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Opinion Paper

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The diagnostic potential and barriers of microbiome based therapeutics

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Abstract: High throughput technological innovations in the past decade have accelerated research into the trillions of commensal microbes in the gut. The ‘omics’ technologies used for microbiome analysis are constantly evolving, and large-scale datasets are being produced. Despite of the fact that much of the research is still in its early stages, specific microbial signatures have been associated with the promotion of cancer, as well as other diseases such as inflammatory bowel disease, neurodegenerative diseases etc. It has been also reported that the diversity of the gut microbiome influences the safety and efficacy of medicines. The availability and declining sequencing costs has rendered the employment of RNA-based diagnostics more common in the microbiome field necessitating improved data-analytical techniques so as to fully exploit all the resulting rich biological datasets, while accounting for their unique characteristics, such as their compositional nature as well their heterogeneity and sparsity. As a result, the gut microbiome is increasingly being demonstrating as an important component of personalised medicine since it

not only plays a role in inter-individual variability in health and disease, but it also represents a potentially modifiable entity or feature that may be addressed by treatments in a personalised way. In this context, machine learning and artificial intelligence-based methods may be able to unveil new insights into biomedical analyses through the generation of models that may be used to predict category labels, and continuous values. Furthermore, diagnostic aspects will add value in the identification of the non invasive markers in the critical diseases like cancer.

Keywords: biomarker; diagnostics; machine learning; microbiota.

Background

Technological innovations in the past decade have accelerated research into the trillions of commensal microbes in the gut, the microbiome. Across gut, inflammatory, and neurological disorders, there are alterations in gut microbiome composition and function [1, 2]. As a result, a plethora of machine learning approaches are increasingly being developed targeting the microbiome due to its diagnostic potential [3]. While the majority of these machine learning studies are critically flawed [4], the microbiome remains a potentially invaluable source for identifying novel biomarkers [5].

Culture-based diagnostics for monitoring

A simple, albeit limited, diagnostic approach involves selectively-culturing fecal samples to monitor antibiotic-resistant bacteria, such as *Enterococci* [6]. However, this approach is limited to well-identified organisms that are easily cultured [5] and have been successfully demonstrated to aid the diagnosis of gastrointestinal infections [7, 8]. For many exploratory studies, culture-based methods are far too limited in their scope but could provide additional functional information that can form valuable components for developing novel machine learning-based analysis approaches.

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16S rRNA diagnostics

The availability and declining cost of sequencing has rendered RNA-based diagnostics more common in the microbiome field. Amplifying and sequencing conserved bacterial ribosomal regions results into microbiome diversity and composition high resolution data [9, 10]. Such datasets have the potential of aiding prediction approach to depict patients that are likely to develop bacterial-resistant infections; for example, *Morganella* and *Prevotella* abundance conferred resistance [7]. 16S rRNA sequencing offers the tantalising possibility of catering the development of models that cater the prediction subsequent infection with antibiotic-resistant bacteria [11].

Beyond gastrointestinal disorders, several studies suggest that 16S microbiome biomarkers may be useful for differentiating Alzheimer's from dementia, as well as detecting schizophrenia, Parkinson's disease, inflammatory bowel disease, colorectal and other cancers, osteoporosis, bipolar disorder, and other immune-related diseases [12–22]. Other applications include predicting Crohn's disease relapse and drug responses [23, 24].

Metagenomic diagnostics

Metagenomic sequencing provides a more precise microbiome data resolution which can detect fungal and viral commensals [9, 25]. In addition to the increasingly accurate species-level resolution, it also detects specific genes that are being expressed in the microbiome, providing functional information [26]. Some studies leverage this extra layer of information to generate diagnostics, though it isn't always clear whether they provide a cost-effective advantage.

Thus far, several studies have employed metagenomic features or variables generated from experiments to diagnose early-stage colorectal cancer, different stages of Parkinson's disease, fatty-liver disease, schizophrenia, as well as other brain and gut disorders [27–31]. Importantly, future diagnostic metagenomic based approach may be able to facilitate the early detection of neurodegenerative diseases or depict particular patient drug-responses within psychiatric settings [32].

Metabolomic diagnostics

Fecal metabolomics provides more functional information that can be used to detect microbiome metabolites, such as short-chain fatty acids, tryptophan metabolites and secondary bile acids [33]. Since it is used less frequently

than other methods, it has been employed by fewer diagnostic applications compared to those utilising 16S or metagenomic data. Nevertheless, metabolomics datasets have been proven useful for differentiating healthy controls from patient cohorts suffering from certain cancers, myasthenia gravis, lupus, cardiovascular disease, liver disease, and other disorders [29, 34–37].

