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# Overexpression of BUNDLE SHEATH DEFECTIVE 2 improves the efficiency of photosynthesis and growth in *Arabidopsis*

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## SUMMARY

Bundle Sheath Defective 2, BSD2, is a stroma-targeted protein initially identified as a factor required for the biogenesis of ribulose 1,5-bisphosphate carboxylase/oxygenase (RuBisCO) in maize. Plants and algae universally have a homologous gene for BSD2 and its deficiency causes a RuBisCO-less phenotype. As RuBisCO can be the rate-limiting step in CO<sub>2</sub> assimilation, the overexpression of BSD2 might improve photosynthesis and productivity through the accumulation of RuBisCO. To examine this hypothesis, we produced BSD2 overexpression lines in *Arabidopsis*. Compared with wild type, the BSD2 overexpression lines *BSD2ox-2* and *BSD2ox-3* expressed 4.8-fold and 8.8-fold higher BSD2 mRNA, respectively, whereas the empty-vector (EV) harbouring plants had a comparable expression level. The overexpression lines showed a significantly higher CO<sub>2</sub> assimilation rate per available CO<sub>2</sub> and productivity than EV plants. The maximum carboxylation rate per total catalytic site was accelerated in the overexpression lines, while the number of total catalytic sites and RuBisCO content were unaffected. We then isolated recombinant BSD2 (rBSD2) from *E. coli* and found that rBSD2 reduces disulfide bonds using reductants present *in vivo*, for example glutathione, and that rBSD2 has the ability to reactivate RuBisCO that has been inactivated by oxidants. Furthermore, 15% of RuBisCO freshly isolated from leaves of EV was oxidatively inactivated, as compared with 0% in BSD2-overexpression lines, suggesting that the overexpression of BSD2 maintains RuBisCO to be in the reduced active form *in vivo*. Our results demonstrated that the overexpression of BSD2 improves photosynthetic efficiency in *Arabidopsis* and we conclude that it is involved in mediating RuBisCO activation.

**Keywords:** *Arabidopsis thaliana*, BSD2, disulfide bonds, oxidative stress, protein disulfide reductase, redox, RuBisCO, Zn finger domain.

## INTRODUCTION

The first step of CO<sub>2</sub> fixation is catalyzed by ribulose 1,5-bisphosphate carboxylase/oxygenase (RuBisCO) in the chloroplast stroma, and is often the rate-limiting step of photosynthesis in the natural environment (Sage *et al.*,

2008). RuBisCO is required in high amounts in C<sub>3</sub> plant chloroplast due to its low catalytic turnover rate and limited specificity for CO<sub>2</sub> versus O<sub>2</sub> (Parry *et al.*, 2013). Therefore, RuBisCO has been targeted to be artificially modified to improve photosynthesis and plant productivity.

However, manipulations of RuBisCO in either catalytic properties or quantity are not straightforward partly because putting 16 protein subunits together ( $L_8:S_8$ ; eight large and eight small subunits) in one RuBisCO is a complex task requiring numerous chaperones (Whitney *et al.*, 2011; Aigner *et al.*, 2017). Furthermore, auxiliary factors are also required for the activation of mature RuBisCO, imposing additional limitations on the apparent catalytic performance (Portis, 2003; Sage *et al.*, 2008).

RuBisCO activity is determined in a light-dependent manner and is active in the light and inactive in the dark. Sequential binding of  $CO_2$  and  $Mg^{2+}$  is a pre-requisite for RuBisCO catalysis (Portis, 2003). This process is inhibited by a range of sugar phosphates that tightly bind to the active site in the large subunit. Upon illumination, ATP-dependent RuBisCO activase promotes RuBisCO activation by releasing these ligands when the stromal ATP/ADP ratio increases with photophosphorylation (Scales *et al.*, 2014). Active photosynthetic electron transport also triggers the release of  $Mg^{2+}$  from the thylakoid to the stroma pool, thereby facilitating RuBisCO activation.

Aside from its regulation by carbamylation, RuBisCO activity is also redox regulated. It has been shown that reactive oxygen species (ROS), such as hydrogen peroxide ( $H_2O_2$ ), inactivate RuBisCO (Moreno *et al.*, 2008). The production of ROS is accelerated in conditions where the absorbed light energy is excessive for photosynthesis, for example during environmental stress. Therefore, it is assumed that RuBisCO tends to be inactivated by ROS under such stressful conditions (Zaffagnini *et al.*, 2012). In laboratory experiments, the oxidatively inactivated RuBisCO has been demonstrated to be reactivated by the addition of reducing agents, for example dithiothreitol (DTT), indicating that this inactivation is reversible (Sudhani and Moreno, 2015). However, it is unknown if oxidatively inactivated RuBisCO is reactivated *in vivo* or if any enzyme is involved in this process.

BUNDLE SHEATH DEFECTIVE2 (BSD2), a small stroma-targeted protein, was initially identified as an essential factor for RuBisCO biogenesis in maize (Brutnell *et al.*, 1999), and now appears to be universal in green algae and plants (Doron *et al.*, 2014; Aigner *et al.*, 2017). BSD2-deficient plants cannot accumulate the large subunit of RuBisCO despite its active translation and transcription (Brutnell *et al.*, 1999; Wostrickoff and Stern, 2007). Several studies have proposed that BSD2 acts in post-translational modification of nascent large subunit or RuBisCO assembly (Brutnell *et al.*, 1999; Wostrickoff and Stern, 2007; Doron *et al.*, 2014; Aigner *et al.*, 2017). The recent success in recombinant expression of plant RuBisCO in *Escherichia coli* identified BSD2 as the late-stage assembly factor which seems to stabilize the premature  $L_8$  complex (Aigner *et al.*, 2017). This view is consistent with the protein annotation of BSD2 homologues, which uniquely conserve the cysteine

(Cys) rich Zn finger domains of the DnaJ chaperone in *E. coli* (Brutnell *et al.*, 1999).

