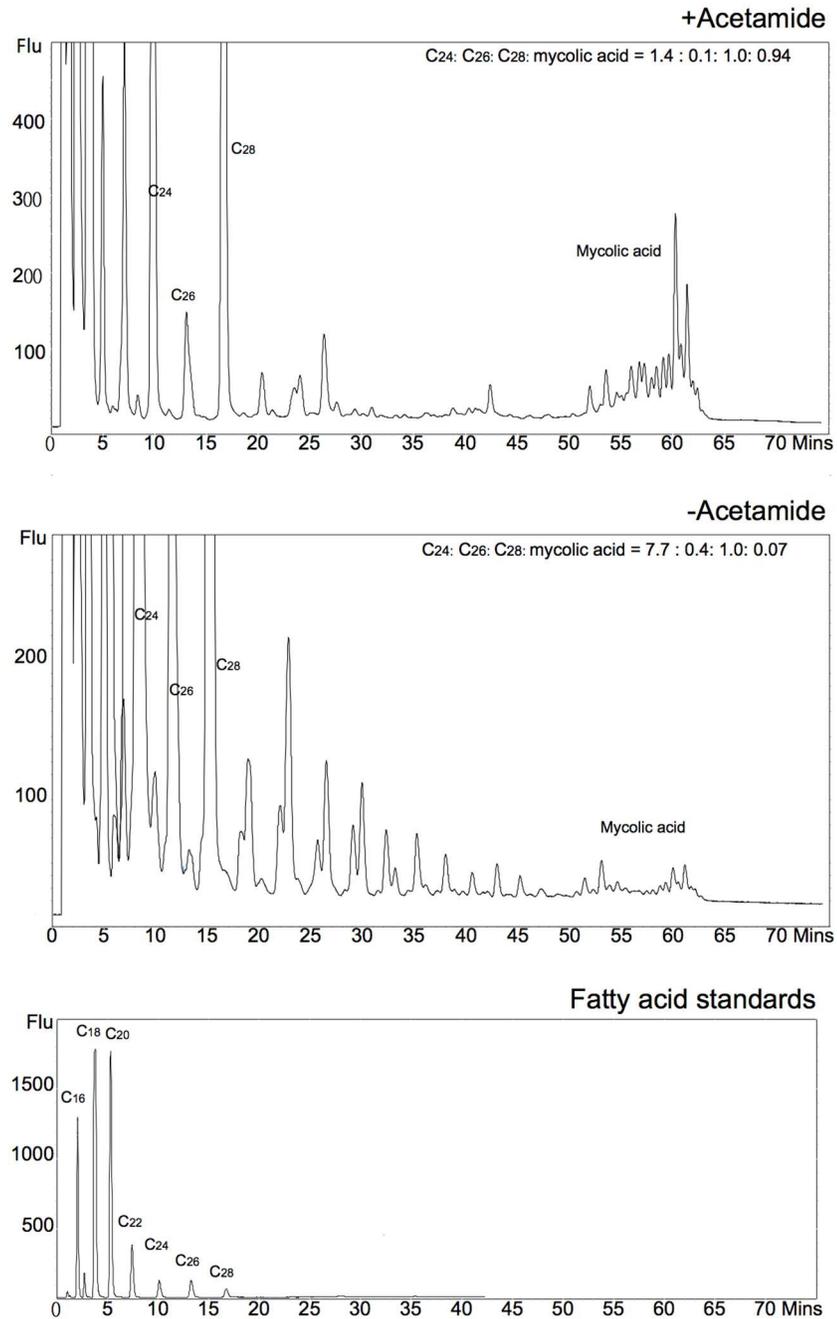


Fig 11. Reverse phase HPLC analysis of dimethoxycoumarin esters of fatty and mycolic acid species. Ratio of injected C₂₆ internal standard to key fatty acyl species and mycolates is indicated in the inset.

doi:10.1371/journal.pone.0164253.g011

Discussion

The aim of this study was to determine whether *desA1*, one of three mycobacterial genes encoding putative aerobic desaturases, played a role in the biosynthesis of mycolic acids. We demonstrated that *MSMEG5773 (desA1)*, homologue of *M. tuberculosis desA1*, is an essential