Outlook

The high number of features or variables within the microbiome, as well as its key role in physiology renders it an ideal source of diagnostic biomarkers across multiple diseases. Most studies select differentially abundant microbial features that are applied within diagnostic models. Predominantly, these microbial features are detected through 16S, whole genome metagenomics, or metabolomics. However, many studies still lack consistency and replicability and have yet to be validated in clinical trial settings.

Emergent problems in microbiome datasets

Both sequencing, as well as metabolomic, microbiome datasets are compositional nature [38–40]. Simply put, sequencing technology imposes an arbitrary detection limit of individual elements. Similarly, to a forest or other ecosystems, the abundance of one species or metabolite affects the abundance of other elements – prohibiting these elements to be treated as independent variables.

It follows that each detected element is present within this interdependent composition, rendering them more challenging to interpret and analyze [38, 41]. Thus we cannot consider this data as true counts of elements in the gut environment, but instead we need to contemplate them as a proportional count of molecules associated with microbes in the gut [38].

Normalization to account for sparsity and zeroes

The first step in analyzing any raw counts table involves normalizing the values it contains to allow for their comparison. However, typically, upon inspecting such data, it becomes obvious that it is sparse, and unevenly distributed. Moreover, such data usually contains several zeroes which need to be properly interpreted as true zeros or artefacts [38, 42].

Running robust multivariate comparisons involves generating Monte Carlo samples of the Dirichlet distribution within each sample before normalization [39, 43]. While many studies use rarefaction subsampling methods, these, typically, result in information loss across the overall composition [44]. Although, Ddta can be transformed using simple log-ratios, such an approach does not sufficiently address the sparsity and zeroes within the dataset [39, 40]. Tools, such as ALDEx2, allow users to perform central, isometric, or additive log ratios for normalization that conserves the relationships between elements better [39, 43].

Distance and ordination

Typically machine learning discrimination models attempt to separate two or more independent groups based on their unique features or variables. However, many common forms of ordination, typically employed in microbiome data analysis, employ non-compositional methods, such as UniFrac, Bray-Curtis, and Jensen-Shannon divergence [38, 41]. The Aitchison's distance is a more reliable method for measuring distance or volatility of compositions over time, providing a geometric measure of distance between principal components [41, 45].

Differential abundance and effect size

Following a multivariate statistics analysis, the abundance and effect size differences need to be determined. While 95% confidence intervals and effect size cut-offs may help identify salient features within a dataset, there is no specific cut-off to indicate clinical or real-world relevance [33]. It is unclear if it is necessary for some microbes or metabolites to reach a certain threshold before exerting strong effects. Small changes in other important microbes by contrast, could destabilize a microbial ecosystem if these microbes serve as important hubs within the network [46–50].

Correlation, causality and directionality

The compositional nature of microbiome datasets dictate that any correlations and comparisons are susceptible to negative correlation bias [38, 51, 52], an affect that necessitated the development of methods, such as SparCC and SpiecEasi [38]. While there aren't any robust methods for

determining causality and directionality, most research methods are now focusing on proportionality i.e., how much of a particular phenotype is explained by a feature within the dataset [53–59]. While Granger causality has also been suggested as a potential method for microbiome analysis, it is seldom used and yet to be validated [58]. It should finally be noted that whether any amount of data or any method development will ever suffice to determine causality is still questionable [60].

Outlook

The compositional nature of microbiome data imposes several challenges and biases that renders their analysis challenging. In addition to the sparsity and zeroes within the data, methods need to account for negative correlation bias, as well as many other uncertainties relating to the importance of different effect sizes.

Machine learning methods and workflows

There are many different types of machine learning methods, broadly split into supervised and unsupervised learning. In supervised learning, data is fully labelled whereas in unsupervised clustering, no data is labelled. Different types of semi-supervised approaches classify a small portion of the dataset for use as a test set.

Unsupervised clustering

Unsupervised clustering methods, such as K-means, have been implemented to characterize microbial features across stages of periodontitis [61]. Principal component analysis is a valuable approach to cluster compositional data, measuring distances based on the Aitchison's metric to determine whether there is a significant difference between two different groups of samples [38, 39, 41]. Non-negative matrix factorization and t-distributed stochastic neighbor embedding (t-SNE) are seldom used with microbiome datasets – though they may be appropriate for describing some cohorts [62].