As BSD2 was shown to be involved in the production and accumulation of RuBisCO in plants it is conceivable that overexpression of BSD2 improves photosynthesis and productivity through accumulating RuBisCO. Conlan *et al.* (2018) recently showed that overexpressing BSD2 in tobacco chloroplasts did not alter RuBisCO content, activation status, leaf photosynthesis rate, or plant growth. Here, we produced BSD2-overexpressing lines, *BSD2ox-2* and *BSD2ox-3*, in Arabidopsis. Our results demonstrate that the overexpression of BSD2 increases the  $CO_2$  assimilation rate and plant biomass by increasing the maximum carboxylation rate by increasing the proportion of active catalytic sites, but not by increasing the number of total catalytic sites or RuBisCO content. To understand its mechanism, we isolated recombinant BSD2 (rBSD2) from *E. coli* and found that BSD2 has the ability to reactivate oxidatively inactivated RuBisCO *in vitro*. In this study, we examined how overexpression of BSD2 improves photosynthesis and productivity and discuss a physiological function of BSD2 in plants. Our results demonstrated that BSD2 is a key component in the mechanism to reactive oxidized RuBisCO, thereby maintaining photosynthetic efficiency in plants.

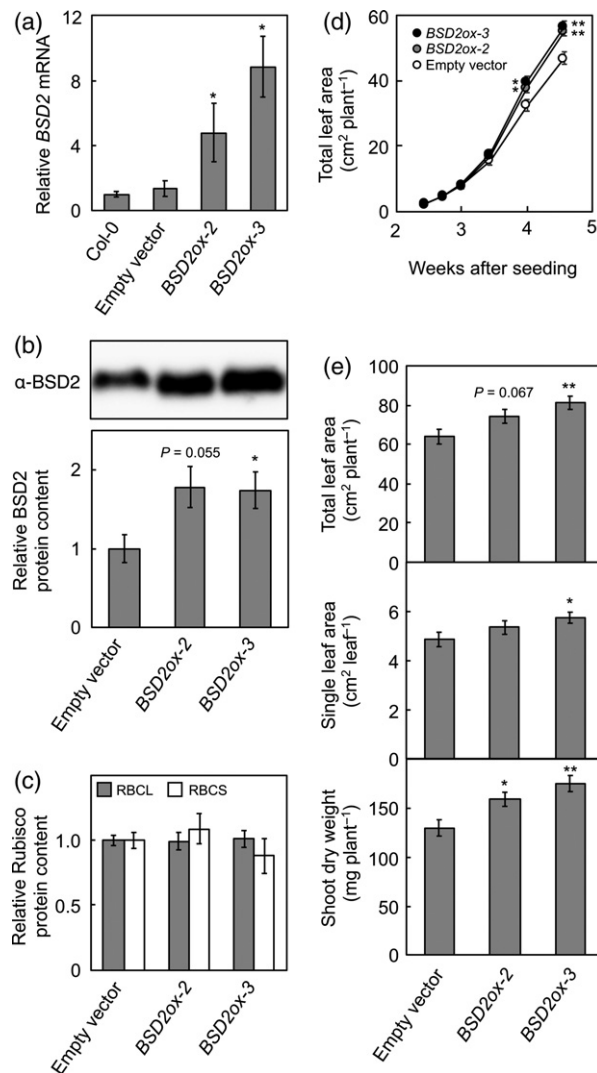
## RESULTS

### Overexpression of BSD2 improves photosynthetic efficiency and increases plant biomass

The full-length open reading frame of BSD2 cDNA was used to generate an overexpression vector pG2CYO2. Following floral dipping, transgenic Arabidopsis plants were selected on both kanamycin-containing and hygromycin-containing medium (Nakagawa *et al.*, 2007). We isolated 15 BSD2-overexpressing lines, and the two lines (*BSD2ox-2* and *BSD2ox-3*) showing the highest levels of BSD2 (At3g47650) transcripts at T3 generation were selected for this study. The levels of BSD2 transcripts in *BSD2ox-2* and *BSD2ox-3* were 4.8-fold and 8.8-fold higher than in the wild type, respectively, and there was no significant difference between wild type and the empty-vector control (Figure 1a). Along with their higher BSD2 gene expression, the *BSD2ox* lines accumulated 74–78% more BSD2 protein in leaves than the control plants (Figure 1b). Conversely, the results of SDS-PAGE analysis indicated the amounts of both RuBisCO large and small subunits were comparable between the *BSD2ox* and control leaves (Figure 1c; Figure S1). Nevertheless, the *BSD2ox* lines increased their leaf area relative to the control (Figure 1d; Figure S2). The increased leaf area was due to an increase in the size of individual leaves (Figure 1e; Figure S3) but not the number of leaves (Figure S4). Eventually, the *BSD2ox* lines had a 1.23–1.35-fold larger shoot dry mass than the control at 35

days after seeding (Figure 1e). In gellan gum-grown plants, the shoot/root ratio was similar between the *BSD2ox* lines and control (Figure S5), suggesting that the carbon-partitioning was unaltered.

To see the effect of overexpression of *BSD2* on photosynthesis, CO<sub>2</sub> assimilation rate per leaf area (*A*) in response to varying CO<sub>2</sub> concentrations was measured under saturating irradiance. The response of *A* to the CO<sub>2</sub> concentration in either the intercellular airspace (*C<sub>i</sub>*;