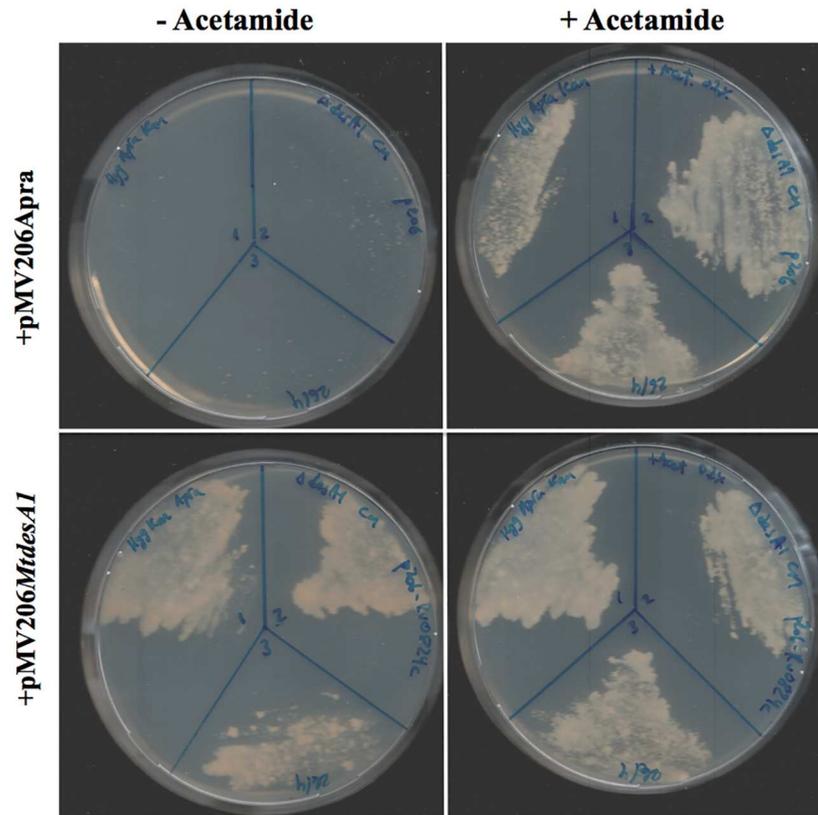


Fig 12. *M. tuberculosis desA1* rescues the growth of $\Delta MsdesA1$ in medium lacking acetamide. $\Delta MsdesA1$ strains containing either empty vector (pMV206Apra) or pMV206MtdesA1 grown on TSB-agar plates with or without acetamide.

doi:10.1371/journal.pone.0164253.g012

gene in *M. smegmatis*, required for growth in laboratory media. We also showed that depletion of DesA1 in a conditional mutant caused loss of mycolic acid biosynthesis and cell viability, and resulted in distinct phenotypes. The first was the accumulation of a species that co-migrated with α -MAMEs in the first dimension of 2D-TLC plates. A comparative Mass Spectroscopic analysis showed that while extracts obtained from cultures grown in the presence of absence of acetamide both contained both monoenoic and dienoic α -mycolate species (based on expected masses), depletion of DesA1 led to change in the relative proportion of the two classes, with levels of the potential monoenoic forms increasing. A conclusive role in double bond addition, and potential identification of double bond position would require a fine structural analysis of these unique species using NMR, however bacterial cell lysis in DesA1-depleted cultures limited our ability to conduct these studies, which would require milligram quantities of purified lipid.

In addition to the phenotype described above, we also observed fatty acyl species of sizes ranging from $\sim C_{28}$ - C_{48} in apolar lipid extracts of cells of the conditional mutant following depletion of DesA1, with a concurrent reduction in levels of free mycolic acids. As FAS-I derived fatty acids are not known to extend beyond C_{26} , these intermediate length fatty acyl species ($\sim C_{28}$ - C_{48}) were thus likely to be truncated mero-mycolate species produced by aborted FAS-II elongation events resulting from a loss of DesA1-catalysed desaturation. Thus, while monoenoic α -mycolate-related species were initially identified in DesA1 depleted cultures, it was likely that FAS-II mediated merochain elongation was not efficient in the absence of

DesA1 and led to the observed series of truncated fatty acyl derivatives. These findings co-relate with earlier hypothesis that desaturation of mycolic acids occurs during merochain elongation [3,26].