For multifactorial disease datasets, such as depression related data, unsupervised clustering might provide powerful means for feature-based stratification that could indicate the best course of treatment.

Supervised learning

Supervised learning methods are far more common across microbiome studies and are especially useful in identifying associations between host phenotypes and microbial features [62]. These methods are used for classification or regression analyses.

Regression

Lasso, ridge, and elastic net are different types of penalized regression. These are especially useful when there are many more features within the data than sample size. The penalty function is important to prevent overinflation within the regression model [62]. The L1 penalty is used in lasso regression which shrinks the size of coefficients while setting the values of coefficients for unimportant features to zero; additionally, if two variables show collinearity, only one of these two variables will have a non-zero coefficient [63, 64].

In ridge regression, the L2 square root function penalty is used to shrink values near zero though it does not remove these variables [65]. Since the variables are included as non-zero coefficients, their effects are incorporated into the machine learning model. Meanwhile the lasso net method is a linear combination of lasso and ridge, retaining all features as distinct groups of variables [62, 66].

In the literature, regression methods are used across different microbiome studies to identify brain, immune, diabetic, and gastrointestinal disorders as well as cancers based on a host phenotype [19, 28, 67–71].

Classification

The support vector machine (SVM) approach employs linear or non-linear distances and margins to segregate different groups [72]. Linear applications use L1 and L2 penalty functions along with the SVM, thereby retrieving the most salient differentiating features and determining the maximal boundary between multiple groups, a task not performed by simple L1 or L2 regression. A comparison to common regression methods demonstrated that microbiome models using various SVMs, although outperforming random forest methods, don't always outperform simpler L2 regressions [73]. However, a scoping review of different machine learning techniques revealed an opposing trend, with random forest performing well against other types of models [74].

The random forest approach uses a re-sampling method, termed bootstrapping, along with random selection, to

generate different types of classifier decision trees [75]. It determines the variable importance of different features from these decision trees, allowing users to select the most salient metabolites or genes. When used appropriately, these models can detect features of colorectal cancer [76]. These pre-selected important features are also useful in future mechanistic studies.

Another popular method used within the microbiome literature involves linear discriminant analysis, part of the LefSe package [77]. It is similar to a supervised version of Principal Component analysis albeit inappropriate for compositional datasets [38].

Artificial neural networks and deep learning

Advances over the last decade allowed for bioinformatics approaches to employ advanced approaches, such as artificial neural networks. These networks relay an input layer of information, an activation function that combines the information, and finally an output function [62]. By manipulating the number of computational layers, or how many times data is passed back and forth between input and output, users can increase or decrease the complexity and power of the model. The goal of an artificial neural network is to find the ideal weights for different features to optimize classification [62, 78, 79].

Deep learning models, such as tensor flow, employ many more layers, allowing them to outperform other algorithms when handling large amounts of data [62]. This, however, comes at the expense of model and feature interpretability. Some tools also combine Bayesian prediction with deep learning, to assess whether some known host features, combined with microbiome data, can predict disease [80].

Reinforcement learning

Other machine learning methods employ rules instead of labels to guide the model development [81]. These methods could be harnessed for finding bacterial proteins or metabolites whose structures may fit human protein receptors.

Machine learning and AI workflows

Many python-based tools incorporate genomic data into reproducible workflows. The BioBakery workflow encompasses several different tools for metagenomic analysis developed within the Huttenhower lab; many of these tools

use linear discriminant analysis [82]. The individual programs and components within this workflow can also be adapted to integrate multi-omics data [83]. Microbiome Analyst is another example of a complete workflow albeit one that incorporates several tools that are inappropriate for compositional data analysis [84, 85].

DeepMicro converts high-dimensional data into low-dimensional representations thereby rendering their interpretation and analysis easier [86]. NFnetFu is a new pipeline demonstrated to perform well compared to other commonly used microbiome pipelines and tools, using a novel combination of approaches to process the data [87]. Briefly, an adaptive neuro fuzzy inference system pre-processes the data to overcome sparsity, even across smaller sample sizes, while maintaining the collinearity and relationships between features [87]. Then, a density-based clustering method is applied to reduce the collinearity into a simpler matrix for further analysis, followed by a lasso L1 regression [87]. Finally, it concludes with a taxon set enrichment analysis to identify relevant biological networks within the dataset [87]. A graphical overview is presented in the Figure 1.