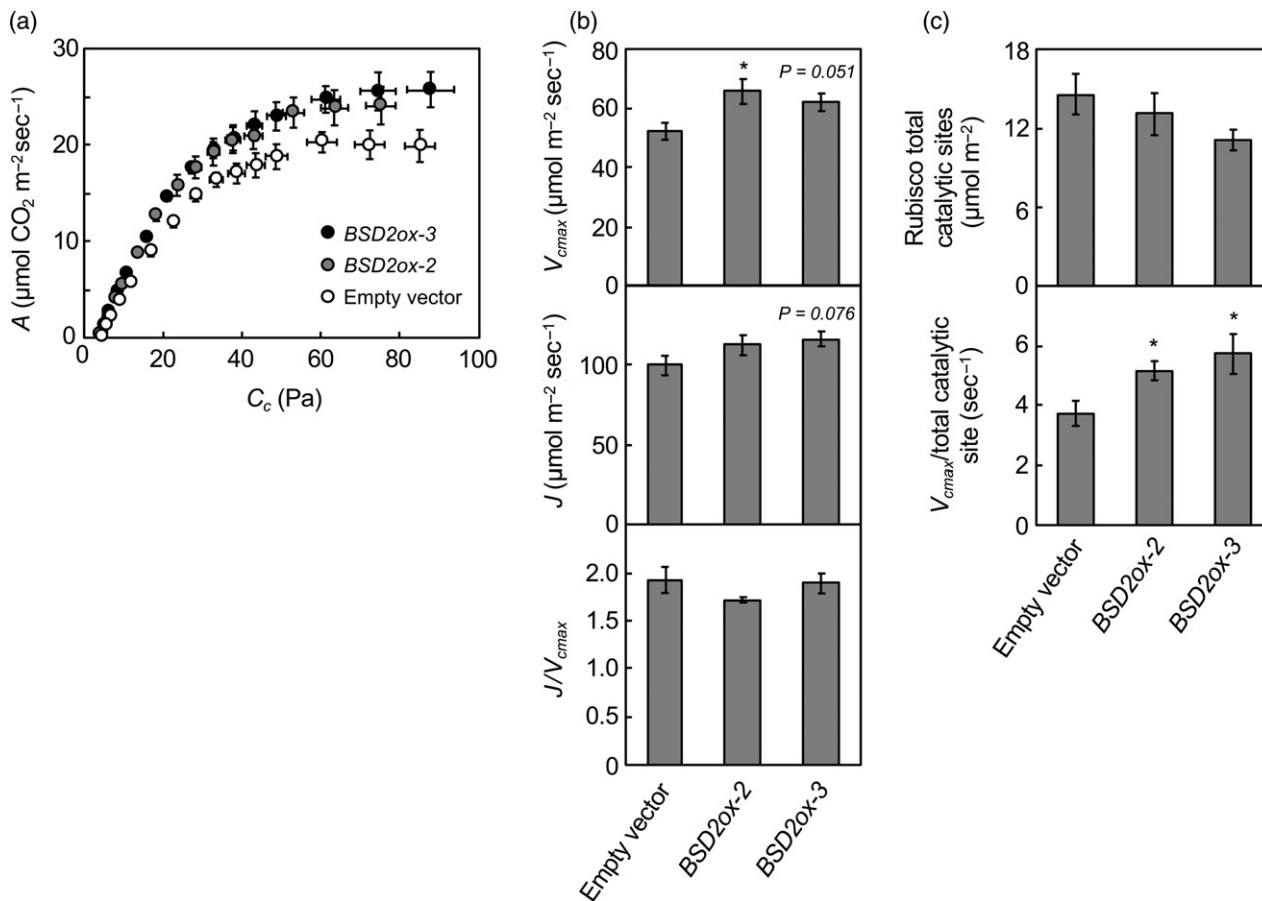


**Figure 1.** Gene expression, protein accumulation, and growth in wild type (Col-0) and transgenic plants transformed either with an empty vector (*Empty vector*) or the *BSD2* overexpression vector (*BSD2ox-2*, *BSD2ox-3*). (a) Expression of *BSD2* gene transcript in leaves of 3-week-old transgenic lines relative to Col-0 (mean  $\pm$  SE;  $n = 4$ ). (b) Western blotting analysis for BSD2 protein in leaves of 3-week-old plants. Values relative to Col-0 are shown (mean  $\pm$  SE;  $n = 3$ ). (c) SDS-PAGE analysis for large and small subunits of RuBisCO (RBCL and RBCS) in leaves of 3-week-old plants. Relative values to Col-0 are shown (mean  $\pm$  SE;  $n = 6$ ). (d) Change of total leaf area at 2 and 5 weeks after seeding (mean  $\pm$  SE;  $n = 7$ ). (e) Total leaf area per plant, average leaf area per individual leaf, and shoot dry weight of 35-day-old plants (mean  $\pm$  SE;  $n = 7$ ). *P*-values by Student's *t*-test are indicated if less than 0.1 (\* $P < 0.05$ , \*\* $P < 0.01$ ).

Figure S6) or at the site of carboxylation (*C<sub>i</sub>*; Figure 2a) showed greater photosynthetic capacity in the *BSD2ox* leaves. Fitting a photosynthetic model to these *A*-*C<sub>i</sub>* plots, we estimated the maximum RuBisCO carboxylation rate (*V<sub>cm</sub>*) and the photosynthetic electron transport rate (*J*) (Figure 2b). The *BSD2ox* leaves had higher *V<sub>cm</sub>* than control leaves by 19–26%. *J* also tended to be higher in the *BSD2ox* leaves, resulting in similar *J/V<sub>cm</sub>* ratios (Figure 2b). This indicates that photosynthetic electron transport capacity is increasing in parallel with *V<sub>cm</sub>* in the *BSD2ox* plants. It is therefore likely that the increase in biomass in the *BSD2ox* plants is caused by this increase in photosynthetic capacity. Furthermore, the number of RuBisCO total catalytic sites *per* leaf area in those leaves for the photosynthesis measurements was not significantly different between the overexpression lines and control (Figure 2c). Nevertheless, the apparent catalytic efficiency for RuBisCO carboxylation, estimated as *V<sub>cm</sub>* *per* total catalytic site, was higher in the *BSD2ox* lines (Figure 2c). This indicates that proportion of active catalytic sites to total catalytic sites of RuBisCO was increased in the *BSD2ox* leaves. Overall, *in vivo* catalytic efficiency rather than RuBisCO content was enhanced in the *BSD2ox* leaves, resulting in higher overall photosynthetic capacity and growth.