We also observed two unexpected phenotypes in the conditional mutant. The first was that levels of a cyclopropanated derivative of α -mycolates remained unaffected by the depletion of DesA1. This was an unexpected outcome as double bond formation precedes cyclopropanation. Fatty acid cyclopropanation is known to increase cell envelope rigidity and it is likely that additional cyclopropanation occurs in residual α -mycolates in lysing cultures to counteract damage to the cell envelope following overall depletion of mycolic acid biosynthesis. The second unexpected outcome of DesA1 depletion was the reduction in the levels of PIMs (Fig 10). This was surprising, as we did not see significant alterations in levels of fatty acids that comprise PIMs in the GC-MS analysis (Fig 8). It seems likely that the observed effect of DesA1 depletion on PIMs is an indirect one.

Our studies have identified *desA1* as a gene that plays a role in mycolic acid biosynthesis. Also, the conditional loss of DesA1 eventually led to cell death, and the viability of the strain could be rescued upon introduction of plasmid-borne *M. tuberculosis desA1*, highlighting the potential of DesA1 as a new anti-*M. tuberculosis* drug target.

Materials and Methods

Generation and characterisation of a *M. smegmatis* MSMEG5773 (*desA1*) conditional mutant

The genetic tool CESTET was used to generate a *M. smegmatis* conditional $\Delta MsdesA1$ mutant [27]. MSMEG5773 was PCR amplified from *M. smegmatis* mc²155 genomic DNA and cloned downstream of an acetamidase promoter fragment into the integrating vector pMV306 [28], to generate pMV306-MSMEG5773. *M. smegmatis* mc²155 [29] was electroporated with pMV306-MSMEG5773 to generate a merodiploid strain mc²155::pMV306-MSMEG5773. In parallel, ph Δ MSMEG5773, a temperature sensitive, recombinant phage designed to replace MSMEG5773 with a hygromycin resistance marker (*hyg*), was generated using protocols described by Larsen *et al.* [20]. Specialized Transduction using the merodiploid strain and ph Δ MSMEG5773 was done as previously described [21,27]. Transductants were selected at the non-permissive temperature of 37°C on selective plates containing 150 μ g/ml hygromycin and 0.02% acetamide. Replacement of the native copy of MSMEG5773 by *hyg* was confirmed by Southern blot. One bonafide knockout, designated as $\Delta MsdesA1$, was selected for further analysis. Conditional depletion of MSMEG5773 (DesA1) in $\Delta MsdesA1$ to monitor mycolic acids and other lipids was done using methods described previously [27].

Complementation of $\Delta MsdesA1$ with *M. tuberculosis desA1*

M. tuberculosis desA1 (Rv0824c) was PCR amplified along with promoter sequences 300bp upstream of the start codon, from genomic *M. tuberculosis* H37Rv genomic DNA and cloned into pMV206Apra, an *E. coli*-*Mycobacterium* shuttle plasmid [30]. The plasmid was introduced into $\Delta MsdesA1$ by electroporation [30]. Three independent apramycin resistant transformants were tested for their ability to grow on TSB-agar plates, with or without acetamide. The $\Delta MsdesA1$ strain was also transformed with pMV206Apra for use as a control strain.