Outlook

While there exist numerous different types of machine learning models and bioinformatics applications and frameworks, it is often difficult to select the best one without benchmarking them. Additionally, it is unclear where the increase of the model complexity leads to better predictions or more actionable insights. Among existing machine learning algorithms, linear discriminant analysis is most commonly used across microbiome studies to identify differential features between groups [77].

Fecal microbiota transplantation (FMT) as diagnostic process

Fecal microbiota transplantation (FMT) is the delivery of solutions of fecal material from donors entering the digestive system of a receiver in an attempt to actively affect the patient’s microbiological diversity and bestow a therapeutic advantage [88, 89]. FMT originated in Chinese

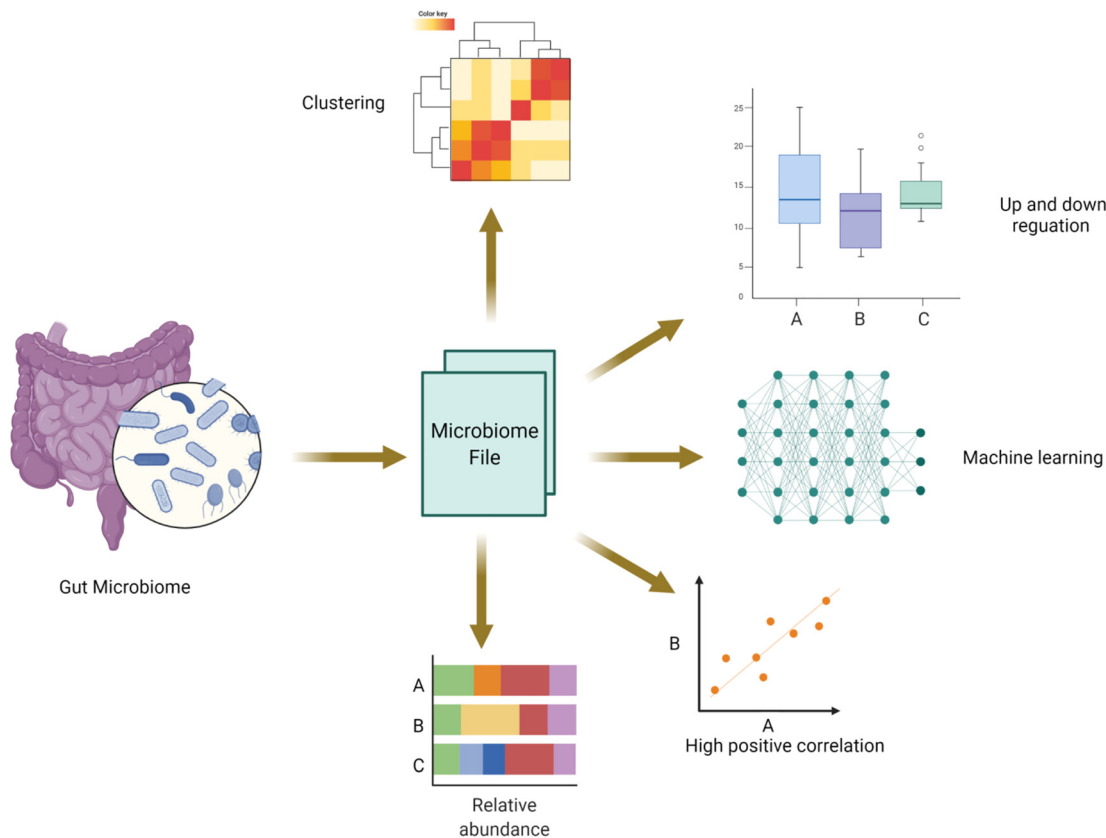


Figure 1: Machine learning-based analysis approaches employing microbiome datasets.

civilization over 1,700 years ago in the 4th century, when a well conventional Chinese medical practitioner entitled Ge Hong effectively cured sufferers of foodborne poisoning and/or chronic diarrhea via oral utilizing a humanoid fecal suspension recognized as “yellow soup” [90, 91]. Dr. Ben Eiseman, Chief of Surgery at Denver General Hospital, along with his team commented on their successful usage of fecal enemas to cure four individuals with pseudo-membranous colitis [92]. Scientists didn’t recognize that the illness they had been treating, pseudomembranous colitis, has been induced by a virulent bacteria called *Clostridioides difficile* until 1978, 20 years later [93].