#### BSD2 catalyzes reduction of disulfide bonds

To understand how the overexpression of BSD2 enhanced the catalytic efficiency for RuBisCO carboxylation *in vivo*, we isolated rBSD2 protein from *E. coli* and studied its characteristics. BSD2 orthologues in green plants contain two Cys4-type Zn finger motifs (CXXCXGXG) (Brutnell *et al.*, 1999; Wostrickoff and Stern, 2007; Doron *et al.*, 2014), which show either chaperone activity or thiol-disulfide oxidoreductase activity in *E. coli* DnaJ (Tang and Wang, 2001). In chloroplasts, four other proteins likewise contain a DnaJ-like domain: CYO1/SCO2 (Shimada *et al.*, 2007; Albrecht *et al.*, 2008), LQY1 (Lu *et al.*, 2011) and HCF222 (Hartings *et al.*, 2017) in/at the thylakoid membrane, and PSA2 in the thylakoid lumen (Fristedt *et al.*, 2014). They all exhibit protein disulfide reductase (PDR) activity like DnaJ (de Crouy-Chanel *et al.*, 1995). We then tested whether rBSD2 also has PDR activity with a protein disulfide isomerase (PDI) assay (Raturi and Mutus, 2007; Muranaka *et al.*, 2012). In this assay, fluorescence increases if the reduction of disulfide bonds is mediated by diethylenetriamine (Di-E-GSSG). When purified rBSD2 and a non-specific reductant (i.e. DTT) was added to PDI assay solution containing Di-E-GSSG, the fluorescence signal significantly increased (Figure 3a). However, such an increase did not happen in the presence of DTT without rBSD2. These results demonstrate that rBSD2 has PDR activity, which is further supported by the dependency of the reaction rate on the substrate concentration (Figure 3b).



**Figure 2.** Photosynthesis in leaves of empty-vector control plants (*Empty vector*) and *BSD2*-overexpressing plants (*BSD2ox-2,3*). (a) Response of CO<sub>2</sub> assimilation (*A*) to chloroplastic CO<sub>2</sub> concentration in leaves (*C<sub>c</sub>*) (mean ± SE; *n* = 4–5). (b) Maximum RuBisCO carboxylation rate (*V<sub>cmax</sub>*) and photosynthetic electron transport rate (*J*) fitted to the *A*–*C<sub>c</sub>* response curves in (a), and the *J/V<sub>cmax</sub>* ratio. (c) Number of RuBisCO total catalytic sites per leaf area, and the maximum RuBisCO carboxylation rate (*V<sub>cmax</sub>*) per total catalytic site (mean ± SE; *n* = 4–5). Both *V<sub>cmax</sub>* in (b) and total catalytic sites in (c) were measured in the same leaves as used in (a). *P*-values by Student's *t*-test are indicated if less than 0.1 (\**P* < 0.05).

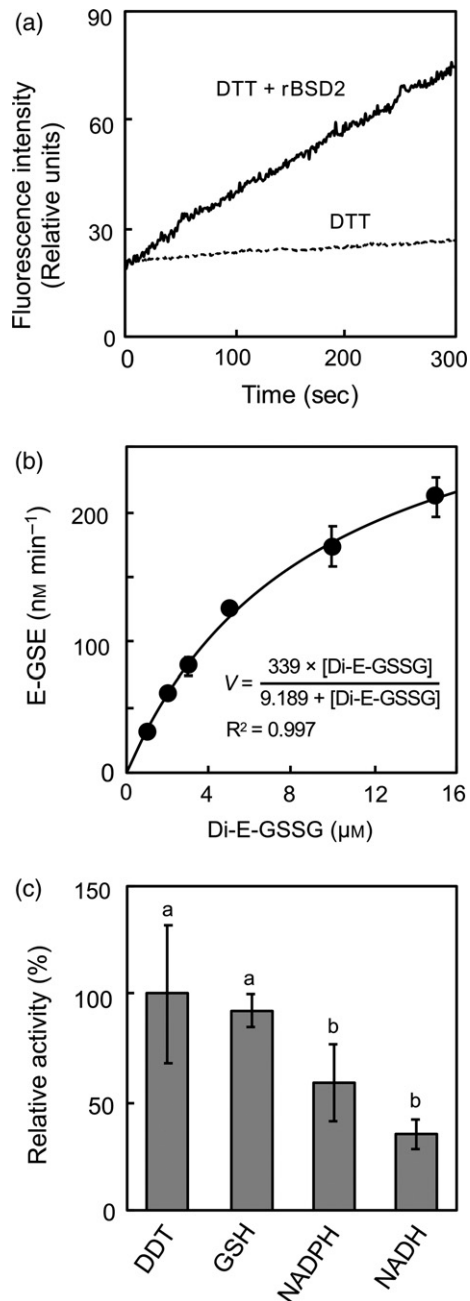
Each of CYO1, LQY1, HCF222 and PSA2 is associated with membrane-embedded photosynthetic complexes involved in thylakoid biogenesis/homeostasis (Lu *et al.*, 2011; Muranaka *et al.*, 2012; Tanz *et al.*, 2012; Fristedt *et al.*, 2014). Considering BSD2 is a stromal protein, we expect that the catalytic properties of BSD2 are different from the membrane-embedded proteins. We then estimated PDR kinetics of rBSD2 from the Michaelis–Menten curve (Figure 3b), as was done for the recombinant CYO1 in *Arabidopsis* (Muranaka *et al.*, 2012) and rice (Tominaga *et al.*, 2016). *K<sub>m</sub>* and *K<sub>cat</sub>* values for rBSD2 were 9.19 μM and 3.39 min<sup>-1</sup>, respectively, which are higher than those of CYO1 (Table 1), suggesting that BSD2 has a lower affinity for its substrate. This distinctive feature may reflect differences in accessibility to and/or abundance of their substrates in the highly crowded macromolecular solution.