Extraction and characterisation of mycolic acids and other lipids

Extraction of polar, apolar lipids and mycolic acid methyl esters (MAMEs) from the conditional mutant, and subsequent TLC analysis was carried out using protocols described by

Dobson *et al.* [25]. Dry weights of cell pellets were used as a measure to equalise loading on TLC plates. Argentation TLC was carried out as described earlier [28]. Characterisation of the unknown lipid observed in Fig 9 was done by preparative TLC of long-chain acidic components in the apolar lipid using toluene/acetone, 8:2. To convert the acidic components into fluorescent dimethoxycoumarin (DMC) esters [31], a 0.4% solution of 4-bromomethyl-6,7-dimethoxycoumarin in dichloromethane, was allowed to react with the extracts overnight under phase-transfer conditions [25,32]. 0.1mg of each sample was dissolved in 50 μ l heptane. 20 μ l of each sample was placed in a new vial. A concentration of 40 μ g/1000 μ l DMC derivatised C28 was used as an internal control. 20 μ l of the internal control was added to vial prior to HPLC analysis. DMC-esters of saturated C₂₀, C₂₆ and C₂₈ fatty acids were used as standards. DMC esters were separated on an Alltech 205250 C₁₈ reverse phase cartridge, using a water-methanol/dichloromethane elution sequence. Purified DMC esters were eluted by 100% DCM and analysed by reverse phase high performance liquid chromatography (HPLC). HPLC analyses used a VWR Hitachi Elite Pump L-2130, LaChrom Autosampler L-2200, Column Oven L-2300, Fluorescence Detector L-2480 and Foxy Jr. Fraction Collector; data were processed by Scientific Software Inc. EZChrom Elite, version 3.1.6 software. A reverse phase (Alltech 81412 Alltima C18, 3 μ m 50 x 4.6mm) column was used. HPLC analysis conditions are shown below. Samples were dissolved in HPLC grade heptane (500 μ l) and 1 to 5 μ l was injected. Using a flow rate of 1ml/min, a gradient of acetonitrile/tetrahydrofuran; 100:0 to 40:60 over 70 min was used for elution. The ratio of DMC derivatised C₂₄, 26, 28 and mycolic acids (MA) were calculated by the area under peaks from the HPLC profiles. The ratio of the net internal standard C₂₈: MA were calculated by subtracting the background C₂₈ from the total C₂₈.

The GC-FID analysis of fatty acid methyl esters (FAMES) was performed on a Shimadzu GC-2010. The FAMES separation was achieved using a DB-225 column (30 m x 0.25 mm i.d., 0.25 μ m film thickness, J&W Scientific) with a column flow of 1.26ml/min (Helium at a constant pressure of 100kPa). Injector and detector temperatures were 50°C. The Injection volume was 5 μ l. The column temperature was programmed initially at 50°C for 2 mins, increased at the rate of 4°C/min to 220°C, and then maintained at 220°C for a further 30.5 mins. The total program time was 75 mins.

Purification and mass analysis of MAMES

MAMES were detected on TLC described previously [33]. The TLC was developed with the solvent system; benzene or *n*-hexane/diethylether (90:15, vol/vol), three runs. The MAMES were purified from the TLC plate. To assign the structures, the molecular weight of each MAME was measured by MALDI Spiral-TOF neIons generated by irradiation with a 349-nm Nd:YLF laser were accelerated at 20 kV. The settings for delay time and grid voltage were optimized to stay constant at $\Delta M < ca.$ 0.03 Da at full width at half maximum (FWHM) over the range of m/z 500 to 1500. The matrix was 10 mg/mL 2,5-dihydroxybenzoic acid in tetrahydrofuran (THF). A sodium iodide solution of 1 mg/mL in THF was used as a cationization agent. MALDI mass spectra were observed in positive spiral mode by averaging 1500 individual laser shots. Three or five mass spectra for each sample were collected. The structures of MAMES were assigned by the R_f value of TLC and the molecular weight.

Supporting Information

S1 Fig. Spiral MALDI-TOF of MAMES isolated from Δ Msd_{esA1}.
(PDF)

S2 Fig. Spiral MALDI-TOF of MAMES isolated from $\Delta MsdesA1$.
(PDF)

S1 Table. Spiral MALDI-TOF of MAMES isolated from $\Delta MsdesA1$.
(PDF)

S2 Table. Spiral MALDI-TOF of MAMES isolated from $\Delta MsdesA1$.
(PDF)

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Supervision: AB GSB DM.

Writing – original draft: AB NF.

Writing – review & editing: AB NF.

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