Individuals can receive fecal microbiome through a number of methods, involving enema [94], nasogastric or nasoduodenal tubes [95] colonoscopy [96], or oral capsules [97, 98]. Furthermore, it is increasingly normal to use refrigerated or freeze-dried feces via anonymously, pre-screened, fit contributors. Despite other antimicrobial treatments, FMT does not produce an underlining dysbiosis which makes the sufferer vulnerable to infections [99]. The processes that enable FMT to prevent infections are still being studied [100], but they most probably include intestinal ecological restorations, intestinal flora composition and functioning, and microbe-host signaling transmission.

Emerging medical research, including animal model research, is increasingly pointing to the value of FMT in inflammatory bowel disease and metabolic syndrome patient treatment. FMT has also been proposed as a viable therapy for various psychiatric diseases, notably autism spectrum disorder. Although experimental animal models have revealed that FMT is effective in preventing energy metabolism disorder and other disorders linked to the gutmicrobiome these results should be carefully considered [101].

Outlook and challenges

As outlined previously, microbiome data offers immense therapeutic potential but remains challenging to analyse and interpret for a variety of reasons including its compositional structure nature that introduces a negative correlation bias, collinearity and sparsity (i.e., multiple types of zeroes value). Currently, the majority of existing microbiome diagnostic approaches suffer from proper validation which result in leakage or overfitting [4].

Lost in translation

Assume that the comparison between a disease and a control group reveals that a microbe is differentially expressed and quantified – how is its clinical relevance then determined? There are no standard cut-offs for effect sizes nor any requirements for reporting 95% confidence intervals and frequently the confidence interval for these differentially abundant features includes zero [33]. But even if these challenges are addressed there remains the issue of determining the directionality of the microbiome diversity impact, as well as the proportion of the disease phenotype that is attributed to the unique microbial features. Moreover, and perhaps more importantly, there needs to be a systematic framework in place that will enable the testing and validation of the efficacy and hypotheses generated through these studies by other laboratories and/or clinical settings.

Applying macro-ecological approaches

Nonetheless, microbiome researchers continue to adapt and develop classic macro-ecological microbiome approaches. Future applications may include developing machine learning models with multi-label classification capacity [102] as well as capable of taking into account volatility and temporal microbiome changes [45, 103]. Given that there is no “standard” or “healthy” microbiome, it remains a challenge to develop replicable diagnostic microbiome pipelines.

This necessitates the need of including more personalized information accompanied by a description of potential factors impacting ecosystems. Many studies now carefully integrate dietary information along with other types of metadata to define the impact of a patient specific microbiome [3, 50, 81, 104–107]. Unfortunately, it isn’t always clear whether the increasing complexity resulting from the application of advanced machine learning approaches, that take into account larger and more diverse datasets, is beneficial and to what degree. Undeniably, in certain contexts, simple linear regression can still outperform more expensive, time-consuming and resource heavy approaches [108].

Fecal microbiota transplantation is now a recognised therapy for *Clostridium Difficile* Infection, Inflammatory Bowel Disease, as well as for several intestinal disorders [94]. Next-generation sequencing technology improvements

have dramatically increased the scope of microbiome variations that may be defined [109], enabling the discovery of dysbiosis biomarkers, as well as the development of increasingly tailored therapy methods, to enhance the effectiveness or accessibility of FMT. Several majors, as well as linked difficulties, should be solved in an attempt to enhance FMT procedures. Firstly, the definition of a “good health” or “perfectly natural” microbe needs to be clearly determined in an attempt to get a deterministic comprehension of host-microbiome interactions. Then the definition of a “good health” or “regular” microbe needs to be clearly stated in sequence to enable more easy detection as well as adjustment of dysbiotic states. Finally, modifications in gastrointestinal microbiology have to be effectively characterized in the literature. Although FMT has clear limitations, it offers the potential of a readily available treatment approach that benefits from reduced toxic effects in comparison to existing synthetic pharmaceutical based approaches.

Outlook

Despite an abundance of recent, novel machine learning approaches and frameworks developed within the context of translational research, very few examples have been demonstrated to successfully achieve and materialise such translations in clinical contexts. Each tool has its own advantages and limitations, which need to be considered when selecting a workflow or algorithm. Moreover, it is vital to assess the performance of the workflow against other standard algorithms and workflows. Nevertheless, we anticipate the future development and use of machine learning models as invaluable diagnostic or pre-screening tools.

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