PDR activity of rBSD2 relies on the presence of reductant. To examine whether rBSD2 can use biological reductants, we measured the PDR activity of rBSD2 in the presence of reduced glutathione (GSH), NADPH, or NADH.

Our results demonstrated that GSH is the most effective reductant among them and its efficiency is comparable to DTT (Figure 3c).

#### BSD2 reactivates oxidized RuBisCO

Previous studies have demonstrated that RuBisCO loses activity through the oxidation of its cysteine residues (Marcus *et al.*, 2003; Moreno *et al.*, 2008). We therefore tested whether BSD2 can reduce and thereby reactivate oxidized RuBisCO. Carboxylation rates of purified RuBisCO decreased by 60% after being oxidized with 5 mM H<sub>2</sub>O<sub>2</sub> (Figure 4a). We then tested the effect of rBSD2 on the recovery of the carboxylation rate in the presence of 1 mM DTT. In our experimental condition, the carboxylation rate did not change in the absence of rBSD2, although higher concentrations of DTT might be able to reactivate oxidized RuBisCO (Sudhani and Moreno, 2015). However, in the presence of rBSD2, the rate partially, but significantly, recovered (Figure 4b). Our results demonstrate that rBSD2 can reactivate oxidized RuBisCO.



**Figure 3.** PDR assay of rBSD2. (a) Change in fluorescence intensity after a solution containing 1  $\mu\text{M}$  Di-E-GSSG was incubated with 5  $\mu\text{M}$  DTT in the presence (solid line) or absence (broken line) of 100 nM rBSD2. (b) Michaelis–Menten curve for PDR kinetics with 5  $\mu\text{M}$  DTT.  $K_m$  and  $V_{\text{max}}$  of rBSD2 were estimated to be 9.189  $\mu\text{M}$  and 339  $\text{nM min}^{-1}$ , respectively (mean  $\pm$  SD;  $n = 3$ ). (c) Effect of 5  $\mu\text{M}$  reductants on the PDR activity. Values relative to DTT are shown (mean  $\pm$  SD;  $n = 3$ ). Different letters indicate significant difference among reductants (Tukey's test,  $P < 0.05$ ).

To examine whether the overexpression of BSD2 influences the redox state of RuBisCO in leaves, we isolated crude protein samples from leaves without adding reductants and subsequently measured the RuBisCO carboxylation rate after incubating the sample with or without DTT

**Table 1** Kinetic properties of BSD2 and CYO1 in *Arabidopsis thaliana* and *Oryza sativa*. Data for CYO1 were obtained in previous studies (Muranaka *et al.*, 2012; Tominaga *et al.*, 2016)

	Species	$K_m$ ( $\mu\text{M}$ )	$K_{\text{cat}}$ ( $\text{min}^{-1}$ )	$K_{\text{cat}} K_m^{-1}$ ( $\times 10^3 \text{M}^{-1} \text{sec}^{-1}$ )
BSD2	<i>A. thaliana</i>	$9.19 \pm 0.93$	$3.39 \pm 0.16$	$6.2 \pm 0.8$
CYO1	<i>A. thaliana</i>	$0.82 \pm 0.09$	$0.53 \pm 0.02$	$10.7 \pm 1.5$
	<i>O. sativa</i>	$3.16 \pm 0.88$	$0.53 \pm 0.05$	$2.8 \pm 0.4$

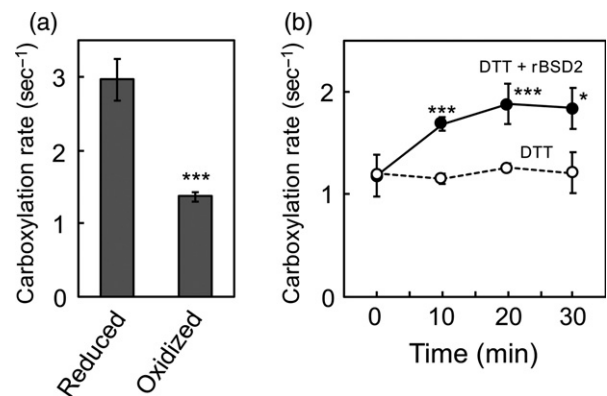
Means  $\pm$  SE ( $n = 3$ ).

for 30 min. In the empty-vector control, the RuBisCO carboxylation rate in samples incubated with DTT was 15% higher than when incubated without DTT, indicating that 15% of RuBisCO catalytic sites was inactive due to oxidation (Figure 5). Conversely, RuBisCO carboxylation rate in the BSD2 overexpression lines could not be increased by adding the reductant DTT, showing that there was no activity loss due to oxidation in the extracted protein samples (Figure 5). Our results demonstrated that RuBisCO from freshly isolated leaves consists of a higher proportion of the reduced active form in the BSD2 overexpression lines than in the control.

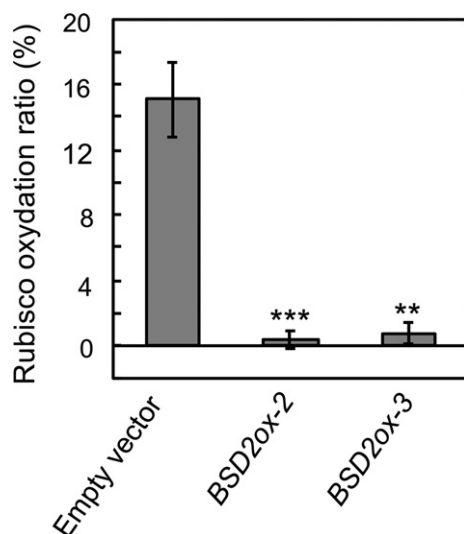
## DISCUSSION

Our results demonstrate that BSD2 overexpression improves both photosynthesis and productivity in our study organism *Arabidopsis*. BSD2 overexpression did not change the RuBisCO content or the number of total catalytic sites, but improved the *in vivo* maximum carboxylation efficiency probably through increasing the pool of reduced RuBisCO.

Incubation of isolated RuBisCO in oxidants, including ROS, decreases RuBisCO activity (Marcus *et al.*, 2003). The



**Figure 4.** RuBisCO reactivation assay. (a) RuBisCO carboxylation rate after incubation with 1 mM DTT (reduced) or 5 mM  $\text{H}_2\text{O}_2$  (oxidized) for 30 min (mean  $\pm$  SD;  $n = 3$ ). (b) Change in carboxylation rate of the pre-oxidized 80 nM RuBisCO in the presence (closed circles) or absence (open circles) of 40 nM rBSD2 (mean  $\pm$  SD;  $n = 3$ ). ( $*P < 0.05$ ,  $***P < 0.001$ , by Student's *t*-test).



**Figure 5.** Redox-dependent inactivation of RuBisCO. The RuBisCO oxydation ratio was calculated as a remaining proportion of the RuBisCO carboxylation rate estimated without DTT present (w/o DTT) relative to that with DTT present (w DTT)  $\left[ \left( 1 - \frac{w/o DTT}{w DTT} \right) \times 100 \right]$ . Means  $\pm$  SE ( $n = 3$ ) are shown. (\*\* $P < 0.01$ , \*\*\* $P < 0.001$ , by Student's *t*-test).

oxidatively inactivated RuBisCO has been demonstrated *in vitro* to be reactivated by reductants, such as DTT (García-Ferris and Moreno, 1993; Marcus *et al.*, 2003; Sudhani and Moreno, 2015). In the present study, we demonstrated that rBSD2 can reactivate the oxidatively inactivated RuBisCO and can act as a catalyst for this reduction. As a consequence, photosynthetic efficiency is improved despite no observed increase in absolute RuBisCO content. This is consistent with results showing that the overexpression of BSD2 increases the apparent carboxylation efficiency of RuBisCO (Figure 2) and that the reduced form of RuBisCO was more abundant in the BSD2 overexpression lines than in the control (Figure 5). However, we need to note that the effect of overexpression of BSD2 on the RuBisCO catalysis and photosynthesis might differ among plants and environments, depending on the sensitivity of RuBisCO to the oxidants, such as ROS. Indeed, the overexpression of BSD2 has not improved photosynthesis in tobacco (Conlan *et al.*, 2018).

#### PDR activity of BSD2 reactivates oxidatively inactivated RuBisCO

Oxidation of RuBisCO by oxidants, including ROS, inactivates RuBisCO, but its mechanism remained unknown. However, as RuBisCO possesses cysteine residues that are sensitive to oxidants, it is conceivable that the oxidation of cysteine residues is associated with the inactivation of RuBisCO. Importantly, RuBisCO oxidized by disulfides such as cystamine can be reactivated by thiols like DTT and cysteamine (García-Ferris and Moreno, 1993; Marcus *et al.*, 2003; Sudhani and Moreno, 2015). Therefore, the

inactivation of RuBisCO by oxidants is reversible, most likely through the reduction of oxidized cysteine residues. In the present study, we showed that rBSD2 has PDR activity, suggesting that reactivation of oxidatively inactivated RuBisCO by BSD2 is associated with the direct thiol transactions between oxidized RuBisCO and BSD2. In practice, this reaction occurs only if effective reductants are available at the site of RuBisCO (i.e. in the stroma). We confirmed that glutathione, NADPH, and NADH, which also exist in the stroma, could initiate the PDR reaction and that glutathione was the most effective of these reductants (Figure 3c). A similar preference for reductants has also been observed in CYO1 (Muranaka *et al.*, 2012; Tominaga *et al.*, 2016). It is known that RuBisCO is a potential target of both thioredoxins (Balmer *et al.*, 2004; Lemaire *et al.*, 2004) and glutaredoxins (Zaffagnini *et al.*, 2012). Although thioredoxins can target vicinal Cys pairs of RuBisCO, they are unlikely to be involved in the reactivation as these disulfides are not critical for the catalytic activity (Ranty *et al.*, 1991; Moreno and Spreitzer, 1999; Marín-Navarro and Moreno, 2006), and the concentration of thioredoxins is not sufficient to reduce the large pool of RuBisCO (Peltier *et al.*, 2006; Sudhani and Moreno, 2015). Conversely, glutathione and ascorbate are very abundant in the chloroplast stroma ranging between 0.5–3.5 and 20–300 mM, respectively, thereby constituting a pool of redox buffers against oxidants (Foyer and Noctor, 2011; Dietz *et al.*, 2016). In contrast with the light-dependent ferredoxin–thioredoxin system, the glutathione pool (i.e. GSH/GSSG ratio) may not be oxidized very much when photosynthetic electron transport is low (e.g. in the dark, under stress conditions, and during senescence) owing to decreased metabolic demands and catabolic supply of reducing equivalents (Foyer and Noctor, 2011). Therefore, chloroplastic glutathione would allow BSD2 to continuously reduce, and thereby maintain RuBisCO to be in the active form (Sudhani and Moreno, 2015). It is noteworthy that a drop in the GSH/GSSG ratio in response to excess ROS may cause a translational arrest of the RuBisCO large subunit, related to oxidation of the nascent polypeptide (Irihimovitch and Shapira, 2000).

In our experiments, rBSD2 reactivated oxidatively inactivated RuBisCO, but the RuBisCO activity did not recover to the initial level (Figure 4). This result suggests that the oxidatively inactivated RuBisCO includes reversible and irreversible states. Previous studies have demonstrated that the oxidation of RuBisCO decreases the number of catalytic sites, resulting in an irreversible deactivation (Marcus *et al.*, 2003). Furthermore, it has been demonstrated that ROS can cause direct fragmentation of the large subunit *in vitro* (Ishida *et al.*, 1998) and in intact chloroplasts (Nakano *et al.*, 2006). The unrecoverable activity of H<sub>2</sub>O<sub>2</sub>-oxidized RuBisCO after BSD2 treatment (Figure 4b) might therefore represent either irreversible

conformational change or decomposition of the RuBisCO complex.

### Physiological function of BSD2 *in vivo*

It has been widely accepted that BSD2 is a critical chaperone in RuBisCO assembly in plants and green algae. During RuBisCO biogenesis BSD2 appears to function in preventing an end-state assembly intermediate from the aggregation until the small subunits become available (Brutnell *et al.*, 1999; Wostrickoff and Stern, 2007; Aigner *et al.*, 2017). It has also been suggested that BSD2 is a co-translational chaperone that protects nascent large subunit chains from misfolding (Doron *et al.*, 2014). Here, we confirmed the hypothesis that BSD2 may also target the post-assembled RuBisCO and reactivate oxidatively inactivated RuBisCO. It has been frequently noted that oxidation of RuBisCO is physiologically relevant to senescence or stress scenarios, which are known to trigger a fast catabolism of RuBisCO (García-Ferris and Moreno, 1993; Moreno and Spreitzer, 1999; Marcus *et al.*, 2003; Marín-Navarro and Moreno 2003). The reactivation of RuBisCO by BSD2 might be therefore important for alleviating senescence or stress scenarios, as well as for maintaining functional photosynthesis.

## EXPERIMENTAL PROCEDURES

### Plant materials and growth conditions

The full-length open reading frame of BSD2 cDNA was used to generate an overexpression vector pG2CYO2. The full-length ORF of BSD2 cDNA was amplified using primers At3g47650-F-attB1 (5'-AAAAGCAGGCTATATGGCGAATTCATGCTTCTTCTTCTCC-3') and At3g47650-attB2 (5'-AGAAAGCTGGGTACTCATGAAGGTGCTCAAGAACCACC-3'). The resulting amplified product was cloned into pDONR/Zeo (Invitrogen, ThermoFisher, Carlsbad, CA, USA) to make pD3gORFull, and the sequence was confirmed. The full-length ORF was introduced into the pGWB2 Gateway vector (Nakagawa *et al.*, 2007) by recombination from the pD3gORFull to make pG2BSD2. The pG2BSD2 vector was introduced into wild type *Arabidopsis thaliana* (Columbia-0) by floral dipping (Clough and Bent, 1998) using *Agrobacterium tumefaciens* GV3101. Fifteen transgenic lines were obtained, and T3 was used for the study.

Plants were grown at 22–23°C, 60–85% (relative humidity) under a 16-h fluorescent light (100  $\mu\text{mol m}^{-2} \text{sec}^{-1}$ )/8-h dark cycles on 1/2MS plates or in soil.

### Photosynthesis measurements

Photosynthetic gas exchange of 5-week old Col-0 and BSD2 overexpression plants were measured using a Licor 6400XT gas exchange system equipped with a leaf chamber fluorometer (LICOR, Lincoln, Nebraska USA). Measurements were performed at a leaf temperature of 25°C and a VPD of 1 kPa. CO<sub>2</sub> response curves were measured at a saturating light intensity of 1200  $\mu\text{mol m}^{-2} \text{sec}^{-1}$ . The curves were fitted for  $V_{\text{cmax}}$ ,  $J$ , and  $g_m$  with the Excel fitting tool provided by Sharkey *et al.* (2007), which was parameterized with the RuBisCO kinetic parameters of Arabidopsis described elsewhere (Walker *et al.*, 2013). CO<sub>2</sub> concentrations at the site of carboxylation ( $C_c$ ) were calculated according to Fick's law as  $C_c = C_i - A/g_m$ .

### SDS-PAGE and immunoblot analysis

Rosette leaves (100 mg fresh weight) from each 3-week-old plant were ground with a mortar and pestle in liquid nitrogen and transferred to a new 1.5-ml tube and homogenized in 5 volumes (v/w) of extraction buffer containing 15 mM Tris-HCl, pH 8.0, 50 mM NaCl, 10 mM ethylenediaminetetraacetic acid (EDTA) and 1.0% protease inhibitor cocktail (Sigma-Aldrich, Tokyo, Japan). The samples were incubated on ice 10 min with vigorous vortexing every minute and centrifuged (20 000  $g$ , 20 min, 4°C). The supernatant equivalent of 1.5 mg of fresh weight was electrophoresed on a 16.5% Tris-tricine SDS-PAGE gel and electroblotted onto polyvinylidene difluoride (PVDF) membrane. The membrane was immunoreacted with anti-BSD2 antibody (a kind gift from Spencer M. Whitney, Australian National University) and detected using an ECL Select kit (GE Healthcare, Chicago, IL, USA). Ribulose 1,5-bisphosphate carboxylase/oxygenase subunits RBCL and RBCS were visualized by staining the gel with Coomassie Brilliant Blue R250.

### Expression and purification of the rBSD2 protein

Truncated BSD2 cDNA was amplified by reverse transcriptase polymerase chain reaction (RT-PCR) using primer that contained sites for *NdeI* (At3g47650-*Nde*, 5'-GGGCATATGGCCGCAACAA TAATCCTCAGGGCACTAAACC-3') and *XhoI* (At3g47650-*Xho*, 5'-CCCTCGAGCTCATCGAAGGTGCTCAAGAACCACC-3'). The amplified DNA fragments encoded the BSD2 protein containing amino acids 57 to 136. The DNA fragments were digested with *NdeI* and *XhoI* and ligated into the expression vector pET-24b(+) (Novagen, Sigma-Aldrich, St. Louis, MD, USA). The sequence of the DNA fragment was confirmed. The *E. coli* strain BL21(DE3) was used for expression of the truncated BSD2 fusion proteins with 6× His tags. Overnight cultured BL21(DE3) cells harbouring the truncated BSD2 gene were diluted 1:20 with fresh culture medium and grown at 37°C for 1 h. Isopropyl- $\beta$ -D-thiogalactopyranoside was then added to a final concentration of 1.0 mM, and the cells were cultured at 20°C overnight. The cells were centrifuged for 10 min at 5000  $g$  and 4°C. The cell pellets were suspended in suspension buffer (15.0 mM Tris-HCl, pH 7.0, 50.0 mM NaCl, 0.1 mM DTT, and 1.0 mM phenylmethylsulfonyl fluoride). The cells were sonicated and the crude extract centrifuged for 10 min at 12 000  $g$  and 4°C. The supernatant was incubated for 1 h with nickel-nitrilotriacetic acid agarose (Qiagen, Venlo, the Netherlands) at 4°C. The agarose was washed with suspension buffer containing 30 mM imidazole. rBSD2 protein bound to the agarose was eluted with suspension buffer containing 200 mM imidazole. The eluted proteins were dialyzed with phosphate-buffered saline buffer at 4°C overnight.

### Assay for PDR-dependent disulfide reduction

Dieosin glutathione disulfide, Di-E-GSSG, was prepared as described in previous study (Muranaka *et al.*, 2012). PDI disulfide reduction assay was conducted in PDI assay buffer (100 mM potassium phosphate, pH 7.0) containing 100 nM rBSD2 and Di-E-GSSG (100–1500 nM) with 5  $\mu\text{M}$  DTT. The increase in fluorescence at 545 nm was monitored with excitation at 525 nm. The kinetic parameters were calculated using KaleidaGraph software (Synergy Software, Reading, PA, USA).

### RuBisCO purification and assay

Leaves from 4-week-old Arabidopsis plants were frozen in liquid nitrogen, and RuBisCO was purified by ion-exchange chromatography (Sharwood *et al.*, 2008) as follows. The frozen leaves were homogenized in glass homogenizer in ice-cold 50 mM Tris-HCl pH



7.6, 1 mM EDTA, 1% (w/v) polyvinylpyrrolidone, 5 mM DTT, plant protease inhibitor cocktail (Sigma, St. Louis, MD, USA, Product # P9599; 200  $\mu$ l per 25 ml solution), then centrifuged at 16 000 *g* for 15 min at 4°C. Next, the supernatant was added to a 5-ml Bio-Scale Mini Macro-prep High Q cartridge (Bio-Rad, Hercules, CA, USA) equilibrated with 50 mM Tris-HCl, pH 7.6, 1 mM EDTA under the control of an AKTA chromatography system. The elution system consisted of a 35-ml gradient of a 0–1 M NaCl at a flow rate of 2 ml min<sup>-1</sup>, and 2-ml fractions were collected. Fractions containing RuBisCO activity (determined by the <sup>14</sup>CO<sub>2</sub> RuBisCO activity assay described below) were pooled and dialyzed overnight at 4°C against 1 L of 50 mM Tris-HCl, pH 7.6, 1 mM EDTA, 20% (v/v) glycerol. Aliquots were frozen in liquid nitrogen and stored at –80°C. Before the RuBisCO activity assay, purified RuBisCO was activated in 50 mM EPPS-NaOH, pH 8.0, 15 mM NaHCO<sub>3</sub>, 15 mM MgCl<sub>2</sub> for 10 min, and then duplicate aliquots were used to measure RuBisCO activity by the [<sup>14</sup>C]CABP assay (10 or 30  $\mu$ M) (Ruuska *et al.*, 1998) in the presence of 1 mM DTT or 5 mM H<sub>2</sub>O<sub>2</sub> for 30 min to reduce or oxidize RuBisCO, respectively. To minimize the effects of excess DTT or H<sub>2</sub>O<sub>2</sub>, each reaction solution was diluted 20-fold with 50 mM EPPS-NaOH, pH 8.0, 20 mM MgCl<sub>2</sub>, 20 mM NaHCO<sub>3</sub>, 1 mM EDTA and then concentrated on an Amicon Ultra-15 Centrifugal Filter Unit (Merck Millipore, Burlington, MA, USA). The dilution/concentration process was repeated before the use.

The carboxylation rate was measured with the NADH-coupled spectrophotometric assay at 25°C by measuring the absorption at 340 nm with spectrophotometer (Sharwood *et al.*, 2016). The final concentrations of purified RuBisCO or rBSD2 in the assay solution were 80 nM (i.e. 10 nM per RuBisCO active site) and 40 nM, respectively. The assay was initiated by adding 0.5 mM ribulose 1,5-bisphosphate purified according to Kane *et al.* (1994) and Sharwood *et al.* (2016).

To estimate the ratio of oxidatively inactive to active RuBisCO in leaf, the frozen leaf was homogenized in glass homogenizer in ice-cold 50 mM Tris-HCl pH 7.6, 1 mM EDTA, 1% (w/v) polyvinylpyrrolidone, plant protease inhibitor cocktail (200  $\mu$ M/25 ml solution) without DTT, and centrifuged at 16 000 *g* for 15 min at 4°C. The supernatants were incubated with or without 5 mM DTT (final concentration) on ice for 30 min, and RuBisCO carboxylation rate was measured with NADH-coupled spectrophotometric assay (Sharwood *et al.*, 2016) but without adding DTT.

## DATA STATEMENT

All relevant data are provided within the manuscript and its supporting materials.

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## AUTHOR CONTRIBUTION

ST and HS designed research; FAB, JT, MM, NS, ST, TK, KN, EM, CH, AM, and HS performed research; FAB, JT, ST, WY, SEM, YO, TT, SD, TK, WS, AS, and HS analyzed data; FAB, JT, ST, WY, SEM, YT, TT, WW, TK, WS, AS, and HS wrote the article.

## CONFLICT OF INTEREST

The authors declare no conflict of interest.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article.

**Figure S1.** SDS-PAGE gel for RBCL and RBCS in leaves.

**Figure S2.** Measurements of total leaf area in Figure 1(d).

**Figure S3.** Comparisons of single leaf length and area in Figure 1(e).

**Figure S4.** Number of leaves in Figure 1(e).

**Figure S5.** Growth of 3-week-old plants on half-strength Murashige and Skoog medium plates.

**Figure S6.** Response of *A* to various *C*<sub>i</sub> in leaves.

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